

In Cooperation with the University of Arizona, School of Natural Resources

# Vascular Plant and Vertebrate Inventory of Coronado National Memorial



Open-File Report 2007-1393

U.S. Department of the Interior  
U.S. Geological Survey  
National Park Service

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## **Vascular Plant and Vertebrate Inventory of Coronado National Memorial**

Edited by Cecilia A. Schmidt , Brian F. Powell, Don E. Swann, and William L. Halvorson

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**U.S. Department of the Interior  
U.S. Geological Survey  
National Park Service**

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## Executive Summary

We conducted inventories for amphibians and reptiles, birds, and mammals; and summarized past inventories for vascular plants at Coronado National Memorial (NM) in Arizona. We used our data as well as data from previous research to compile species lists for the memorial, assess inventory completeness, and make suggestions on future monitoring efforts.

There have been 940 species of plants and vertebrates recorded at Coronado NM (Table 1), of which 46 (5%) are non-native. The species richness of the memorial is one of the highest in the Sonoran Desert Network of park units, third only to park units that are two and one-half (Chiricahua National Monument), 19 (Saguaro National Park) and 70 (Organ Pipe Cactus National Monument) times larger in area. The high species diversities are due to the large elevational gradient, overlap of biogeographical

regions, wide range of geology and soils, and diverse vegetation communities present at the memorial.

Changes in species composition have occurred at the memorial over the last 20 years in all major taxonomic groups. These changes are likely due to increases in grassy plant species (both native and non-native) at the lower elevations of the memorial. We suspect that grassy plant cover has increased because of changes in grazing intensity, introduction of some non-native species, and a recent fire. All recent vertebrate inventories have yielded grassland obligate species not previously recorded at the memorial.

Based on the review of past studies, we believe the inventory for most taxa, except bats, is nearly complete, though some rare or elusive species will likely be added with additional survey effort.

**Table 1. Summary of vascular plant and vertebrate inventories at Coronado NM.**

Taxonomic group	Number of species recorded	Number of non-native species
Plants	649	41
Amphibians and Reptiles	43	0
Birds	196	2
Mammals	52	3
<b>Totals</b>	<b>940</b>	<b>46</b>





# Chapter 1: Introduction to Biological Inventories At Coronado National Memorial

Brian F. Powell, Cecilia A. Schmidt, and William L. Halvorson

## Project Overview

*Inventory: A point-in-time effort to document the resources present in an area.*

In the early 1990s, responding to criticism that it lacked basic knowledge of natural resources within park units, the National Park Service (NPS) initiated the Inventory and Monitoring Program (NPS 1992). The purpose of the program is to increase scientific research in NPS units and to detect long-term changes in biological resources (NPS 1992). At the time of the program's inception, basic biological information, including lists of plants and animals, were absent or incomplete for most park units. In fact, as of 1994, more than 80% of national park units did not have complete inventories of major taxonomic groups (Stohlgren et al. 1995).

Species inventories have both direct and indirect value for management of natural areas. Species lists facilitate resource interpretation and visitor appreciation of natural resources. Knowledge of which species are present, particularly sensitive species, and where they occur is critical for making management decisions (e.g., locating new facilities). Inventories are also the cornerstone for long-term monitoring. Thorough biological inventories provide a basis for choosing parameters to monitor and can provide initial data (i.e., a baseline) for monitoring ecological populations and communities. Inventories can also assist in testing sampling strategies, field methods, and data collection protocols, and can provide estimates of variation that are essential in prospective power analysis.

## Report Format

This report is intended to be useful for internal planning, outreach, and education. We report only

common names in the text unless we reference a species that is not listed later in an appendix; in this case we present both common and scientific names. For each taxonomic group we include an appendix of all species that have been recorded in the memorial (Appendices A–D), and species whose presence is possible (except for plants and birds; Appendices E and F). Species lists are in phylogenetic sequence and include taxonomic order, family, genus, species, subspecies or variety (if applicable), and common name. Units of measurement are presented in accordance with the International System of Units.

## Species Conservation Designations

We indicate species conservation designations by the following agencies: U.S. Fish and Wildlife Service (responsible for administering the Endangered Species Act), Bureau of Land Management, USDA Forest Service, Arizona Game and Fish Department, and Partners in Flight (a partnership of federal, state and local governments, non-governmental organizations, and private industry).

## Voucher Specimens

Voucher specimens are an indisputable form of evidence of a species occurrence. For plants, we electronically searched the Herbaria at Arizona State University, Desert Botanical Gardens, Northern Arizona University, and the University of Arizona for existing specimens from Coronado NM (see Appendix A for results). We searched for existing vertebrate vouchers in records from 28 natural history museums (Table 1.1; see Appendix H for results).

**Table 1.1. Museums that were queried, in 1998, for vertebrate voucher specimens with “Arizona” and “Coronado National Memorial” in the collection location. Collections in bold-faced type had specimens from the memorial.**

<b>Collection</b>	<b>Collection cont.</b>
<b>Brigham Young University</b>	Oklahoma Museum of Natural History, Norman
<b>Chicago Academy of Sciences</b>	Peabody Museum, Yale University
Cincinnati Museum of Natural History & Science	Saguaro National Park
Cornell Vertebrate Collections, Cornell University	Strecker Museum, Baylor University, Waco
George Mason University (Fairfax, VA)	Texas Cooperative Wildlife Collection
Marjorie Barrick Museum, University of Nevada-Las Vegas	<b>University of Arizona</b>
Michigan State University Museum (East Lansing)	<b>University of Colorado Museum</b>
Milwaukee Public Museum	<b>University of Illinois, Champaign-Urbana</b>
<b>Museum of Comparative Zoology, Harvard University</b>	<b>University of Kansas, Natural History Museum</b>
Museum of Texas Tech University	<b>University of Michigan</b>
Museum of Vertebrate Zoology, University of California, Berkeley	University of Texas, Arlington
Museum of Life Sciences, Louisiana State University, Shreveport	Walnut Canyon National Monument, Arizona
<b>Natural History Museum of Los Angeles County</b>	Western Archaeological and Conservation Center, Tucson
North Carolina State Museum of Natural Sciences	Wupatki National Memorial, Flagstaff

## Chapter 2: Memorial Overview

Cecilia A. Schmidt, Brian F. Powell, and William L. Halvorson

### Memorial Area and History

Coronado National Memorial is located in southeastern Arizona approximately 30 km south of the city of Sierra Vista (Figs 2.1, 2.2). The memorial is bounded by the U.S./Mexico border to the south, private and Arizona State Trust land to the east, and USDA Forest Service land to the west and north (NPS 1998). Nearby Sierra Vista, one of the fastest growing cities in Arizona, has an estimated population of 40,000 (NPS 1998). Other small towns, such as Hereford and Palominas on the U.S. side of the border, and Ejido Jose Maria Morales on the Mexican side of the border, are within 20 km of the memorial.

Coronado NM was established in 1941 as an international memorial, then changed status in 1952 to a national memorial. The memorial was created to commemorate the 15th century explorations of Francisco Vazquez de Coronado, the first Spanish explorer of southern Arizona (NPS 1998). When originally founded, the memorial encompassed approximately 1,165 ha. In 1978, the memorial was expanded by 744 ha to include the entire Montezuma Canyon Watershed (Fig 2.2; NPS 1998, NPS 2004a). Currently, the memorial encompasses 1,922 ha (NPS 1998). Annual visitation to the memorial averages 90,000 (NPS 2004b).

### Natural Resources Overview

#### *Physiography, Geology, and Soils*

Within the Basin and Range Physiographic Province, the memorial is located at the southern end of the Huachuca Mountains, one of the region's "sky island" mountain ranges. Topography in most parts of the memorial is steep, climbing from 1,400 m in the grassland plain of the memorial's southeast corner, to 2,350 m at Montezuma Peak, located along the memorial's northern border.

The geology of the area is complex, with high-angle reverse faults juxtaposing sedimentary, igneous, and metamorphic rock. At least nine caves, most notably Coronado Cave, have been discovered at the memorial (NPS 1998). The

memorial is dominated by rock outcrop and alluvium. Deep soils (up to 1.5 m) predominate on the lower slopes and are especially notable in the southeastern portion of the memorial. However, shallow soil associations (from 25–50 cm) with high rock-fragment content are typical of the steeper slopes and are the most extensive (Richardson et al. 1979). For a complete soil survey, see Denny and Peacock (1996).

#### *Hydrology*

The deep soils in alluvium typically have low permeability and widely varying water-holding capacity, while the shallow soil associations, with their high rock fragment content, have low water-holding capacity (Richardson et al. 1979). Intense precipitation generates flash floods and it is not uncommon for especially heavy storms to reshape drainages. The main drainage, the east-southeast/west-northwest trending Montezuma Canyon, bisects the memorial. Main drainages in the mountainous terrain channel the flow to Montezuma Canyon, while the numerous drainages within the grasslands flow to the San Pedro River (east of the memorial). Springs present at the memorial, such as Yaqui Springs, trickle rather than flow, and permanent surface waters are not present.

#### *Climate*

Coronado NM experiences an annual bimodal pattern of precipitation which is characterized by heavy summer (monsoon) storms brought about by moisture coming from the Gulf of Mexico and less intense frontal systems coming from the Pacific Ocean in the winter. On average, approximately one-half of the annual precipitation falls from July through September (Table 2.1; WRCC 2004). The area's hot season occurs from April through October; maximum temperatures in July can exceed 35°C. Winter temperatures dip below freezing and snow is occasional.

#### *Vegetation*

According to NPS (1998) and Ruffner and Johnson (1991), the memorial contains four vegetation

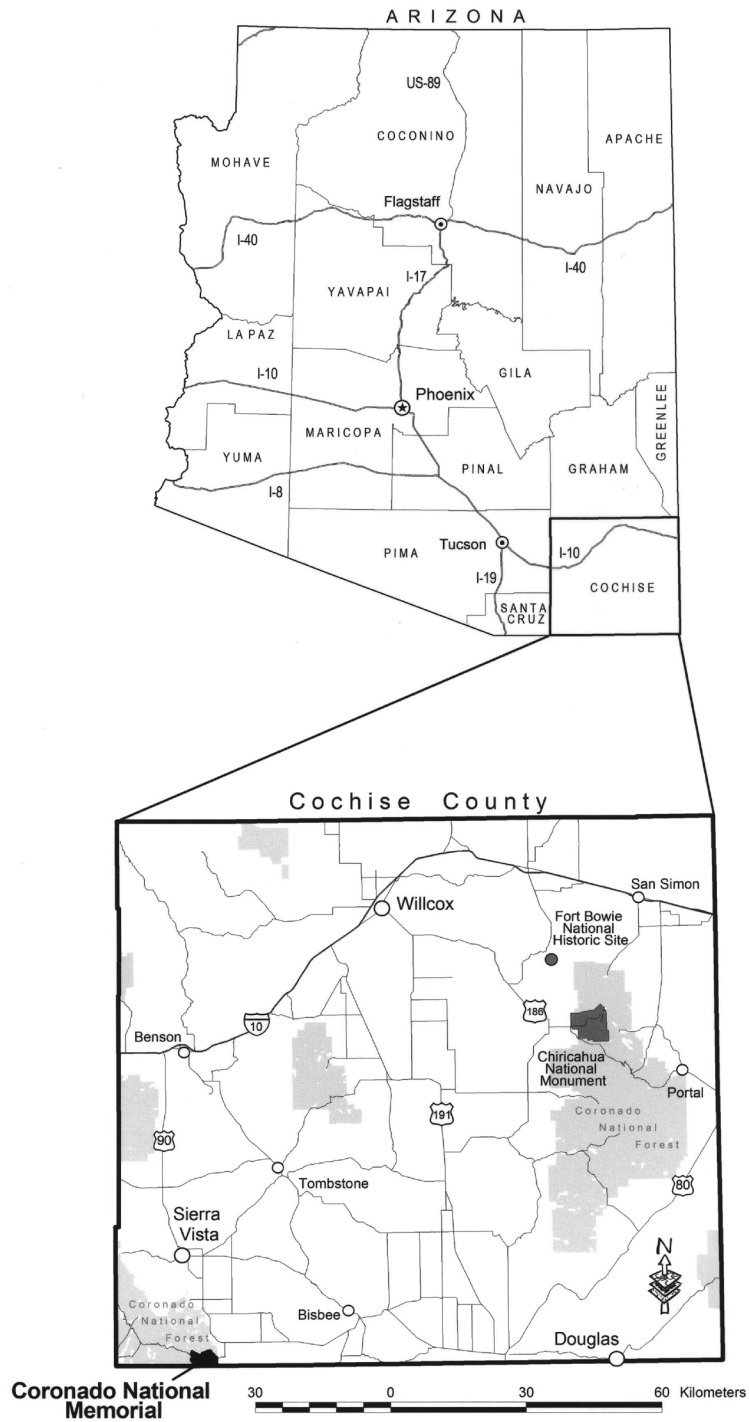
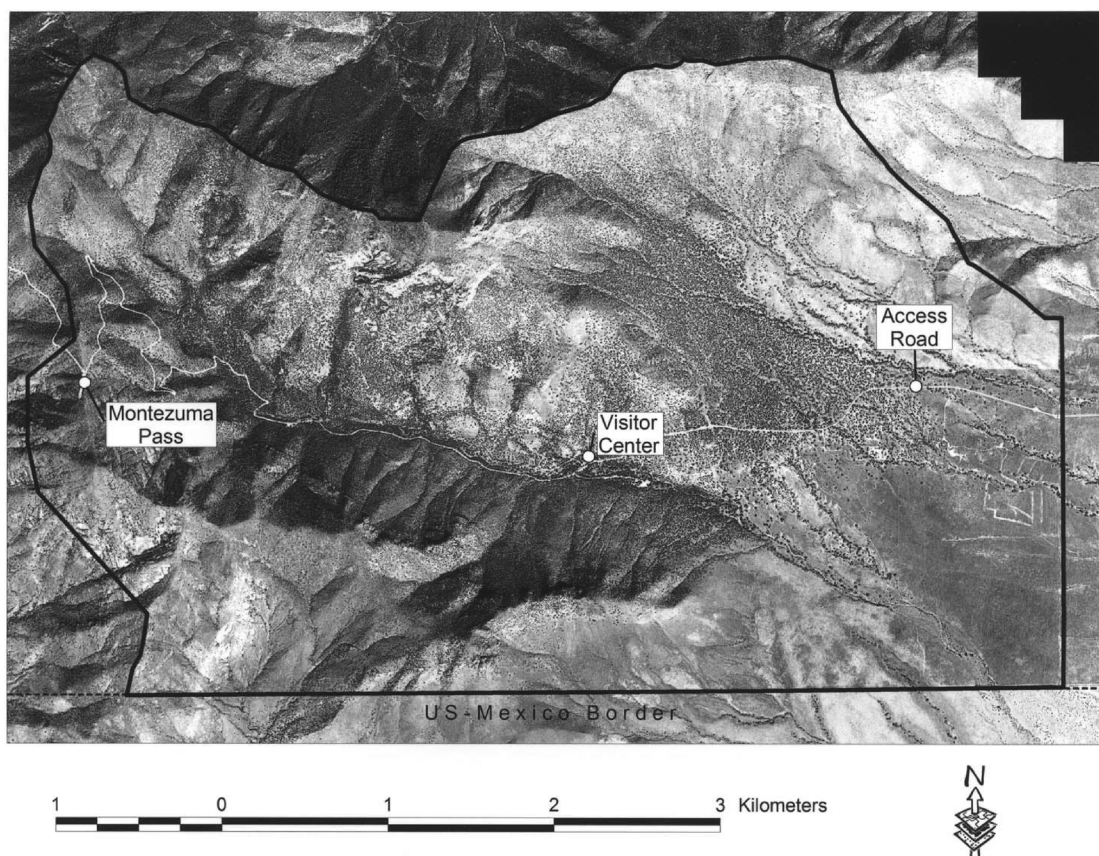


Figure 2.1. Location of Coronado National Memorial, Arizona.





**Figure 2.2. Boundary and major features, Coronado NM.**

types: Oak-Mexican Pinyon Pine-Juniper Woodland (evergreen woodland community), the most extensive community, occurs mainly on the north and southwest facing slopes of the memorial; Grama Grass-Mixed Grass-Mixed Shrub Grassland (Chihuahuan semi-desert grassland community) occurs on the easternmost part of the memorial; Velvet Mesquite-Mixed Short Tree Woodland (Desert Riparian Forest community) is found in the easternmost part of the memorial along low elevation ephemeral drainages; and Arizona Sycamore-Arizona Walnut-Oak Riparian Forest (mixed broadleaf forest community) is found along Montezuma Canyon.

## Natural Resource Management Issues

### *Border Crossings*

The most pressing management issue for Coronado NM is the trespass of drug smugglers and undocumented immigrants (border crossers) across the U.S./Mexico border. It is estimated that 150 undocumented immigrants enter the U.S. through the memorial each day (NPS 2003a) and approximately 75,000 to 100,000 pounds of drugs are brought through the memorial each year (NPS 2003b).

In addition to the obvious safety concerns, border crossers impact the natural resources of the memorial. Border crossers have created a network

**Table 2.1. Average monthly climate data for Coronado NM, 1960–2004.** Data from WRCC (2004).

Characteristic	Month												Annual
	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec	
Maximum temperature (°C)	14.7	16.7	19.6	23.8	28.2	33.0	32.2	30.6	29.2	24.8	19.1	14.8	23.9
Minimum temperature (°C)	0.3	1.2	2.9	6.2	10.4	15.0	16.7	15.8	13.9	9.2	3.8	0.5	8.0
Precipitation (cm)	4.3	3.7	2.9	1.1	0.6	1.4	11.5	10.2	5.0	3.9	2.5	5.3	4.3



**Figure 2.3. Trails and roads made by border crossers into Coronado NM.** Photo by Mike Hardin.

of trails and roads in the memorial, particularly in the semi-desert grasslands on the southeast side of the memorial (see Fig. 2.3). They have also damaged vegetation, eroded and compacted soil, and left large amounts of trash behind (Hubbard 2002, Hubbard et al. 2003, NPS 2003a). These impacts may affect water quality and wildlife movement patterns, though the extent of these impacts has not been established. Border crossers often shelter in caves and old mine shafts that provide roosting habitat for bats, including the endangered lesser long-nosed bat, which has been documented at the memorial (Hubbard 2002, Hubbard et al. 2003, NPS 2003a). Border crossers light fires (using trees or wood from historic structures on the memorial), some of which may burn out of control and spread through the memorial (NPS 2003a, Hubbard et al. 2003). In response to growing threats of undocumented immigrants and drug smugglers to memorial resources, the NPS built a vehicle barrier through the southeastern portion of the memorial, however the remainder of the memorial boundary with Mexico is lined with only barbed-wire fence (NPS 2003b).

### *Fire*

Before the 1900s, fire occurred regularly in the area, but since the early 1900s fires have been actively suppressed on the memorial (Ruffner and

Johnson 1991). Fire suppression leads to an increase in woody plant species, eventually increasing the intensity of fires, such as the Peak Fire of 1988, which burned most of the memorial (Ruffner and Johnson 1991). The fire varied from moderate to severe in intensity, in many places killing trees; it also appears to have led to an increase in grasses (Ruffner and Johnson 1991). Fires can also cause air quality and soil erosion problems and result in the loss or alteration of habitat for wildlife. Fire management is made difficult because of the memorial's proximity to Mexican ejido (communal) lands, where there is no fire management plan and few resources for fighting fires. Coronado NM is part of the Huachuca Fire Partners group, which works with local, state, and federal agencies, as well as private landowners, to manage fires on a landscape scale, regardless of political boundaries (Barbara Alberti, pers. com.).

### *Non-Native Species*

The most pressing non-native species issue at the memorial is the spread of Lehmann lovegrass and other non-native grasses. Lehmann lovegrass has become the dominant grass on the southeast side of the memorial where it has replaced many native grasses. This change in species composition can alter the fire regime of the area by supporting higher fire frequencies, thereby leading to other

changes in vegetation composition and structure (Anable et al. 1992). However, abundance and species richness of some taxa, such as small mammals, may be higher in areas with Lehmann's lovegrass.

### *Grazing*

Livestock grazing has degraded an estimated 80% of streams and riparian ecosystems in the western United States through loss of vegetation, stream-bank erosion, soil compaction, flooding, and water pollution (BLM 1994). No single land-use activity has affected western riparian systems as much as cattle grazing (Bahre 1991, Fleischner 1994). Livestock grazing can also increase the number and extent of non-native plants (Belsky et al. 1999) and can negatively affect wildlife through habitat modification and competition for resources (Saab et al. 1995).

Cattle have been grazed on the memorial since the 16th century. At the time of the memorial's establishment, it had four grazing allotments managed by the USDA Forest Service (NPS 1998). In 1992, two of those allotments were retired and the management of the remaining allotments was turned over to the NPS (NPS 1998). The memorial still manages two cattle allotments, consisting of approximately 1,390 ha. or 37% of the memorial, which the staff would like to retire (NPS 1998, NPS 2004a). In the surrounding areas, especially on the Mexico side of the boundary, cattle are still being grazed today (See Fig. 2.2) and trespass of cattle from Mexico onto the memorial is common (Hubbard 2002). At the memorial, cattle spend more time in the Montezuma Wash because of an abundance of shade and palatable forage there (NPS 2004a), which is consistent with their habitat-use patterns (Kauffman and Krueger 1984).

### *Development of Adjacent Lands*

Housing developments east of the memorial are beginning to encroach on the memorial's boundary (NPS 1998, Hubbard et al. 2003). Potential impacts of development include an increase in the number and extent of non-native plants, disruption of animal movement patterns, and increased

harassment and mortality of native animals by pets and feral animals (NPS 1998, Tigas et al. 2002, Hubbard et al. 2003). Development will also likely increase groundwater withdrawal (Hubbard et al. 2003). Because of the small size of the memorial, the memorial itself can not support the home ranges of many large mammals, such as bears, mountain lions, and jaguars. In order for these species to survive, areas surrounding the memorial must also be protected from development.

### *Caves and Abandoned Mines*

The memorial is home to 62 abandoned mines and several caves (NPS 1998). Mining for lead and zinc began on the memorial in the late 1800s and continued through 1947, thereby leaving contaminated tailings that are polluting water at the memorial (NPS 1998). These mines can pose safety threats to visitors but also provide habitat for many species of wildlife, especially bats.

### *Aircraft Noise*

Low-flying military aircraft from Fort Huachuca, law enforcement aircraft from the U.S. Border Patrol, and private aircraft pass over the memorial often (NPS 1998, Hubbard et al. 2003). Both vibrations and noise generated by these aircraft affect the natural quiet of the memorial and may also affect wildlife in the area (NPS 1994). Aircraft overflights can produce changes in the physiology and behavior of some wildlife species (Luz and Smith 1976, Craig and Craig 1984, Ellis and Ellis 1991, Weisenberger et al. 1996).

### *Wildlife Poaching*

There have been some incidents of wildlife poaching at the memorial in the recent past and there may still be some poaching occurring. At least one arrest was made in a reptile poaching incident during the past decade (Barbara Alberti, pers. comm.). Swann et al. (1999) documented evidence of illegal collecting of snakes, although they did not feel that this was a major management concern at the time. The presence of certain highly-prized species has the potential to make illegal collecting a management issue in the future.



## Chapter 3: Plant Inventory

Cecilia A. Schmidt

There have been several studies on the vegetation at Coronado National Memorial. The memorial has kept a list of all plant species identified within its boundaries. Ruffner and Johnson (1991) mapped the vegetation of the memorial, as well as created a plant list based on the memorial's vegetation list and species found on their monitoring plots. Parfitt and Christy (1992) also created a species list based on the memorial's list and specimens previously accessioned into herbaria at Arizona State University and the University of Arizona. Ruyle (2002) established vegetation monitoring plots on one of the grazing allotments (Joe's Spring Allotment) and monitored them from 1998 to 2001. Halvorson and Guertin (2003) mapped common non-native species found at 11 national park units, including Coronado NM. The NPS Sonoran Desert Network (SDN) Inventory and Monitoring Program (unpublished data) conducted vegetation sampling on 33 plots throughout the memorial in 2002. Finally, vegetation sampling at diurnal breeding-season bird stations was conducted in 2004 (see Chapter 5).

### Data Used to Create Plant Species List

The plant species list for the memorial (Appendix A) is based on Ruffner and Johnson (1991), Parfitt and Christy (1992), Ruyle (2002), Halvorson and Guertin (2003), the NPS SDN Inventory and Monitoring Program data (unpublished), and vegetation sampling data from breeding-season bird stations. Taken together, these sources provide a comprehensive list of plants occurring at the memorial.

### Nomenclature

Scientific and common names used in this report are current according to the Integrated Taxonomic Information System (ITIS 2004) and the PLANTS database (USDA 2004).

### Results and Discussion

There have been 649 plant species documented at Coronado NM (Appendix A), 41 (6%) of which are

non-native. Ruyle (2002) lists horehound (*Ballota* species) as being found on one of his plots in the memorial. However, this species is not found in the southwestern United States and therefore we omitted it from the species list (Appendix A). Coronado NM, surprisingly, has one of the lowest percentages of non-native plant species in the Sonoran Desert Network of park units, second only to nearby Fort Bowie National Historic Site. This is surprising because of the history of disturbance to the memorial, most notably cattle grazing that continues to occur today. Although Coronado NM does boast one of the lowest number of non-native plant species, the spatial coverage of a few species is wide-ranging. Lehmann lovegrass, in particular, covers most of the eastern portion of the memorial and was found to have the highest frequency and coverage on several of Ruffner and Johnson's (1991) monitoring plots. It was also found on almost every one of Ruyle's (2002) monitoring plots and often in high frequencies. Halvorson and Guertin (2003) also found this species to be widespread at the memorial. Ruffner and Johnson (1991) found that native species that are more tolerant to trampling (by cattle and humans), such as blue grama, may be more common under the current conditions than were historically present. These species may have replaced other grasses such as big sacaton, historically common in southern Arizona, and now rare at the memorial.

Two species of concern under the Endangered Species Act, the Huachuca Mountain milkvetch and netted globecherry, have also been documented at the memorial (Ruffner and Johnson 1991, HDMS 2003). Ruffner and Johnson (1991) mention that woody species, such as mesquite, may be increasing in the grassland areas of the park. Encroachment of woody vegetation into grasslands, with the subsequent loss of these grasslands and the species that depend on them, is a major natural resource issue in southern Arizona (NPS 2000, Taylor 2004). Ruffner and Johnson (1991) suggest that periodic fires, which kill off woody species' seedlings and saplings, are important to reducing encroachment and therefore restoring the semi-desert grasslands.

### *Peak Fire*

The Peak Fire occurred during Ruffner and Johnson's (1991) study. This allowed them the unique opportunity to observe the effects of the fire and the subsequent flash flood events on vegetation at the memorial. Overall, they found that the fire had little effect on the long-term species composition and densities (Ruffner and Johnson 1991). Only individuals from two species, rabbitbrush (*Ericameria* species) and hedgehog cactus (*Echinocereus* species), were killed by the fire and had not returned within the year (Ruffner and Johnson 1991). However, there were short-term changes: species whose germination is encouraged by fire or disturbance such as morning glory (*Ipomoea* species), Sonoita nightshade, and Orcutt's threeawn, increased quickly after the fire, but a year after the fire were found in low numbers or not at all (Ruffner and Johnson 1991). Short-term effects on trees were minimal; within a year many had sprouted back (Ruffner and Johnson 1991).

Subsequent flash flood events after the Peak Fire altered wash and stream characteristics and washed away many individual plants.

### **Inventory Completeness**

With each additional study conducted at the memorial the number of species identified has increased. Ruyle (2002) added 16 species to the lists by Ruffner and Johnson (1991) and Parfitt and Christy (1992). The Inventory and Monitoring (NPS, unpublished data) study conducted only one year after Ruyle's study added an additional 13 species. Based on the number of studies and collections, we believe the plant inventory is near 90% complete. However, new species will continue to be found and increasing development around the memorial may result in more non-native species being established at the memorial in the future.

## Chapter 4: Reptiles and Amphibians Inventory

Don E. Swann and Cecil R. Schwalbe

Although specimens of amphibians and reptiles have been collected in Montezuma Canyon sporadically throughout the latter half of the twentieth century, the first formal survey of herpetofauna at the memorial was by University of Arizona researchers during 1978. Based on generalized searches, Johnson and Lowe (1979) confirmed 16 species of reptiles and six species of amphibians, and believed that at least another 13 species of reptiles and one species of amphibian occurred on site. However, the Lowe-Johnson study was of short duration and the boundaries of the memorial have changed significantly since 1978. Other studies of herpetofauna at Coronado NM have included barking frogs (Goldberg and Schwalbe 2000) and montane rattlesnakes (Swann et al. 1999).

The main purpose of the present study was to conduct field surveys for reptiles and amphibians at Coronado NM and to combine this information with historic data to produce a baseline inventory. In addition, we proposed to develop a repeatable study design for monitoring species diversity of herpetofauna. We used a variety of techniques to confirm as many of the memorial's reptiles and amphibians as possible. We recorded all individual reptiles and amphibians observed during 1997–1998 and determined the approximate location of each observation; we also continued to collect species presence data opportunistically through 2001. Specific objectives were:

1. Survey reptiles and amphibians using techniques that have minimal impact on the animals and the environment;
2. Take voucher photographs and collect voucher specimens to document which species are present on the memorial;
3. Gather historic information on herpetofauna at Coronado NM from published sources, museum collections, and historic reports;
4. Document abundance of selected species in different vegetation communities at the memorial;
5. Provide information for the basis of long-term monitoring; and
6. Provide this information to the memorial in a format useful for making management decisions and interpretation.

Several products are provided in the appendices of this report. Additionally, a separate report (Swann and Schwalbe 2002) provides detailed species accounts and distribution maps for all species. Printed and electronic copies of data sheets, data tables, Global Positioning Systems (GPS) files, field notes, and voucher photos and other photos, have been sent separately to the memorial.

### Methods

Fieldwork on this study began in April 1997 and was completed in October 1998. However, observations on reptiles were also made during a study of mammals at the memorial starting in October 1996 (Chapter 6). Additionally, this report includes specimens collected by park staff and the authors through May 2001. Because reptiles and amphibians are very diverse in their habits and habitats, we used many different methods to detect species. Methods included visual encounter surveys on large and small plots; visual surveys on transects established by Johnson and Lowe (1979); road transects (“road cruising”) with an automobile; historic and museum records; and sightings by staff and visitors. Due to the sensitivity of cultural resources on the memorial, which required archaeological approval, and then illness of the major field investigator (DES) during a period in the summer of 1998, we only established one pitfall array, which had very low trap success. We employed extensive cover-turning, particularly turning of debris at the Montezuma Ranch, to observe and capture reptiles and amphibians. However, we were careful to return all cover to its natural place and appearance (most of this cover was removed during a cleanup of Montezuma Ranch at the end of the field season in 1998).

As an extension of this study, we established a study area to search for and estimate abundance of banded rock rattlesnakes and Sonoran mountain kingsnakes, and did focused searches for two other species of montane rattlesnakes — twin-spotted rattlesnake and ridgenose rattlesnake. Results of these searches are reported here, but significantly greater detail is available in a separate report, “Ecology of Banded Rock Rattlesnakes and Sonoran Mountain Kingsnakes at Coronado National Memorial,” (Swann et al. 1999) funded by and submitted to Southwest Parks and Monuments Association (SPMA).

### *Visual Encounter Surveys*

The visual encounter survey (Crump and Scott 1994) is a standard method for inventory and monitoring of reptiles and amphibians that are difficult to capture in traps. The method utilizes systematic, timed searches in a defined habitat or area. During visual encounter surveys a variety of specific techniques are used to detect animals on the ground surface, above the ground in trees and other vegetation, within rock cracks and other geological features, and below ground under inorganic and organic debris (Table 4.1).

In 1997, we systematically and repeatedly searched the entire land area of the memorial using visual encounter surveys within broad (5–10 ha) areas based on different vegetation communities. We made an effort to alternate searches in different vegetation communities so that on every field trip we spent time searching in each community. In 1998, we continued these searches, but also

established smaller plots for monitoring purposes (see monitoring plots in Figure 4.1).

Visual encounter surveys were generally conducted during morning and early evening hours. During each survey, we recorded the start and end time, the researcher, the area searched, and the percentage of time spent on each type of search activity. For each individual amphibian, reptile, and mammal observed, we recorded the date and time of the observation, the location, the method used, and habitat characteristics such as vegetation community and substrate.

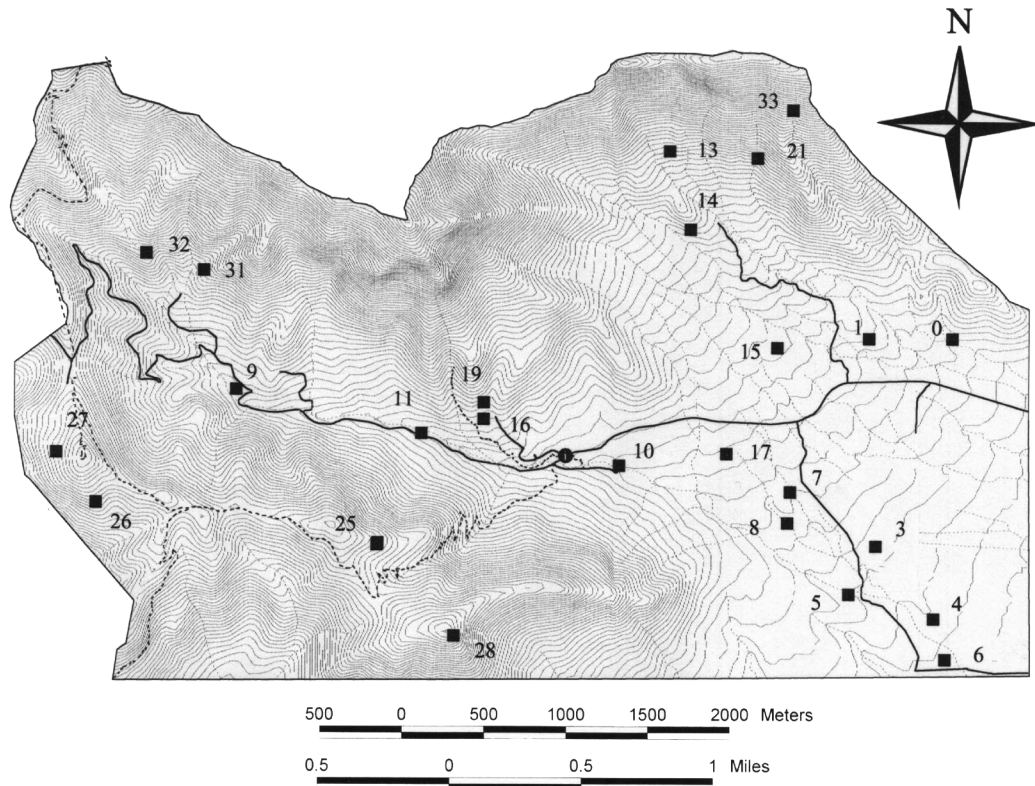
### *Monitoring Plots*

The monitoring phase of this study was conducted during April–October 1998. Coronado NM was divided into six sections based on vegetation communities and elevation, and 25 plots were randomly located within each section (Figure 4.1). Ten plots were in oak woodland, five below 1,800 m and five above this elevation. Four plots were in semi-desert grassland, two below 1,500 m and two above this elevation. Nine plots were located within Montezuma Canyon and its tributaries: two in mesquite riparian, two in oak riparian below 1,500 m, three in oak riparian between 1,500 m and 1,800 m, and two in oak riparian above 1,800 m. Two plots were located at perennial seeps (the “blue waterfall” near the residence area and the “fern grotto” below Bob Thompson Peak). All plots were 0.5 ha except for the four grassland plots, which were increased in size to 1 ha due to very low numbers of observations. Grassland plots were 100 m by 100 m, while other plots were rectangles with sides of 50 m and 100 m. All plots

**Table 4.1. Methods used during visual encounter surveys at Coronado NM, 1997–1998.**

Method	Description
Visual search on ground	Observer walks slowly while observing ground or rock surfaces, using eyes and ears to observe surface-active animals
Visual search in trees	Observer inspects tree trunk, branches, leaves, and bark to observe tree-dwelling animals
Binocular search	Observer uses binoculars to scan rock surfaces, exposed tree trunks and branches, etc., to observe basking animals
Search in rock cracks	Observer uses mirror to reflect sunlight or shines flashlight into rock cracks, caliche caves, burrows, etc., to observe crack-dwelling animals
Cover-turning	Observer turns over (and carefully replaces) rocks, logs, leaf litter, etc., to observe animals hidden underneath
Search in water	Observer uses visual search, or dip-nets, to find aquatic adults, larvae, and eggs
Listen for calls	Observer listens for calling amphibians, usually during summer rain events





**Figure 4.1. Coronado NM, indicating locations of monitoring plots established during this study .**

were aligned with the major axis east/west (except for riparian plots, which were aligned to follow the major stream channel) and the southeast corner was located using a Trimble GeoExplorer GPS unit. Each plot was sampled a minimum of eight times during the study, four times during spring (May-early July), and four times during summer (late July-September). Plots were sampled for 30 minutes each time. To evaluate the effects of temporal variability within morning surveys, plots were usually sampled two times each per sampling day. To evaluate observer effect, different observers usually conducted surveys on any given day. Thus on a typical morning, two observers would sample four plots for 30 minutes each, and each plot would be sampled two times, once by each observer.

To better understand temporal variability within plots, we selected a subset of four plots (one each in woodland, oak riparian, mesquite riparian, and seep) for repeated samples. These plots were sampled 12–16 times throughout the study.

During each 30-minute survey, the entire plot was systematically searched. The major goals

of each survey were to achieve complete (geographic) coverage of the plot, to observe as many species as possible (without guessing at the identity of species), and to observe as many individuals of each species as possible. All observers were herpetologists with >2 years experience in finding reptiles and amphibians in Arizona under different weather conditions, in different habitats, and using different techniques. Each observer was asked to use his experience to choose the appropriate survey technique for the conditions on the plot.

During each survey we recorded a number of variables, including shaded temperature at 1.5 m, 0.5 cm, and ground surface at the start of the sampling period. We did not attempt to sample reptiles if air temperatures were below 25°C. For each reptile and amphibian observed (and for most mammals), we recorded the time, technique used, vegetation community, and substrate. We also made notes on substrate, and located the individual on a hand-drawn map of the plot.

### *Historic Transect*

To compare potential changes in relative abundance of herpetofauna at Coronado NM between the study by Johnson and Lowe (1979) and our study, we relocated five, 1-km transects established by Johnson and Lowe in 1978 and resurveyed them. During each survey we recorded, as they did, all individuals of each species encountered. Unfortunately, it is not possible to determine from Johnson and Lowe's report when or how often they surveyed the transects, or which data used in relative abundance estimates were from the transect and which were obtained from road riding and "additional careful search of the habitats on site and directly off site." Their original data appears to be lost and the field notes provided by Terry Johnson (pers. comm.) were incomplete. To provide the best coverage possible, we sampled each transect at least twice during each spring and summer season of our two-year study. We also recorded the time, approximate location, and habitat characteristics of each observation using the same methods as on our plots.

### *Road Transects*

We searched for reptiles at all times while driving on East Montezuma Canyon road within the memorial boundary. These included trips (particularly at night) specifically to search for amphibians and reptiles, as well as any time we drove on the roads, such as travel to and from monitoring grids. During each trip we recorded the start and end times and start and end locations. If the trip ended within the memorial and did not involve any turns or returns (for example, if we drove from Montezuma Ranch to the visitor center), we simply recorded the start and end information for that "segment". If the trip included a return (for example, if we drove from the visitor center to Montezuma Pass and returned), we ended the segment at the turn-around point, then started a new segment for the return trip. If the trip took us outside the memorial boundary, we began and ended segments at the memorial boundary (we did search outside the boundary for specimens, but do not include these data in the road transect results). For each amphibian, reptile, and mammal observed on road segments, we recorded the time, odometer location, and species identification. Odometer

locations were translated to GPS coordinates in the database for species mapping.

### *Historic Records*

We obtained historic records of reptiles and amphibians at Coronado NM from a variety of sources, including Johnson and Lowe (1979), museum specimens, and historic records located at the memorial. Additionally, we obtained museum records from natural history collections throughout the United States, including 17 university museums, the American Museum of Natural History in New York City, and the Smithsonian Institute in Washington, D.C. A large number of records were obtained from the University of Arizona herpetological collection (UAZ). We compiled historic records in a database with all available information for each record, such as collecting date, collector, and other data on the specimen tag itself.

### *Additional Methods*

During this study we recorded all individuals of all amphibians and reptiles observed, including individuals observed when we were in transit between plots or occupied in other activities. Throughout most of the study we recorded all observations of mammals (summarized in Chapter 6). We also collected reports of sightings, photographs, and road-killed specimens of amphibians and reptiles from staff and visitors. For uncommon species, we evaluated the reliability of different observations for species accounts. We did not record all individual amphibians and reptiles observed off-site, but collected data on individuals of species that were uncommon or were of unknown status within the boundaries of the memorial; we sometimes collected voucher specimens from off-site.

### *Analyses*

All observations of reptiles and amphibians made during the study were entered in a Microsoft Access database. Separate tables were created for both effort and results of visual encounter surveys, historic transects, road transects, monitoring plots, and incidental sightings. For each observation not on a mapped plot, we determined a general (+ 200 m) location using a grid overlay on a large detailed

topographic map of Coronado NM (undated NPS map, datum NAD 27 CONUS). Observations were assigned to points to generate distribution maps for each species in the GIS software program ArcView 3.2.

Summary statistics for monitoring plots were generated using the software programs Microsoft Excel and JMP-IN (SAS Institute, Inc.). Temporal and observer variability were evaluated using Analysis of Variance (ANOVA; Zar 1996). Results were considered significant if  $P < 0.05$ . Species richness (the number of species present at the site) was estimated using the mark-recapture methods of Program CAPTURE for closed populations (Otis et al. 1978). This program can estimate species richness based on patterns of observations in the field; that is, whether each species is either observed (“captured”) or not observed (“not captured”) during successive field-days (Swann 1999). In our analysis, species richness was estimated based on seven periods of 20 sequential field person-days.

### Species List

Development of a species list of vertebrates for any park or land area is complicated because some species are transitory and the occasional observation of an individual of a particular species does not necessarily indicate the species is resident on site. In addition, some species look alike and can be confused with one another. Even trained herpetologists have difficulty distinguishing among species of whiptail lizards (*Cnemidophorus* spp.) without detailed observations and measurements

that can only be made on captured individuals or preserved specimens. During this study, a species was considered “Confirmed Present” at the memorial only if a voucher specimen or unambiguous voucher photograph was obtained during this study or had been previously obtained by Johnson and Lowe (1979).

In general we collected only species that had not been collected in previous studies or that we found dead (usually on roads). We used voucher photographs whenever possible. Records of all voucher specimens can be found in Appendix H. All specimens have been deposited in the University of Arizona herpetology collection.

### Nomenclature

Due to rapidly improving genetic technology and other tools, taxonomists are continuously updating the classification of amphibians and reptiles. Unfortunately, this activity often leads to changes of long-familiar names, and in recent years the rate of name changes has exceeded the ability of field guide authors to keep pace. In this paper, we use the standard names of Collins (1997), which are different in many cases from the names used in the current Peterson field guide (Stebbins 1985) and the most recent list published by the Society for the Study of Amphibians and Reptiles (Crother 2000).

## Results

### Overall

We made 396 individual observations of six species of amphibians (not including larvae; Table

**Table 4.2. Species of amphibians confirmed at Coronado NM, and number of observations of adults during 1997–1998 on general visual encounter surveys (VES), monitoring plots (Plots), road transects (Roads), the Lowe-Johnson transect (Transect), and incidental observations (Other). HR refers to historic records: C = confirmed by specimen during previous study; SO = historic sight observation.**

Species	HR	VES	Plots	Roads	Trans	Other	Total
barred tiger salamander	SO	62	0	0	0	0	62
Great Plains toad	C	0	0	1	0	0	1
red-spotted toad	C	137	1	48	3	0	189
Woodhouse's toad	C	0	0	0	0	0 <sup>a</sup>	0
barking frog	SO	21 <sup>b</sup>	0	0	0	1	22
canyon treefrog	SO	12	4	0	0	0	16
New Mexico spadefoot	C	91	0	15	0	0	106
<b>Total amphibians</b>		<b>323</b>	<b>5</b>	<b>64</b>	<b>0</b>	<b>1</b>	<b>396</b>

<sup>a</sup> Woodhouse toads (15 individuals) were observed by B. N. Alberti in 2000.

<sup>b</sup> Minimum estimate of barking frogs based mainly on number of calling individuals; does not include frogs collected during study.

**Table 4.3. Species of reptiles confirmed at Coronado NM, and number of observations during 1997–1998 on general visual encounter surveys (VES), monitoring plots (Plots), road transects (Roads), the Lowe-Johnson transect (T rans), and incidental observations (Other). HR refers to historic records: C = confirmed by specimen during previous study; SO = previous sight observation; NR = no known records from memorial. Potential records from Montezuma Canyon are labeled with an asterisk (\*). Identification of whiptail lizards, which are difficult to identify without handling, are based on recognizable field features.**

Species	HR	VES	Plots	Roads	Trans	Other	Total
<b>Lizards</b>							
Sonoran spotted whiptail	C	257	172	23	75	5	532
desert grassland whiptail	C	141	87	2	15	7	252
unidentified whiptails ( <i>Cnemidophorus</i> spp.)	—	199	31	15	11	9	265
eastern collared lizard	NR*	1	0	0	0	0	1
Madrean alligator lizard	C	9	0	0	0	1	10
mountain skink	SO	6	0	0	0	0	6
Great Plains skink	NR*	11	0	0	0	1	12
Gila monster	SO	0	0	0	0	0 <sup>a</sup>	0
lesser earless lizard	C	25	5	2	4	1	37
short-horned lizard	C	2	0	1	1	2	6
Clark's spiny lizard	C	79	45	42	12	16	194
Yarrow's spiny lizard	C	158	15	4	23	9	209
bunch grass lizard	NR	8	0	0	0	1	9
prairie lizard	NR*	13	3	4	0	0	20
tree lizard	C	84	82	2	12	15	195
<b>Subtotal (lizards)</b>		<b>993</b>	<b>440</b>	<b>95</b>	<b>153</b>	<b>67</b>	<b>1748</b>
<b>Turtles and Tortoises</b>							
ornate box turtle	NR	1	0	0	0	0	1

<sup>a</sup>Individuals of Gila monsters, western hognose snakes, and western coral snakes were collected after 1998.

4.2) and 1,896 observations of 33 species of reptiles (Table 4.3) at Coronado NM during this study. This total includes all observations made during 1997 and 1998, but does not include observations made by the co-authors and memorial staff during 1996 and after 1998. Four species not observed during our study have been confirmed at the memorial between 1998 and 2001:

Woodhouse's toad, Gila monster, western coral snake, and western hooknose snake. We observed all of the species observed at the memorial by Johnson and Lowe (1979) and confirmed 22 species that they had not confirmed or observed. However, several of these species had been sighted at the memorial by staff or visitors or had been collected on lands that are probably now part of the memorial.

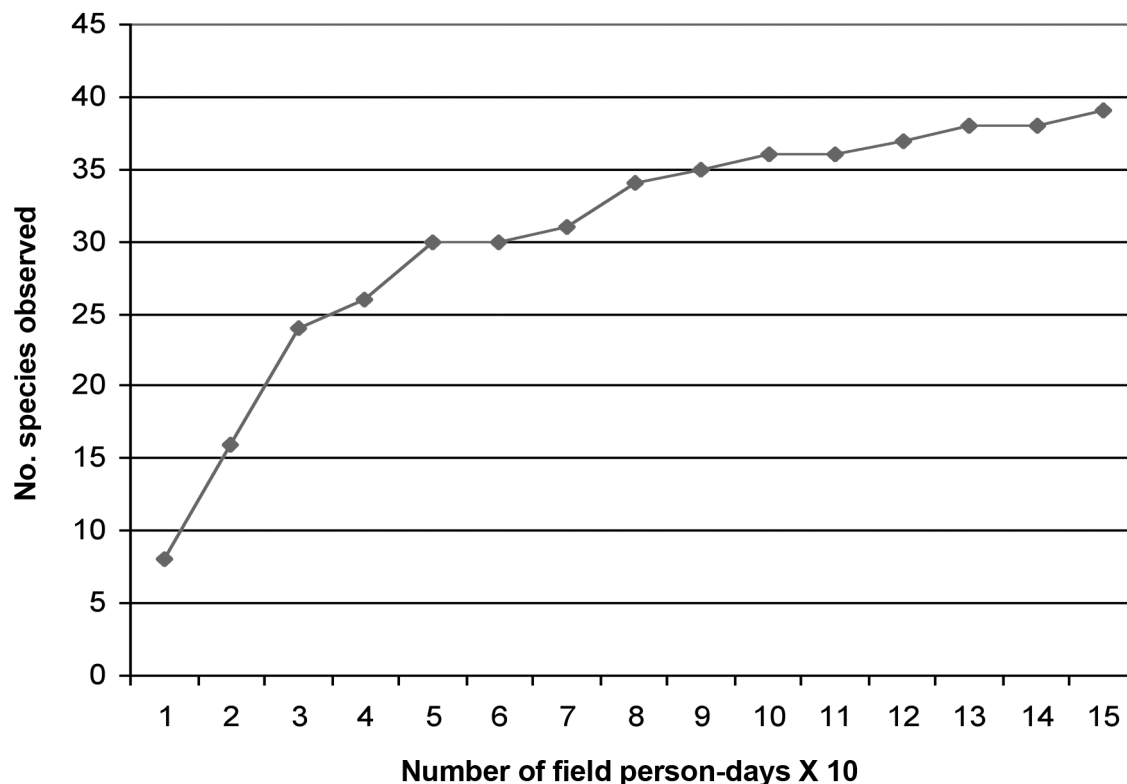
### *Estimates of Species Richness*

Figure 4.2 indicates the rates at which new species were encountered during the approximately 150

field person-days of our study. At the end of the 1997 season 30 species had been observed, and we encountered an additional nine species for the first time in 1998. Estimates of species richness (number of species present) at Coronado NM were obtained using Program CAPTURE. The Program chose Model (bh) (and the estimator generalized removal) as the most appropriate model for the data, suggesting that species observability varied both among species and that field researchers varied their search effort in response to finding species. Program CAPTURE estimated the species richness of amphibians and reptiles at the memorial to be 39 (SE = 1.132, CI = 39–46), which is identical to the number of species detected during our study but is an underestimate since at least 43 species are known to occur on site.

### *Visual Encounter Surveys*

Visual encounter surveys, including observations made while we moved between plots, accounted



**Figure 4.2. Species accumulation curve for reptiles and amphibians combined at Coronado NM, 1997–1998.** Each symbol represents the total number of species observed (y-axis) after a certain number of field person-days (x-axis); thus, eight species had been observed by the tenth person-day of the study, and 30 species had been observed by the 50<sup>th</sup> day.

for 57% of all observations of reptiles and amphibians during the study. In total, we spent 606 hours on visual encounter surveys (Table 4.4). Because of the great variability and overlap of vegetation communities it was not possible to precisely quantify the amount of sampling conducted in each vegetation community. However, based on the dominant vegetation in each area sampled, we estimate that time spent was approximately 26% in oak woodland, 17% in semi-desert grassland, 46% in riparian woodland, 7% in mesquite riparian, and 3% at perennial seeps (Coronado NM consists of approximately 72% oak woodland, 20% semi-desert grassland, and 8% riparian woodland, mesquite riparian, and seeps). The exaggerated amount of time in riparian woodland was due to intensive searching of this vegetation community during the rock rattlesnake/Sonoran mountain kingsnake study, which was included in our visual encounter survey time.

A total of 1,073 individual reptiles and 323 individual amphibians (not including larvae) were observed on visual encounter surveys. Mean number of observations/hour of reptiles (Table 4.4) exhibited temporal variability. During months with >20 sampling hours (May–October), observations/hour ranged from a low of 1.1 in October to a high of 2.4 in July. Observation rates varied among researchers as well; for the three observers with >50 sampling hours, observations/hour ranged from 2.23 (DES) to 1.25 (TE) observations per hour. However, the low rate for Taylor Edwards is probably the result of many hours spent searching for rock rattlesnakes and Sonoran mountain kingsnakes in a vegetation community (higher-elevation oak woodland) with lower relative abundance of reptiles than other communities. Rates of encounter of species followed similar patterns as rates of encounters of individuals, with a few exceptions (Table 4.4). The number of species observed per hour was highest

**Table 4.4. Summary of visual encounter survey data at Coronado NM, 1997–1998.** Includes all reptile observations on site; amphibians are excluded because of the extreme variability of observations. “Other/Combined” refers to results by short-term field researchers or where two or more researchers were combined on data sheets.

Month/Personnel	Person hours	Number of individuals observed	Number of observations per hour	Number of species observed	Number of species observed per hour
Month					
April	4.8	6	1.30	3.0	0.63
May	129.2	398	3.10	15.0	0.12
June	53.5	79	1.50	11.0	0.21
July	60.0	142	2.40	14.0	0.23
August	173.2	207	1.20	19.0	0.11
September	137.1	187	1.40	21.0	0.15
October	48.2	54	1.10	14.0	0.29
<b>Totals</b>	<b>606.0</b>	<b>1073</b>	<b>1.80</b>	<b>29.0</b>	<b>0.05</b>
Personnel					
Swann	221.5	493	2.23	20.0	0.09
Bell	104.7	206	1.97	18.0	0.17
Edwards	232.9	289	1.25	18.0	0.08
Other/Combined	46.9	85	1.81	8.0	0.17
<b>Totals</b>	<b>606.0</b>	<b>1073</b>	<b>1.78</b>	<b>29.0</b>	<b>0.05</b>

**Table 4.5. Summary of surveys on monitoring plots at Coronado NM, 1998.** Includes all reptile observations; only five amphibians were observed.

Month/Personnel	Person hours	Number of individuals observed	Number of observations per hour	Number of species observed	Number of species observed per hour
Vegetative Community					
Month					
May	35.0	179	5.1	10	0.29
June	12.5	55	4.4	5	0.40
July	9.0	83	9.2	5	0.56
August	30.0	34	1.1	7	0.23
September	24.0	73	3.0	6	0.25
October	3.5	24	6.9	5	1.43
<b>Total</b>	<b>114.0</b>	<b>448</b>	<b>3.9</b>	<b>29</b>	<b>0.25</b>
Personnel					
Swann	48.5	225	4.6	11	0.22
Bell	32.5	107	3.3	5	0.15
Edwards	26.5	95	3.7	8	0.30
Hare	6.5	21	3.2	5	0.77
<b>Total</b>	<b>114.0</b>	<b>448</b>	<b>3.9</b>	<b>29</b>	<b>0.25</b>
Vegetation Community					
Riparian Woodland High	8.0	11	1.4	4	0.50
Riparian Woodland Mid	18.0	62	3.4	4	0.22
Oak Woodland High	20.0	57	2.9	4	0.20
Oak Woodland Low	24.0	54	2.3	4	0.17
Mesquite Riparian	18.0	186	21.5	8	1.00
Semi-desert Grassland	16.0	49	3.1	6	0.38
Perennial Seeps	10.0	29	2.9	4	0.40
<b>Total</b>	<b>114.0</b>	<b>448</b>	<b>3.9</b>	<b>29</b>	<b>0.25</b>

in October and lowest in August (excluding the small April sample).

### Monitoring Plots

We sampled on 25 regular monitoring plots for a total of 114 hours during 1998, and five observations of two amphibian species (excluding larvae) and 448 observations of 13 species of reptiles were made (Table 4.5). Mean number of reptiles observed during 30-minute surveys was 1.96 (SE = 0.176). Mean number of observations/hour varied by month, with the highest number of observations in July and October (Table 4.5). There was no significant difference among the three primary researchers in number of individuals observed per survey (MS = 4.7774, F = 0.6462, p = 0.525). As with a study at Fort Bowie National Historic Site using nearly identical methods (Swann et al. 2001), encounter rates on monitoring plots for all researchers exceeded encounter rates on generalized visual encounter surveys.

Numbers of reptiles observed on surveys did vary greatly among different vegetation communities (MS = 75.6859, F = 15.0473, p < 0.0001). More reptiles per hour were observed in the mesquite riparian vegetation community, found at lower elevations in Montezuma Canyon, than in other communities sampled (Table 4.5).

### Historic Transects

The five transects established by Johnson and Lowe (1979) were sampled a minimum of four

times, including at least once each during summer 1997, spring 1998, and summer 1998. Mean number of reptiles varied greatly among transects, with highest abundance of reptiles (7.57 individuals/transect) on transect 3 in Oak Woodland along lower Joe's Canyon Trail and the lowest abundance (1.33 individuals/transect) on transect 5 in Pine-Oak Woodland.

Table 4.6 provides a summary of 1978 and 1997–1998 results. It is difficult to compare the two datasets because the original data from Johnson and Lowe (1979) have been lost and it is not known what their sampling effort was. Johnson and Lowe (1979) state that “Relative abundance categories are based on, and indicate, what an observer might expect to see during an activity peak for a particular species of lizard.” They do not provide any information on the length of their surveys, but we might assume that they are for the entire period of peak activity on a given day. Because we surveyed both within and outside “activity peaks”, our data in Table 4.6 are presented as peak values only, or the highest number of that species observed on any transect survey. In general, our peak numbers are lower than those presented by Johnson and Lowe (1979), possibly because our surveys were of shorter duration. It is also interesting that the numbers of lesser earless lizards observed by Johnson and Lowe (1979) greatly exceed the number that we observed. In contrast, they observed very few Sonoran whiptails in semi-desert grassland community, while this species was very abundant on our transect surveys.

**Table 4.6. Comparison of relative abundance of lizards in each vegetation community at Coronado NM based on transect data from Johnson and Lowe (1979) and our study.** The 1978 column represents the range of seasonal (peak value) abundance for each species in the each community, while the 1997–1998 column represents peak values observed on transects sampled during our study.

Species	Semi-desert grassland (Transect 1)		Oak woodland (Transect 3)		Pine-oak woodland (Transects 4-5)		Riparian habitat (Transect 2)	
	1978	1997–1998	1978	1997–1998	1978	1997–1998	1978	1997–1998
lesser earless lizard	5–25	0	5–15	0	1–5	1	5–15	1
Yarrow's spiny lizard	0	0	5–>25	1	1–15	4	0	0
Clark's spiny lizard	1–5	0	5–>25	2	1–5	1	5–>25	2
tree lizard	>25	0	5–15	3	1–5	3	5–>25	1
short-horned lizard	1–5	0	0	0	0	1	0	0
Sonoran spotted whiptail	0	2	5–>25	24	0–5	6	1–25	6
grassland whiptail	1–25	6	0	0	0	0	0	0
Madrean alligator lizard	0	0	0–5	0	0	0	0–5	0

**Table 4.7. Summary of road transect survey data at Coronado NM, 1997–1998.** Includes all reptile and amphibian observations on and off site. Total distances rounded to nearest mile or kilometer.

	Total distance in kilometers	Number amphibians	Number reptiles	Total hours	Amphibians per kilometer	Reptiles per kilometer	Amphibians per hour	Reptiles per hour
Month								
April	21	0	0	0.6	0.00	0.00	0.00	0.00
May	400	0	18	20.3	0.00	0.05	0.00	0.89
June	162	0	11	8.5	0.00	0.07	0.00	1.29
July	520	40	47	33.2	0.08	0.09	1.20	1.42
August	1010	24	29	47.2	0.02	0.29	0.51	0.61
September	502	0	16	23.8	0.00	0.32	0.00	0.67
October	326	0	11	13.1	0.00	0.34	0.00	0.84
<b>Totals</b>	<b>2941</b>	<b>64</b>	<b>132</b>	<b>146.5</b>	<b>0.02</b>	<b>0.04</b>	<b>0.44</b>	<b>0.90</b>
Year								
1997	1258	24	60	64.8	0.02	0.05	0.37	0.93
1998	1683	40	72	81.7	0.02	0.04	0.49	0.88

### Road Transects

A total of 146.5 hours on 113 days and/or evenings was spent searching on roads for reptiles and amphibians during this study (Table 4.7). A total of 2,941 km were driven on East Montezuma Canyon Road or smaller roads within the boundaries of the memorial; we also drove outside of the boundary, but do not include these data in Table 4.7. We made 64 observations of three species of amphibians and 132 observations of 20 species of reptiles.

Observational trends on roads varied by time of year. Amphibians were only observed in July and August, coincident with summer rains. More reptiles were seen per sampling unit in July and August than in other months (Table 4.7). Trends for the two years, 1997 and 1998, were very similar for reptiles, but more amphibians were observed during 1998. The road transects detected one amphibian (Great Plains toad) and one reptile (longnose snake) that were not detected using other methods. However, three species of amphibians and 13 species of reptiles (including eight snakes) that were observed during this study were never observed on roads, nor were three of the four species confirmed during 1999–2000 after the study ended.

### Historic Records

In addition to Johnson and Lowe (1979), we located several important sources of information on herpetofauna at Coronado NM. A total of 37

museum specimens of reptiles and amphibians were found at four museum collections (University of Arizona, Chicago Academy of Sciences, Brigham Young University, and Harvard University). Most specimens from the memorial are located at the University of Arizona and were collected by Terry Johnson in 1978. Two additional specimen records were found in published sources (Gloyd 1937, Fowlie 1965). A few of the older specimens were collected in Montezuma Canyon prior to the inclusion of the entire U.S. portion of this canyon within the boundaries of the memorial. Information (location, collection numbers, collector, and date) for all specimens known to be collected at the memorial is provided in Appendix H.

### Additional Records

Additional records of reptiles and amphibians on, and in, the vicinity of the memorial were obtained from a variety of published and unpublished sources, as well as by incidental observations and road riding during our study. In addition to species observed on monitoring plots, transects, and during visual encounter surveys, nine species of reptiles and one species of amphibian were observed on site during our study; these records are recorded in the study database and summarized in the “Other” column in Tables 4.2 and 4.3. In addition to reptiles and amphibians, we recorded all observations of mammals during our study. Mammal observations are summarized in Chapter



6 and in the COROWILD database kept at the memorial.

## Inventory Completeness

Based on the species accumulation curve, and considering the previous inventory effort (Johnson and Lowe 1979) we believe that our inventory was fairly complete. With one exception, we observed all of the species observed at the memorial by Johnson and Lowe (1979). The species accumulation curve (Figure 4.2) indicates that we continued to detect new species throughout 1997–1998, but we further benefited by the continuing active presence of Barbara Alberti at the memorial following completion of fieldwork, as four species not observed during our study were later confirmed. Program CAPTURE estimated the species richness of amphibians and reptiles at the memorial to be 39 (SE = 1.132, CI = 39–46), which is identical to the number of species detected during our study but is an underestimate of the total count of 43 species for the site.

Nevertheless, it is noteworthy that the Huachuca Mountains and adjacent valleys contain approximately seven amphibian and 26 reptile species not observed by us (see Appendix E). However, many of these species require more surface water than is present at the memorial, and others are species associated with valley bottomland habitats that do not occur at the memorial. The two montane rattlesnake species that occur close by but were not found by us are probably absent, or occur in very low numbers, due to the lack or scarcity of key habitat components.

## Discussion

### *Reptile and Amphibian Community*

Our results indicate that Coronado NM supports a moderately diverse herpetofauna of seven

amphibian and 36 reptile species. Reptiles at the memorial rank second in species richness (number of species) and third in species density (species/1,000 ha) among the four national parks and monuments in southern Arizona that have been intensively inventoried (Table 4.8). Obviously, both species richness and density are important in determining relative biodiversity of each park unit. The smallest two parks inventoried, Tonto NM and Fort Bowie National Historic Site, have species densities 3–4 times that of third smallest, Coronado NM, and about 200 times that of Organ Pipe Cactus NM, the largest of the parks by far. Primary reasons for this variation lie in the relative amounts of the various biomes comprising the respective parks, with some biomes being much more diverse in reptile species than others.

The memorial's diversity results from several factors, including its fairly large elevation range and location in the overlap zones among the Chihuahuan and Sonoran Deserts, the Rocky Mountains, the Sierra Madre Occidental, and the Great Plains. The upper elevations of the memorial contain many of the taxa with restricted U.S. ranges associated with the sky islands, including the mountain skink, Yarrow's spiny lizard, bunch grass lizard, banded rock rattlesnake, Madrean alligator lizard, and Chihuahuan blackhead snake. The unique limestone areas provide rare habitat for barking frogs.

Lower elevations include Great Plains species that are near the western limits of their range, such as the western hognose snake, the Texas blind snake, and the ornate box turtle, as well as a few Sonoran Desert species that are close to the eastern limit or their ranges such as the Gila monster and Sonoran spotted whiptail.

### *Comparisons with Johnson-Lowe Study*

As noted previously, it is difficult to compare our study directly to Johnson and Lowe (1979) because

**Table 4.8. Numbers of species and species densities of native reptiles at the four inventoried parks in southern Arizona, arranged by decreasing size.** Original data from Rosen and Lowe (1996) for Organ Pipe, Swann et al. (1996) for Tonto, and Swann et al. (2001) for Fort Bowie.

Park name	Area in ha	Number reptile species	Species density (spp/1000 ha)
Organ Pipe NM	133,830	43	0.32
Coronado NM	1,900	36	18.90
Tonto NM	461	33	71.60
Fort Bowie NHS	405	25	61.70

of a lack of information on their methodology and changes in the memorial boundaries since that study. However, their report contains a wealth of information on relative abundances of different species in different vegetative communities, information that allows us to make some general observations. Amphibians such as the New Mexico spadefoot and red-spotted toad were observed frequently in both studies, and Johnson and Lowe (1979) were the first to note the presence of barking frogs. Two species that they listed as uncommon were either very rare or absent during our study; we observed only one Great Plains toad within the memorial boundaries, and no Woodhouse's toads were observed during 1997–1998. Likewise, we found two species (canyon treefrog and barred tiger salamander) that they did not observe. However, canyon treefrogs are very restricted in their distribution on the memorial and barred tiger salamanders were possibly introduced only recently. We are not certain that these possible changes in the

amphibian species composition are real, particularly since Woodhouse's toads were regularly observed at the memorial in 2000.

However, we believe that our confirming 20 species of reptiles (Table 4.9) not found by Johnson and Lowe indicates a real change in species composition since the 1970s, and is not simply the reflection of our larger sampling effort or observer differences between the two studies. Terry Johnson and Charles Lowe are both accomplished herpetologists. Lowe is considered the dean of Southwestern herpetologists, conducting numerous studies and surveys throughout Arizona and northern Mexico from the early 1950s into the 1990s. Johnson, now at the Arizona Game and Fish Department, specialized in inventories during and after graduate school at the University of Arizona.

Inspection of the lists of species of amphibians and reptiles (Table 4.9) and mammals (see Chapter 6) confirmed recently at the memorial for the first time leads us to believe that the recent

**Table 4.9. Species of amphibians and reptiles confirmed for Coronado NM that were not confirmed or observed by Johnson and Lowe (1979).**

Taxon	Common Name
Amphibians	barred tiger salamander
	canyon treefrog
Reptiles – Lizards	eastern collared lizard
	mountain skink
	Great Plains skink
	Gila monster
	bunch grass lizard
	prairie lizard
Reptiles – Turtles and Tortoises	ornate box turtle
Reptiles – Snakes	western diamondback rattlesnake
	rock rattlesnake
	Mojave rattlesnake
	western hooknose snake
	western hognose snake
	night snake
	common kingsnake
	Texas blind snake
	western coral snake
	Big Bend patchnose snake
	mountain patchnose snake
	Chihuahuan blackhead snake
	Blackneck garter snake

appearance of some of these species is likely related to habitat changes both within and outside the memorial since the 1970s. At least seven reptile species in Table 4.9 (Great Plains skink, bunch grass lizard, prairie lizard, ornate box turtle, Mojave rattlesnake, western hooknose snake and western hognose snake) show close affinities to grassland habitats. It is noteworthy that nine species of rodents, mostly grassland obligates, that were trapped in grasslands in 1996–1999 (see Chapter 6) were similarly never captured in those same areas in 1978 (Petryszyn and Cockrum 1979).

Within the memorial, there has also been a reduction of grazing extent and intensity since the 1970s. Photographs in Johnson and Lowe (1979) document extensive bare ground in the southwest corner of the memorial in an area now heavily vegetated with non-native and native grasses. We speculate that release from grazing likely caused the observed recent increases in grassland species. That hypothesis could also account for the greater number of coachwhips reported by Johnson and Lowe (their second most common snake) compared to our two observations. Coachwhips are fast-moving hunters usually found in open habitats, where they can see their prey as well as potential predators.

The relative numbers of lesser earless lizards and Sonoran whiptails at the memorial have clearly changed since the 1970s. Johnson and Lowe considered earless lizards to be “common” in most vegetative communities in 1978 and they did not see any Sonoran whiptails in semi-desert grassland habitat. We observed only 37 lesser earless lizards during the entire study, and found Sonoran whiptails to be the most common lizard on grassland plots, constituting 35% of all lizards seen. This dramatic shift in the proportions of the two species does not appear to follow simply from the release-from-grazing hypothesis. We speculate that the Sonoran whiptail, being larger, may have some competitive advantage over the earless lizard in areas where more grass is present. Lesser earless lizards are adapted to terrain with low-lying vegetative cover and loose, friable soils (Degenhardt et al. 1996), while Sonoran spotted whiptails are well-known for running quickly into vegetation when alarmed.

Another surprising change was the total absence of western diamondbacks, rock rattlesnakes, and Mojave rattlesnakes from the

Johnson and Lowe (1979) study. In our study, rock rattlesnakes (26 observations) and diamondbacks (23 observations) were the second and third most common snakes observed. It is possible that the population of rock rattlesnakes has increased at the memorial, but it is difficult to separate the difference between our results and those of Johnson and Lowe (1979) because our search efforts for this species were so intensive during our SPMA study in 1998 (Swann et al. 1999); it is worth noting that we observed only three rock rattlesnakes during 1997, before our focused study of them began. However, the failure of Johnson and Lowe (1979) to find the other two species is of great interest. Since cattle co-occur with western diamondbacks and Mojave rattlesnakes throughout both snake’s range, we can only speculate as to why none were found in the earlier study. A potential explanation is that these rattlesnakes are both nocturnal hunters who feed primarily on rodents, and that they have responded positively to the increase in rodent abundance that has accompanied the relaxation from grazing pressure.

## Monitoring

Establishing long-term monitoring programs in national park units such as Coronado NM is critical for long-term management. In addition, because parks are less likely than other areas to be degraded by human activities in the future, monitoring of park resources can provide great insight into environmental changes that may be taking place on a larger scale. If unlimited funds were available, it would be ideal to monitor the absolute abundance, distribution, and important life history parameters of every species of amphibian and reptile present in the memorial. Unfortunately, funds are always limited and there are many other important resource management, visitor, and interpretive priorities. A further problem in monitoring amphibians and reptiles is that many species are difficult to locate and identify compared to other taxonomic groups, and natural variability in abundance is high for most species. Gathering meaningful data on this taxonomic group can thus be fairly expensive and usually involves at least oversight by well-trained personnel.

In general, we have found that if monitoring programs in small parks focus first on comprehensive, repeatable inventories, they can be

funded over a longer cycle than is typical for most monitoring programs. We believe the greatest monitoring need is for information on changes in species diversity over time. The National Park Service is committed institutionally to preservation of biological diversity in national parks (NPS 1992), but local extinction of species has occurred in many parks in the past century (see Newmark 1995, Swann 1999). The potential loss of species at Coronado NM in future decades should be considered a real possibility as the memorial (and particularly the lower-elevation riparian areas) becomes isolated due to land development outside of the boundaries. Therefore, we suggest that species diversity of reptiles and amphibians be monitored periodically (every 10–20 years) by conducting site-wide surveys using a repeatable study design, as described in this report.

A second priority is focused monitoring of species of management concern. We did not observe any threatened and endangered species during our study. However, the barking frog is the obvious candidate for continued monitoring (Goldberg and Schwalbe 2004), and the Sonoran mountain kingsnake, banded rock rattlesnake, mountain skink, and bunch grass lizard are other potential candidates.

A third priority is specific studies of the impact of land-use changes on herpetofaunal communities. The greatest changes that may be expected at the memorial during the next decades will probably result from changes in grazing regimes and in the frequency of natural and prescribed fire. These three management implications are discussed in more detail below.

### *Monitoring Species Diversity Using Site-Wide Surveys*

Management would be assisted by conducting site-wide surveys at Coronado NM periodically using a repeatable study design. “Periodically” would ideally be once every five years, but intervals of once every 10–20 years may be more realistic. In part, that is, because site-wide surveys should always include at least two years of spring and summer surveys to account for dry years when some species may not be active. A repeatable study design is one that can be repeated by future researchers. We believe that if the data, methods, and time-frame outlined in this final report can be

accessed by future researchers, it will be possible to repeat our 1997–98 survey and directly compare results of future surveys with ours. However, we assume that there will be improvements in survey and analytical methodology in the years ahead so that our approach can be refined and improved upon. More specific information useful for long-term monitoring at the memorial is in Swann and Schwalbe (2002).

For evaluating changes in species richness (the number of species in an area), capture-recapture methodology (of species) could be used (see discussion earlier in this report). However, specific analytical methods are constantly being refined, and better methods for estimating species richness may become available. Loss of native species richness appears to accompany many types of human-caused environmental impacts (such as deforestation for agriculture, overgrazing, and land development; Rosenzweig 1995).

When species richness does decrease, species with very specific habitat needs are most likely to be extirpated first. Thus, the repeat surveys should take special note of individual species that appear to have declined in relative abundance or distribution at the site, which may then be singled out for more intensive study.

### *Single Species Monitoring*

Species that are intensively studied could be those identified by NPS and other agencies (such as Arizona Game and Fish Department or the U.S. Fish and Wildlife Service) as being of special concern. For species of special concern it is often possible to fund studies that can help identify important life habitat characteristics, habitat needs, and threats. For example, Saguaro National Park is currently studying desert tortoises because of their special status.

Currently, the species most deserving of intensive study at the memorial is the barking frog (Goldberg and Schwalbe 2000). Protocols for monitoring this species are being developed based on experiences gained during monitoring over the past decade. Even more than most amphibians, barking frogs are extremely difficult to locate and capture. Because of the habitat specificity of this species, the most cost-effective approach for long-term monitoring may be to look at changes in the distribution of calling males (C. Goldberg, pers.

comm.). Current demographic studies should be continued as long as sufficient staff and external resources are available. Because this unique species is so rare in the United States, the relatively large population at the memorial deserves special management and continued study.

For banded rock rattlesnakes and Sonoran mountain kingsnakes, we delineated a 0.5 km<sup>2</sup> study area where these species are most abundant (Swann et al. 1999). In 1998, we estimated population size using mark-recapture techniques and measured habitat variables (such as vegetative cover, distance to ant mounds, etc.) associated with these two species. Because of the rarity of Sonoran mountain kingsnakes and because of the difficulty of in-house monitoring of a venomous reptile, monitoring of these species has serious management implications. Both snakes are species of management concern because they are highly prized by collectors. One option, assuming funding is available, would be to use radio telemetry to develop a deeper understanding of the habitat, life history needs, and absolute abundances of banded rock rattlesnakes and Sonoran mountain kingsnakes.

Another option, if only limited funds are available, would be the monitoring of the abundance of these two species using the methods detailed in Swann et al. (1999). Monitoring could be funded through relatively small grants from NPS sources or the Southwest Parks and Monuments Association.

The fourth and fifth candidates for long-term monitoring are the mountain skink, which occurs in riparian woodland in the United States, and the bunch grass lizard, which occurs in grassland areas associated with pine-oak woodlands. Both are limited in their distribution in the United States and are of concern to the Arizona Game and Fish Department (AGFD 1996). Very little is known about the mountain skink (Degenhardt et al. 1996), and monitoring should include a more focused study of their life history and habitat requirements. Recent studies have shown that bunch grass lizards are very sensitive to habitat changes associated with grazing; they have declined dramatically in some areas of southeastern Arizona (Bock et al. 1990). For both species, a need is the designation of study areas based on range maps in Swann and Schwalbe (2002) Appendix C and mark-recapture studies conducted

annually over a period of 3–5 years to determine abundance, recruitment, and survival. Following these initial studies, monitoring can probably be based on periodic surveys using distance sampling line transects (Buckland et al. 1993) or a similar methodology.

### *Land-Use Change Studies*

Efforts to restore damaged habitats, such as the restoration of the grassland community at the memorial, have greatly increased on America's public lands in recent years. Accompanying these restoration efforts have been improvements in research methodology, particularly the Before-After-Control-Impact (BACI) study design (Underwood 1994) to measure and evaluate how restoration affects ecosystem components and processes. Fire, changes in the grazing regime, and other management activities at the memorial will affect reptiles and amphibians as well as communities of plants, invertebrates, small mammals, and birds. Measuring changes in these communities could be an important aspect of evaluating the success of restoration and natural processes. Although our proposed 10–20 year monitoring of species richness of herpetofauna at the memorial will provide data on general trends in restored areas, changes in the reptile and amphibian community changes will be better evaluated through focused studies conducted of specific organisms at a smaller scale. Usually, changes in population size of common species are studied because it is easier to obtain sample sizes that are large enough to detect a significant trend.

At Coronado NM, common high-elevation species most likely to be affected by fire are bunch grass lizards, Sonoran spotted whiptails, tree lizards, and Yarrow's spiny lizards. Bunchgrass lizards favor areas with higher grass cover, while Sonoran spotted whiptails favor areas with more woody cover. Tree lizards are strongly associated with trees in oak woodland areas, and Yarrow's spiny lizards are strongly associated with rocks in areas where the tree canopy is relatively open. Except for the bunch grass lizard, all of these species are common at the memorial and accurate abundance estimates with confidence intervals can be obtained through mark-recapture studies. However, if mark-recapture methods are used simultaneously with a non-capture method such as

line-transect distance sampling (Buckland et al. 1993), it may be possible to calibrate the latter method with the former so that marking may not be required after the first or second season. Whiptails are difficult to capture using lizard nooses, and pitfall traps may not be appropriate because of concerns about cultural resources, but methods that have been successful with other species in this genus include use of hollow cover tubes (Strong et al. 1993), funnel traps with drift fences (Karns 1986), and baited (hookless) fishing line (Strong et al. 1993).

Consistent with the BACI study design for restoration monitoring, use of multiple plots approximately 1 ha in size, which can be in the same location as plots used for small mammal, bird, or vegetation monitoring would help. Plots should be established in both restoration areas and non-restored control sites. The need is for annual sampling for several years to determine natural

among-year variability in population size, with less frequent sampling over a longer time frame.

Long-term monitoring of natural resources is essential in public land management, particularly in national parks that are to be managed in perpetuity. However, it is important to keep in mind that monitoring and research in parks have historically been difficult to sustain because of fluctuating financial resources, shifting agency priorities, and staff turnover. Systems for managing data, including geographic data, should be established in advance and resources should be dedicated to data entry and maintaining archives. More importantly, we believe that the most successful long-term programs require relatively simple but repeatable methods where sampling does not have to occur every year. These efforts can then be supplemented by short-term research on specific species and management activities as outlined above.

## Chapter 5: Bird Inventory

Brian F. Powell

The first comprehensive inventory of birds was conducted in 1977 and 1978 by Russell and Danforth (1979) who surveyed five transect routes multiple times in all seasons. They reported relative abundance based on the number of observations per hour of surveys. Although no original data exist from this effort (Steve Russell, pers. comm.), the transect routes were similar to those used by our survey crews and we therefore make gross comparisons between these two studies. Mike Guest, a volunteer at the memorial and an excellent birder, has maintained a bird-sightings database of records collected since 1991. Although most of the sightings in the database are from near the visitor center, the database nevertheless contains a wealth of information for completing the species list and evaluating the completeness of the inventory. From 1997 to 2003, personnel from the Southern Arizona Bird Observatory (Walraven and Wood 2002) banded birds as a part of the Monitoring Avian Productivity and Survivorship (MAPS) program (DeSante and O'Grady 2000). MAPS banding data from 1997 to 2001 were summarized by Martinez and Hubbard (2003) and data from all banding years were entered into the visitor center database that Mike Guest maintains. Susan Wethington and others banded hummingbirds at the memorial and other areas in the southwest in 2003 and 2004 as part of the Hummingbird Monitoring Network (Wethington 2004). Park personnel conduct annual surveys for Mexican spotted owls. We trained memorial volunteers in the use of the line transect method (described below), which they used for breeding-season and non-breeding season surveys from the fall of 2002 to the spring of 2004. Mendez and Desmond (2004) planned to study resource use by over-wintering grassland birds at the memorial. Since our surveys, the NPS in 2005 collected data for their landbird monitoring program (B. Powell, unpublished data). Finally, we located some records of specimens collected from the memorial (Appendix H). During both our study and the monitoring study, incidental observations of reptiles and amphibians, and mammals were collected (Appendix G).

### Goals

The purpose of this study was to complete an inventory for birds at Coronado NM. This effort was part of a larger biological inventory of eight NPS units in southern Arizona and southwestern New Mexico (Davis and Halvorson 2000, Powell et al. 2002, 2003, 2005a, 2006 Schmidt et al. 2006). This report supersedes Powell et al. (2005b). The goals of our inventory of Coronado NM were to:

1. Conduct field surveys with the goal of documenting at least 90% of the bird species expected to occur at the memorial.
2. Use repeatable sampling designs and survey methods (when appropriate) that allow estimation of parameters of interest (e.g., relative abundance) with associated estimates of precision.
3. Gather historic occurrence data from past studies and voucher specimens.

The bulk of our effort addressed goals number 1 and 2. To maximize efficiency (i.e., the number of species recorded by effort) we used field techniques designed to detect multiple species. We did not undertake single-species surveys for threatened or endangered species.

### Technical Concepts

This section introduces some technical concepts and considerations related to our inventory at Coronado NM.

#### *Sampling Design*

Sampling design is the process of selecting sample units from a population or area of interest (for a review, see Thompson [1992]). Unbiased random samples allow inference to the larger population from which those samples were drawn, and estimate the true value of a parameter. The precision of these estimates, based on sample variance, increases with the number of samples taken. Theoretically, random samples can be taken until all possible samples have been selected and

precision is exact; in this case a census has been taken and the true value is known. Non-random samples are less likely to be representative of the entire population because the sample may (intentionally or not) be biased toward a particular characteristic, perhaps of interest or convenience.

Our survey stations were not randomly located because we were more interested in detecting the maximum number of species than in providing inference to a larger area. Thus, abundance estimates (relative abundance, useful as an index to true abundance) detailed in this report may be biased because we surveyed in areas likely to have high species richness; however, the nature or extent of that bias is difficult to characterize or quantify. If population estimates were a higher priority, avoiding this potential bias would have greater importance.

### *Estimates of Abundance*

Estimating population size is a common goal of biologists, generally motivated by the desire to reduce (e.g., pest species), increase (e.g., endangered species), maintain (e.g., game species) or monitor (e.g., indicator species) population size. Our surveys at Coronado NM were generally focused on detecting species rather than estimating population size. In many cases, however, we present estimates of “relative abundance” by species, which is an index to population size; we calculate it as the number of individuals of a species recorded, scaled by survey effort. Some researchers (particularly plant, marine, and invertebrate ecologists) prefer to scale such frequency counts by the number of observations of other species, which provides a measure of community dominance based on abundance relative to other species present. If we completed multiple surveys in comparable areas (e.g., anywhere within Coronado NM), we included a measure of precision (usually standard error) with the mean of those survey results.

Indices of abundance are presumed to correlate with true population size but do not typically attempt to account for variation in detectability among different species or groups of species under different circumstances. Metrics (rather than indices) of abundance do consider variation in detection probability, and these include density (number of individuals per unit area; e.g.,

two crissal thrashers per km<sup>2</sup>), and absolute abundance (population size; e.g., 10 crissal thrashers at Coronado NM). These latter techniques are beyond the scope of our research. While it is true that indices to abundance have often been criticized (and with good reason, c.f. Anderson 2001), the abundance information that we present in this report is used to characterize the commonness of different species rather than to quantify changes in abundance through space (e.g., habitat-use studies) or time (e.g., monitoring). As such, relative-abundance estimates are more useful than (1) detectability-adjusted estimates of density for only a few species or (2) raw count data for all species without scaling counts by search effort. For a review of methods used to estimate abundance, see Lancia et al. (1996).

## **Data Organization**

### *Spatial Data*

Most spatial data are geographically referenced to facilitate mapping of study stations. Coordinate storage is the Universal Transverse Mercator (UTM) projection, using North American datum 1983 (NAD 83), Zone 12. We recorded UTM coordinates using hand-held Garmin eMap® Global Positioning System (GPS) units (Garmin International Incorporated, Olathe, KS; horizontal accuracy is about 10–30 m) because of their convenience and relative simplicity.

### *Inventory Databases and Data Archiving*

We entered field data into Microsoft Access (version 97) and checked all data for transcription errors. From these databases we reproduced copies of the original field datasheets using the “Report” function in Access. The output looks similar to the original datasheets but all data are easier to read. The database, printouts, and other data such as GIS layers will be distributed to memorial staff and to the University of Arizona, Special Collections, Main Library; Tucson, Arizona. Original datasheets will be given to the NPS Sonoran Desert Network (SDN) Inventory and Monitoring (I&M) program office in Tucson and may be archived at another location. This redundancy in data archiving is to ensure that these valuable data are never lost. Along with the archived data, we will include copies of the original datasheets and a guide to



filling out the datasheets. This information, in conjunction with the text of this report, should enable future researchers to repeat our work.

## Methods

We surveyed for birds at Coronado NM from 2002 to 2004. The majority of our inventory work took place in the spring of 2003 and 2004. We used four field methods: variable circular-plot (VCP) counts for diurnal breeding birds, nocturnal surveys for owls and nightjars, line transects for winter (i.e., non-breeding season) birds, and incidental observations for all birds in all seasons. Although winter bird surveys were not included in the original study proposal (Davis and Halvorson 2000), we felt they were important in our effort to inventory birds at the memorial because many species that use the area during the fall and winter may not be present during spring and summer (breeding season) surveys. We concentrated our primary survey effort on the breeding season because bird distribution is relatively uniform at that time due to territoriality among most landbird species (Bibby et al. 2002), and this uniformity increased our precision in estimating relative abundance and also enabled us to document breeding activity. Our survey period included peak spring migration times for most species, which added many migratory species to our list.

We also sampled vegetation around VCP stations. Vegetation structure and plant species composition are important predictors of bird species richness or the presence of particular species (MacArthur and MacArthur 1961, Rice et al. 1984, Strong and Bock 1990, Powell and Steidl 2002).

In most cases we do not report observations that failed to determine species (e.g., “unknown woodpeckers”). Ravens are an exception. Both Chihuahuan and common ravens occur at the memorial and they are difficult to differentiate unless viewed at short range under certain conditions or if they are seen flying together (Bednarz and Raitt 2002). We were not able to positively determine the species for any raven sighting and therefore report all observations as “unknown raven.”

## *General Vegetation Characteristics at Repeat-Visit VCP Stations*

We subjectively placed the two repeat-visit VCP transects (described below) in areas that we believed would have the highest species richness. The Wash transect incorporated elements of the semi-desert grassland and xeroriparian washes and it had considerably less overstory vegetation than the Riparian transect; three stations of the Wash transect were mostly semi-desert grassland with some areas containing no large trees but with velvet mesquite and Emory oak in the wash (Fig. 5.1; see also Fig. 5.2). The upper reaches of the Wash transect were more typical of an oak savannah, with increasing density of trees at stations numbers 7 and 8 (Table 5.1). The Riparian transect contained dense stands of oaks and alligator juniper in the canyon bottom with some Arizona sycamore along Montezuma Wash. The understory in the riparian area is more dense than upland areas with wait-a-minute bush, Mearn’s sumac, and Arizona baccheris dominating. The south-facing upland areas have a lower density of overstory trees and scattered shrubs in the understory such as Scott’s yucca. The north-facing slopes are oak woodland (Fig 5.1; see also Fig. 5.2 for aerial view).

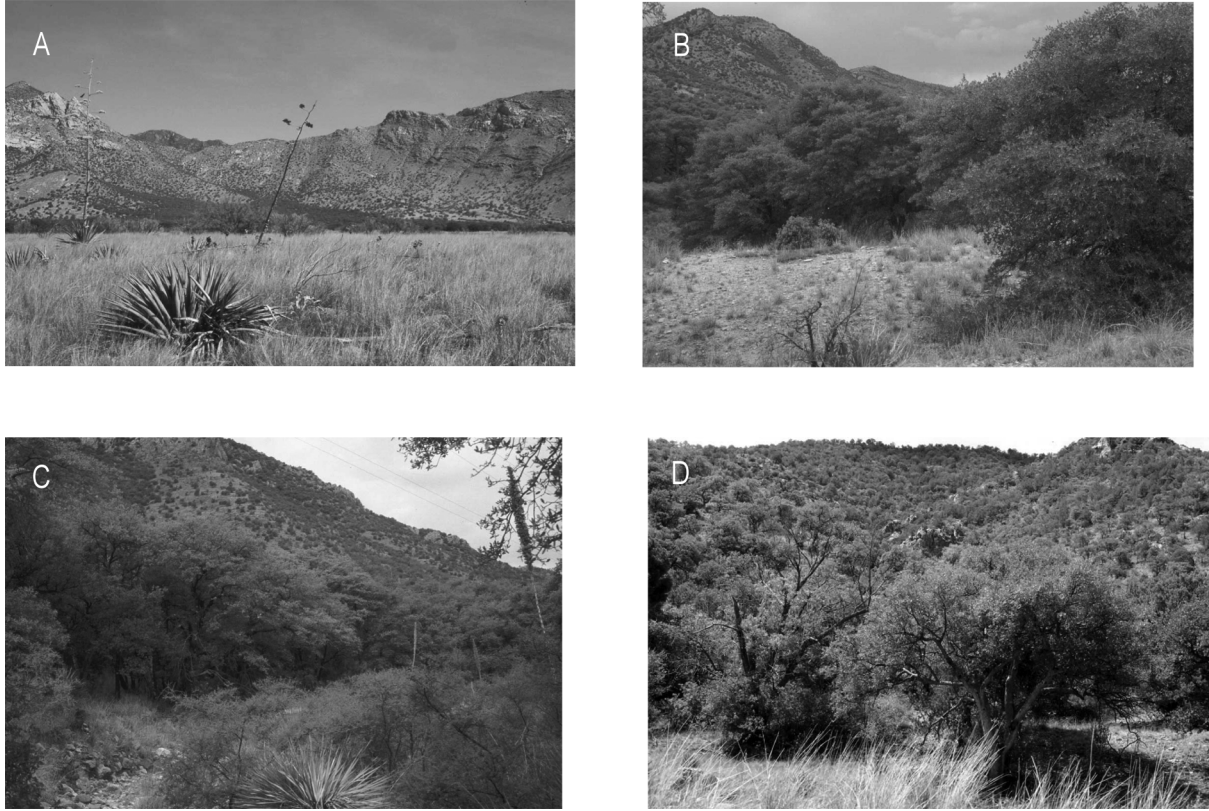
## *Spatial Sampling Designs*

We subjectively located all survey stations and transect sections (Figs. 5.2, 5.3) to encounter as many species as possible. Most repeat-visit VCP survey stations and line transects corresponded approximately to those used by Russell and Danforth (1979). Because of the inaccessibility of most areas of the memorial, we conducted reconnaissance VCP and nocturnal surveys along trails and old roads (Figs. 5.2, 5.3).

## *VCP Surveys*

### Field Methods: Repeat-visit Transects

We used the variable circular-plot (VCP) method to survey for diurnally active birds during the breeding season (Reynolds et al. 1980, Buckland et al. 2001). Conceptually, these surveys are similar to traditional “point counts” (Ralph et. al 1995) during which an observer spends a standardized length of time at one location (i.e., station) and records all birds seen or heard and the distance to each bird or group of birds.



**Figure 5.1. Photographs of bird stations along both repeat-visit VCP transects: Wash (A and B) and Riparian (C and D).** Photo A is looking north from Wash station number 2; B is looking west from Wash station number 8; C is looking west from Riparian station number 3; and D is looking east from Riparian station number 8. See Fig. 5.2 for location of stations.

We established two transects in 2003 that we surveyed repeatedly in both 2003 and 2004. Each transect consisted of eight stations, each located a minimum of 250 m apart to maintain independence of observations at each station. We surveyed each year from mid April through early July, the period of peak breeding activity for most species in the area.

Each year we visited the Riparian and Wash transects five times and four times, respectively. On each visit we alternated the order in which we surveyed stations (along a transect) to minimize bias by observer, time of day, and direction of travel. We did not survey when wind exceeded 15 km/h or when precipitation exceeded an intermittent drizzle. In 2003, we began bird surveys approximately 30 minutes before sunrise and concluded no later than three hours after sunrise. However, in 2004 we began surveys after 0700 hrs because of safety concerns.

We recorded a number of environmental variables at the beginning of each transect: wind speed (Beaufort scale), presence and severity of rain (qualitative assessment), air temperature (°F), relative humidity (%), and cloud cover (%). After arriving at a station, we waited one minute before beginning the count to allow birds to resume their normal activities. We identified, to species, all birds seen or heard during an eight-minute “active” period. For each detection we recorded distance in meters from the observer (measured with laser range finder when possible), time of detection (measured in one-minute intervals beginning at the start of the active period), and the sex and/or age class (adult or juvenile), if known. We did not measure distances to birds that were flying overhead nor did we use techniques to attract birds (e.g., “pishing”). We made an effort to avoid double-counting individuals. If we observed a species during the “passive” count period (between the eight-minute counts) that had not been recorded

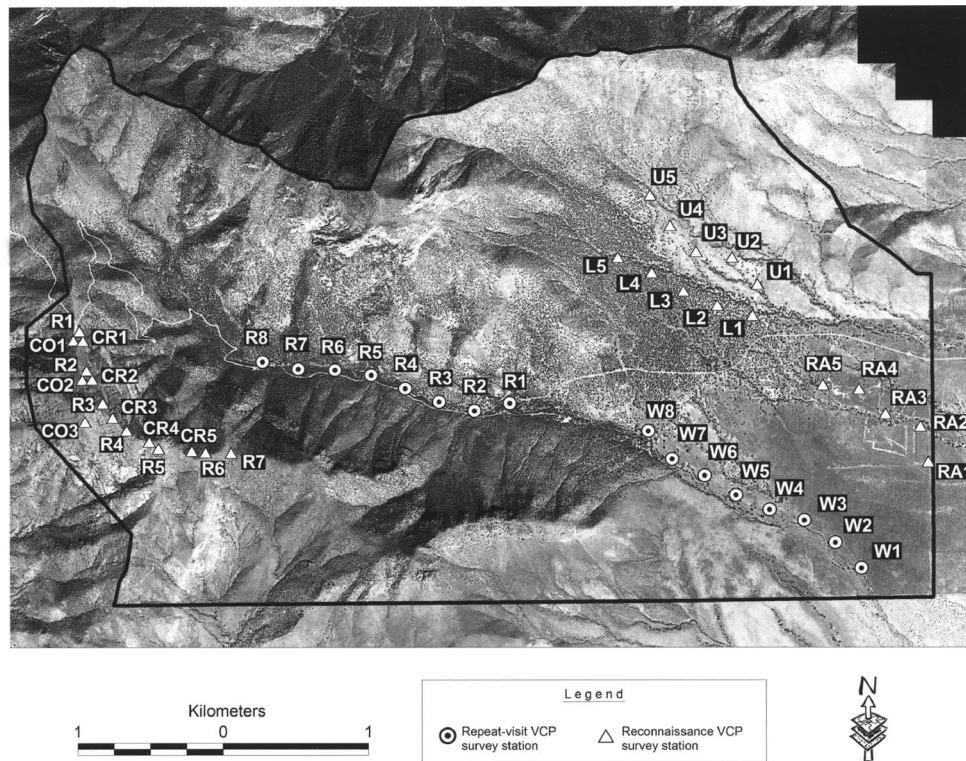


Figure 5.2. Location of VCP survey stations, Coronado NM, 2001 and 2002. See Appendix H for UTM coordinates.

**Table 5.1. Mean density of the most common tree species at each station along the two repeat-visit VCP transects, Coronado NM, 2004.** Data summarized from Appendix J. Density derived from data collected in the “tree” and “potential cavity-nesting” categories from point-quarter sampling (see text for description of field methods). Only species with >5 individuals per station are included in this summary. See Appendix A for scientific names.

Transect	Station	Tree Species						All oak species
		Arizona madrone	desert willow	alligator juniper	Mexican pinyon	Arizona sycamore	velvet mesquite	
Riparian	1			7.0	3.5	4.4		51.4
	2			4.7		4.7		69.3
	3				6.8			81.5
	4			3.0				110.1
	5	5.4		9.7		2.1		86.2
	6			14.6				125.4
	7	3.2		7.9	3.3			45.8
	8	6.0		18.1		14.8		32.6
Wash	1		8.1				10.8	7.8
	2		0.3				2.8	1.6
	3		2.1				1.8	2.4
	4						2.0	1.8
	5						2.1	4.9
	6						2.1	4.4
	7						1.7	14.0
	8						5.9	33.3

**Table 5.2. Summary of bird survey effort by UA inventory personnel, Coronado NM, 2002–2004.** Sample size was used in calculating relative abundance for each transect and each year.

Transect type (group)	Transect name	Year surveyed	Number of survey stations	Number of visits	Sample size
Nocturnal Survey	Owl	2003	variable	variable	15
Repeat-visit VCP	Riparian	2003	8	5	37 <sup>a</sup>
		2004	8	5	40
	Wash	2003	8	4	26 <sup>a</sup>
		2004	8	4	32
Reconnaissance VCP (high elevation)	Coronado Peak	2002	3	1	3
	Crest Trail	2003	5	1	5
		2004	5	1	5
	Ridge	2004	7	1	7
	(semi-desert grasslands)	Level	2004	5	5
		Ranch	2004	5	5
		Uplands	2004	5	5
Winter transect	Riparian	2002/2003	6	3	18
	Wash	2002/2003	6	2	12

<sup>a</sup> Survey effort was reduced on one survey because of wind.

previously at a station on that visit, we recorded its distance to the nearest station.

#### Effort: Repeat-visit Transects

We visited the Riparian transect five times and the Wash transect four times each, in both 2003 and 2004 (Table 5.1). We visited each station for eight minutes.

#### Field Method: Reconnaissance Transects

Most of our effort was focused on the two repeat-visit transects, but this left most of the memorial unsurveyed. Therefore, to get better spatial coverage of the memorial and still be able to make comparisons among transects, we established an additional six transects, located throughout the memorial (Fig. 5.2). For data collection, we followed the same protocol as for repeat-visit VCPs except that we spent five minutes at each station (instead of eight) and the distance between stations was generally >300 m. In 2004, we did not begin field work until after 0700 hrs, as with repeat-visit surveys.

#### Effort: Reconnaissance Transects

The number of survey stations along each transect ranged from three to seven and each transect was surveyed once except the Crest Trail transect, which we surveyed once in both 2003 and 2004

(Table 5.2). We visited each station for five minutes.

#### Analyses: All VCP Methods

We calculated relative abundance of each species along each transect as the number of detections at all stations and visits (including zero values), divided by effort (total number of visits divided by total number of stations). We reduced our full collection of observations for each repeat-visit VCP station ( $N = 1,338$ : 790 and 548 for Riparian and Wash transects, respectively) to a subset of data ( $n = 770$ : 487 and 283 for Riparian and Wash transects, respectively) that was more appropriate for estimating relative abundance. We used only those detections that occurred  $\leq 75$  m from count stations (thereby excluding 164 and 163 observations, respectively) because detectability is influenced by conspicuousness of birds (i.e., loud, large, or colorful species are more detectable than others) and environmental conditions (dense vegetation can reduce likelihood of some detections). Truncating detections may reduce the influence of these factors (Verner and Ritter 1983; for a review of factors influencing detectability see Anderson 2001, Farnsworth et al. 2002). We also excluded observations of birds that were flying over the station (90 and 77 observations,

respectively), birds observed outside of the eight-minute count period (72 and 41 observations, respectively), and unknown species (25 and 17 observations, respectively). Some observations met more than one of these criteria for exclusion from analysis.

For reconnaissance VCP transects, we calculated relative abundance in the same way as the repeat-visit VCP transects. We do not make comparisons between reconnaissance and repeat-visit transects because of the differences in the amount of survey time. Finally, we make comparisons of parameters and communities between years based on qualitative assessment of relative abundance and do not employ statistics, such as t-tests, to establish statistical differences between years, in part because of differences in methods of data collection.

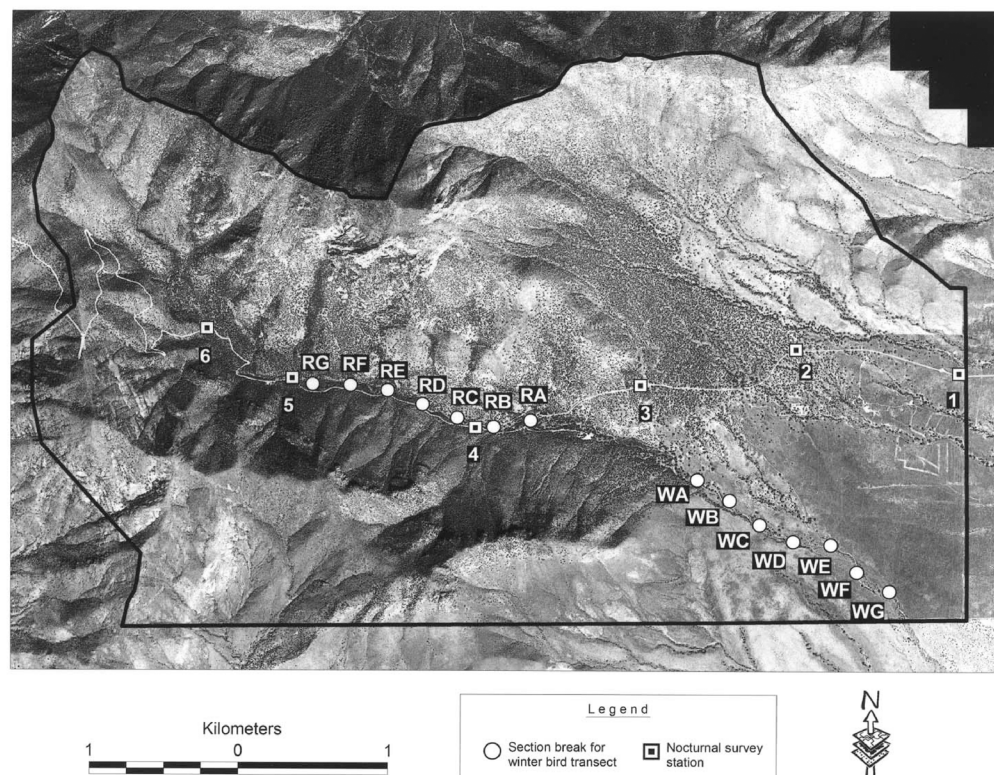
### *Line-transect Surveys*

#### Field Methods

We used a modified line-transect method (Bibby et al. 2002) to survey for birds from November 2002 to February 2003. Line transects differ from station

transects (used in our VCP surveys) in that an observer records birds seen or heard while the observer is walking a line, rather than standing at a series of stations. The transect method is more effective during the non-breeding season because bird vocalizations are less conspicuous and frequent, and therefore birds tend to be more difficult to detect (Bibby et al. 2002).

We established two transects at the memorial (Fig. 5.3). The transects were broken into sections, with the start and finish locations corresponding to repeat-visit VCP stations. Each section was approximately 250 m in length. As with other survey methods, we alternated direction of travel along transects to reduce biases, and did not survey during periods of excessive rain or wind (see VCP survey methods for details). We began surveys about 30 minutes after sunrise and continued until we completed the transect. As with VCP surveys, we recorded weather conditions at the beginning and end of each survey. Prior to beginning a section, we recorded the section name (e.g., “A–B”) and the start time.



**Figure 5.3.** Locations of non-breeding season (“winter”) transect sections and nocturnal survey stations for birds, Coronado NM, 2002 and 2003.

We timed our travel so that we traversed each section in ten minutes, during which time we assigned all birds seen and/or heard into one of the following distance categories:  $\leq 100$  m,  $> 100$  m, or “flyover.” When possible, we noted the sex and age class of birds. We recorded birds observed before or after surveys as “incidentals” (see section below), and we did not use techniques to attract birds (e.g., “pishing”).

#### Effort

We surveyed all six sections of the Riparian and Wash transects three and two times, respectively, in the fall/winter of 2002/2003. We attempted to complete additional surveys, but strong winds precluded effective surveys.

#### Analysis

Due to the low number of observations ( $n = 146$ ) within 100 m of the transect lines, we used all observations ( $N = 162$ ; except unknown species) to estimate relative frequencies of detections (see Methods section of VCP surveys for more details).

### *Nocturnal Surveys*

#### Field Methods

To survey for owls we broadcast commercially available vocalizations (Colver et al. 1999), using a compact disc player and broadcaster (Bibby et al. 2002), and recorded other nocturnal species (nighthawks and poorwills) when detected. We established one nocturnal survey transect along the Montezuma Pass Road (Fig. 5.3). The transect had six stations that were a minimum of 500 m apart. As with other survey methods, we attempted to reduce sampling biases by varying direction of travel along transects and by not surveying during periods of excessive rain or wind. We began surveys approximately 45 minutes after sunset. We conducted nocturnal surveys in 2003 only and did not survey in 2004 because of border-related safety concerns.

We began surveys at each station with a three-minute “passive” listening period during which time we broadcast no calls. We then broadcast vocalizations for a series of two-minute “active” periods and used vocalizations of species that we suspected, based on habitat and range information, might be present: elf, flammulated, northern pygmy, northern saw-whet, western screech, and whiskered screech owls. We excluded the great horned owl from the broadcast sequence

because of its aggressive behavior toward other owls, and we did not survey for Mexican spotted owls because that effort would have required a different protocol and because memorial staff survey annually for this species.

We broadcast recordings of owls in sequence of species size, from smallest to largest size species, so that smaller species would not be inhibited by the “presence” of larger predators or competitors (Fuller and Mosher 1987). During active periods, we broadcast owl vocalizations for 30 seconds followed by a 30-second listening period. This pattern was repeated two times for each species. During the count period we used a flashlight to scan nearby vegetation and structures for visual detections. If we observed a bird during the three-minute passive period, we recorded the minute of the passive period in which the bird was first observed, the type of detection (aural, visual or both), and the distance to the bird. If a bird was observed during any of the two-minute active periods, we recorded in which interval(s) it was detected and the type of detection (aural, visual, or both). As with other survey types, we attempted to avoid double-counting individuals recorded at previous stations. We also used multiple observers, alternated direction of travel along transects, and did not survey during inclement weather.

#### Effort

We surveyed each of the six stations at least once (Table 5.1), but the number of visits to each station varied because strong winds consistently interrupted surveys.

#### Analysis

We calculated relative abundance as per VCP surveys.

### *Line-transect Surveys: Memorial Volunteers*

We trained Coronado NM volunteers in the line-transect survey method, which they used from October 2002 to April 2004. They used the same method during the breeding and non-breeding season because the method relies less on auditory detection and more on visual detection of birds, which was the most accurate method of detection for the volunteers. The datasheets and methods of data collection were similar between our two studies except that volunteers typically used multiple observers because it was safer and more

enjoyable for them. The location of survey stations was similar to ours.

To summarize data from the memorial volunteers, we classified surveys completed from August through March as non-breeding season and surveys completed from April through July as breeding season surveys. We summarized data for only the transects that were visited on multiple occasions (Table 5.3). To calculate mean relative abundance, we used all detections including flyovers and species observed at distances >100 m.

### *Incidental and Breeding Observations*

#### Field Methods

When we were not performing formal surveys and encountered a rare species, a species in an unusual location, or an individual engaged in breeding behavior, we recorded UTM coordinates, time of detection, and (if known) the sex and age class of the bird. We recorded all breeding observations using the standardized classification system

developed by the North American Ornithological Atlas Committee (NAOAC 1990), which characterizes breeding behavior into one of nine categories: adult carrying nesting material, nest building, adult performing distraction display, used nest, fledged young, occupied nest, adult carrying food, adult feeding young, or adult carrying a fecal sac. We made breeding observations during standardized and incidental surveys.

#### Analysis

We report frequency counts of incidental and breeding observations; we could not calculate relative abundance because we did not standardize effort for this survey type.

### *Vegetation Sampling at Diurnal Breeding-Season Stations*

In 2004, we sampled vegetation associated with each of the breeding-season stations along the Riparian and Wash transects. We sampled

**Table 5.3. Bird survey effort by Coronado NM volunteers, 2002–2004.**

Survey season	Transect name	Year	Month	Number of sections surveyed
Non-breeding	Grassland	2002	October	5
		2002	December	6
		2003	January	6
		2003	February	6
		2003	September	6
		2003	December	6
	Ranch	2004	January	6
		2002	November	6
		2002	December	4
		2003	January	5
		2003	February	8
		2003	August	6
Breeding	Grassland	2003	April	4
		2004	April	6
	Ranch	2003	April	20
		2003	May	6
		2003	June	6
		2003	July	10
		2004	April	4
	Upland	2003	April	5
		2003	June	4
		2004	March	4

vegetation at five subplots located at a modified random direction and distance from each station. Each plot was located within a 72° range of the compass from the station (e.g., Plot 3 was located between 145° and 216°), to reduce clustering of plots. We randomly placed plots within 75 m of the stations to correspond with truncation of data used in estimating relative abundance.

At each plot, we used the point-quarter method (Krebs 1998) to sample vegetation by dividing the plot into four quadrants along cardinal directions. We applied this method to plants in one size category: potential cavity-bearing vegetation (> 20 cm diameter at breast height), and three height categories: sub-shrubs (0.5–1.0 m), shrubs (> 1.0–2.0 m), trees (> 2.0 m). If there was no vegetation for a given category within 25 m of the plot center, we indicated this in the species column. For each individual plant, we recorded distance from the plot center, species, height, and maximum canopy diameter (including errant branches). Association of a plant to a quadrant was determined by the location of its trunk, regardless of which quadrant the majority of the plant was in; no plant was recorded in more than one quadrant. Standing dead vegetation was only recorded in the “potential cavity-bearing tree” category. On rare occasions when plots overlapped, we repeated the selection process for the second plot.

Within a 5-m radius around the center of each plot, we visually estimated percent ground cover by type (bare ground, litter, or rock); and percent aerial cover of vegetation in each quadrant using three height categories: 0–0.5 m, > 0.5–2.0 m, and > 2.0 m. For both estimates, we used one of six categories for percent cover: 0 (0%), 10 (1–20%), 30 (21–40%), 50 (41–60%), 70 (61–80%), and 90 (81–100%).

### Analysis

Using point-quarter data, we calculated mean density (number of stems/ha) for all species in each of the four height/size categories. We used the computer program Krebs to calculate density (Krebs 1998). We collected these data to characterize gross vegetation characteristics around survey stations. In the event that future bird surveys detect marked changes in species or communities, the vegetation data reported in Appendix I will provide potential explanatory variables for changes in bird populations.

### *Nomenclature*

Scientific and common names used current according to American Ornithologists’ Union (AOU 1998, 2003).

### *Assessing Inventory Completeness*

Inventory completeness can most easily be assessed by (1) examining the rate at which new species were recorded in successive surveys (i.e., species accumulation curves; Hayek and Buzas 1997) and (2) by comparing the list of species we recorded with a list of species likely to be present based on previous research and/or expert opinion. For the bird species accumulation curve, we randomized the order of the sampling periods to break up clusters of new detections that resulted from temporal conditions (e.g., monsoon initiation) independent of cumulative effort. We used the computer program Species Richness and Diversity III (Pisces Conservation Ltd., IRC House, Pennington, Lymington, UK) to calculate the species accumulation curve where the order of samples is shuffled >10 times and the average is plotted, thereby smoothing the curve.

### **Results**

We recorded 129 species during surveys from 2002 to 2004 (Appendix C). We recorded 84 species during VCP surveys, 31 species during non-breeding season surveys, seven species during nocturnal surveys, and 113 species with incidental observations (Appendix C). We found five new species for the memorial: wild turkey, rock pigeon, yellow-billed cuckoo, Botteri’s sparrow, and northern cardinal. Of the 129 species that we found, a number have important conservation designations including the yellow-billed cuckoo, peregrine falcon, loggerhead shrike, and elegant trogon (Appendix C).

During the breeding season, the most widespread species (based on their occurrence at both repeat-visit and reconnaissance VCP transects) were the ash-throated flycatcher and Bewick’s wren, which we found at all eight transects (Appendix I). The mourning dove, rufous-crowned sparrow, Scott’s oriole, and house finch were also widespread and occurred at seven of the eight transects.



### *VCP Surveys: Repeat-visit Transects*

We recorded 84 species during breeding-season surveys to repeat-visit VCP transects (Appendix I). Species richness was similar between the Riparian and Wash transects, but composition of each community was quite different; we found 24 species on the Wash transect that we did not find on the Riparian transect and 29 species on the Riparian transect that we did not find on the Wash transect (Appendix I). Differences in community composition were not restricted to rare species; some of the most abundant species on each transect were not represented on the other transect. For example, the sulphur-bellied flycatcher and Hutton's vireo were found on the Riparian transect but not the Wash transect, whereas the verdin, cactus wren, Botteri's and Cassin's sparrow, and eastern meadowlark were found on the Wash transect but not on the Riparian transect (Appendix I; Tables 5.4, 5.5).

A difference in bird communities between these two transects is also evident in relative abundance estimates (Tables 5.4, 5.5). On the Riparian transect, the dusky-capped flycatcher, Mexican jay, black-throated gray warbler, spotted towhee, and black-headed grosbeak were all far more abundant than on the Wash transect. Conversely, the mourning dove and blue grosbeak were more abundant on the Wash transect than the Riparian transect. We found similarities in relative abundance between the two transects for the white-winged dove, bushtit, rufous-crowned sparrow, and house finch.

There were some striking inter-year differences in species richness and relative abundance estimates for each transect, yet caution should be exercised in making comparisons between 2003 and 2004 because of differences in sampling protocols (see Methods section). On the Wash transect, the Cassin's sparrow and blue grosbeak were two of the most common species in 2004 but were not abundant enough to estimate relative abundance in 2003 (Table 5.4). The ash-throated flycatcher and Mexican jay were far more abundant in 2004 than 2003, while the mourning dove had the opposite trend. The Bewick's wren and verdin had similar relative abundance scores in both years. On the Riparian transect, we found six greater roadrunners in 2003 but none in 2004 (Table 5.5). Relative abundance scores were

considerably higher in 2004 than in 2003 for the dusky-capped flycatcher, Mexican jay, and brown-headed cowbird. Relative abundance was similar between years for the bridled titmouse, Bewick's wren, black-throated gray warbler, spotted towhee, rufous-crowned sparrow, and Scott's oriole (Table 5.4).

### *VCP Surveys: Reconnaissance Transects*

We recorded 45 species at six reconnaissance VCP transects (Table 5.6). Species richness among sites ranged from 16 to 24 species, though the transect with the most species (Crest Trail) was surveyed only twice. The mourning dove, ash-throated flycatcher, and Bewick's wren were the most widespread species. Species with the highest relative abundance scores were the bushtit and Bewick's wren on the Level transect and the bushtit on the Ranch transect (Table 5.6).

### *Line-transect Surveys*

We found 31 species along two transects during non-breeding season surveys (Table 5.7, Appendix C). We found nine species that were not recorded during breeding-season surveys including the western bluebird, which was abundant during line-transect surveys (Table 5.7). We found 22 and 23 species along the Riparian and Wash transects, respectively, though the mean number of detections was much higher on the Riparian transect ( $7.2 \pm 3.4$  SE) than on the Wash ( $2.0 \pm 1.2$  SE) transect.

The Mexican jay and bushtit were the most abundant species along the Riparian transect and the bushtit and ruby-crowned kinglet were the most abundant species along the Wash transect (Table 5.7). The western bluebird and dark-eyed junco were very abundant on the Riparian transect but were not found on the Wash transect, and we found verdin along the Wash transect but not on the Riparian transect. Bushtit and ruby-crowned kinglet had similar relative abundance estimates for both transects. For other species, there were marked differences between transects, most notably for the Mexican jay, Bewick's wren, and spotted towhee, which were much more abundant on the Riparian than on the Wash transect.

### *Nocturnal Surveys*

We recorded five species of owls and two species of nightjars during nocturnal surveys in 2003

**Table 5.4. Sum (number of observations) and relative abundance (mean + SE) of birds observed during breeding-season surveys along the Wash transect, Coronado NM, 2003 and 2004.** Relative abundance estimates exclude birds observed > 75 m from stations, flyovers, and observations made outside of the eight-minute count period. See Methods section for additional details on estimation of relative abundance and effort used in those calculations. See Appendix C for scientific names and Appendix I for complete list of species observed.

Species	2003 (n =26)			2004 (n =32)			2003-2004
	Sum	Mean	SE	Sum	Mean	SE	Mean
white-winged dove	4	0.15	0.091				0.08
mourning dove	10	0.38	0.125	3	0.09	0.052	0.24
greater roadrunner	1	0.04	0.038				0.02
black-chinned hummingbird	1	0.04	0.038	2	0.06	0.063	0.05
ladder-backed woodpecker				4	0.13	0.059	0.06
Arizona woodpecker	2	0.08	0.053				0.04
western wood-pewee				1	0.03	0.031	0.02
Hammond's flycatcher	4	0.15	0.091				0.08
gray flycatcher	1	0.04	0.038				0.02
ash-throated flycatcher	7	0.27	0.105	22	0.69	0.130	0.48
Cassin's kingbird	3	0.12	0.064				0.06
western kingbird				2	0.06	0.043	0.03
Bell's vireo	1	0.04	0.038				0.02
plumbeous vireo	1	0.04	0.038				0.02
crissal thrasher	1	0.04	0.038				0.02
Mexican jay	2	0.08	0.053	22	0.69	0.263	0.38
bridled titmouse	1	0.04	0.038	10	0.31	0.176	0.18
verdin	7	0.27	0.089	8	0.25	0.078	0.26
bush-tit	5	0.19	0.136	5	0.16	0.156	0.17
cactus wren				2	0.06	0.063	0.03
Bewick's wren	24	0.92	0.156	23	0.72	0.136	0.82
house wren	1	0.04	0.038				0.02
ruby-crowned kinglet	9	0.35	0.123				0.17
phainopepla				2	0.06	0.063	0.03
orange-crowned warbler	1	0.04	0.038				0.02
Lucy's warbler	3	0.12	0.064	10	0.31	0.138	0.21
black-throated gray warbler	1	0.04	0.038				0.02
Townsend's warbler				1	0.03	0.031	0.02
hepatic tanager	1	0.04	0.038				0.02
western tanager				1	0.03	0.031	0.02
green-tailed towhee	2	0.08	0.053	1	0.03	0.031	0.05
spotted towhee	1	0.04	0.038				0.02
canyon towhee	5	0.19	0.096	9	0.28	0.112	0.24
Cassin's sparrow				12	0.38	0.125	0.19
Botteri's sparrow	8	0.31	0.133	18	0.56	0.127	0.44
rufous-crowned sparrow	7	0.27	0.105	9	0.28	0.112	0.28
chipping sparrow	10	0.38	0.385				0.19
lark sparrow	2	0.08	0.077	2	0.06	0.063	0.07
black-headed grosbeak				1	0.03	0.031	0.02
blue grosbeak				13	0.41	0.126	0.2
eastern meadowlark	1	0.04	0.038	6	0.19	0.070	0.11
brown-headed cowbird	1	0.04	0.038	1	0.03	0.031	0.03
Scott's oriole	1	0.04	0.038	1	0.03	0.031	0.03
house finch	2	0.08	0.077	8	0.25	0.100	0.16

**Table 5.5. Sum (number of observations) and relative abundance (mean + SE) of birds observed during breeding-season surveys along the Riparian transect, Coronado NM, 2003 and 2004.** Relative abundance estimates exclude birds observed > 75 m from stations, flyovers, and observations made outside of the eight-minute count period. See Methods section for additional details on estimation of relative abundance and effort used in those calculations. See Appendix C for scientific names and Appendix I for complete list of species observed.

Species	2003 (n =37)			2004 (n =40)			2003-2004
	Sum	Mean	SE	Sum	Mean	SE	Mean
turkey vulture	1	0.03	0.027				0.01
Cooper's hawk	3	0.08	0.045	1	0.03	0.025	0.05
white-winged dove	7	0.19	0.076				0.09
mourning dove	3	0.08	0.060				0.04
yellow-billed cuckoo				1	0.03	0.025	0.01
greater roadrunner	6	0.16	0.061				0.08
black-chinned hummingbird	1	0.03	0.027	2	0.05	0.035	0.04
Anna's hummingbird				3	0.08	0.042	0.04
broad-tailed hummingbird				4	0.10	0.048	0.05
acorn woodpecker	1	0.03	0.027				0.01
ladder-backed woodpecker				1	0.03	0.025	0.01
Arizona woodpecker	3	0.08	0.045	4	0.10	0.048	0.09
northern flicker	9	0.24	0.090	2	0.05	0.050	0.15
western wood-pewee	2	0.05	0.038				0.03
Hammond's flycatcher	3	0.08	0.060	2	0.05	0.035	0.07
gray flycatcher				1	0.03	0.025	0.01
western flycatcher <sup>a</sup>	2	0.05	0.038				0.03
cordilleran flycatcher				1	0.03	0.025	0.01
dusky-capped flycatcher	8	0.22	0.069	27	0.68	0.145	0.45
ash-throated flycatcher	11	0.30	0.085	6	0.15	0.067	0.22
sulphur-bellied flycatcher	6	0.16	0.082	4	0.10	0.060	0.13
Cassin's kingbird	2	0.05	0.054	1	0.03	0.025	0.04
plumbeous vireo	3	0.08	0.045				0.04
Hutton's vireo	5	0.14	0.057	14	0.35	0.111	0.24
crissal thrasher	1	0.03	0.027				0.01
western scrub-jay				1	0.03	0.025	0.01
Mexican jay	21	0.57	0.244	47	1.18	0.214	0.87
bridled titmouse	15	0.41	0.152	17	0.43	0.160	0.42
bushtit	11	0.30	0.122	5	0.13	0.064	0.21
white-breasted nuthatch				1	0.03	0.025	0.01
canyon wren	4	0.11	0.052				0.05
Bewick's wren	38	1.03	0.137	59	1.48	0.152	1.25
ruby-crowned kinglet	7	0.19	0.076	2	0.05	0.050	0.12
blue-gray gnatcatcher	1	0.03	0.027				0.01
hermit thrush	1	0.03	0.027				0.01
Lucy's warbler				4	0.10	0.070	0.05
yellow-rumped warbler				3	0.08	0.042	0.04
black-throated gray warbler	17	0.46	0.100	17	0.43	0.138	0.44
Townsend's warbler				1	0.03	0.025	0.01
Wilson's warbler	1	0.03	0.027	2	0.05	0.035	0.04
painted redstart				1	0.03	0.025	0.01
hepatic tanager	2	0.05	0.038	7	0.18	0.061	0.11
western tanager				3	0.08	0.055	0.04
spotted towhee	14	0.38	0.112	15	0.38	0.085	0.38

Species	2003 (n =37)			2004 (n =40)			2003-2004
	Sum	Mean	SE	Sum	Mean	SE	Mean
canyon towhee	1	0.03	0.027	3	0.08	0.042	0.05
rufous-crowned sparrow	9	0.24	0.105	12	0.30	0.096	0.27
chipping sparrow				1	0.03	0.025	0.01
black-throated sparrow				4	0.10	0.060	0.05
dark-eyed junco	1	0.03	0.027				0.01
black-headed grosbeak	6	0.16	0.073	5	0.13	0.053	0.14
blue grosbeak				1	0.03	0.025	0.01
brown-headed cowbird	2	0.05	0.038	12	0.30	0.073	0.18
hooded oriole				1	0.03	0.025	0.01
Scott's oriole	11	0.30	0.128	16	0.40	0.086	0.35
house finch	1	0.03	0.027	4	0.10	0.060	0.06

<sup>a</sup> Cordilleran or Pacific-slope flycatcher.

(Table 5.8). The most abundant species were the elf and whiskered screech owls.

### *Incidental and Breeding Observations*

We recorded observations of 113 species outside of other formal bird surveys (Appendix C). Of these species, 29 were not recorded by any other method and included: wild turkey, peregrine falcon, rock pigeon, elegant trogon, and northern cardinal. We made 32 observations that confirmed breeding for 19 species (Table 5.9). The most breeding observations were for the Cooper's hawk and Mexican jay (four breeding observations each). We made 12 observations of nest contents or adults feeding juvenile birds and we did not observe any brown-headed cowbird eggs, nestlings, or fledglings.

### *Line-transect Surveys: Volunteer Data*

Volunteers surveyed three transects during the breeding season and two transects during the non-breeding season and found 51 and 43 species, respectively, during those surveys (Appendices L, M). Of the 71 species recorded on the Ranch and Grassland transects, 21 were observed during the breeding season but not during the non-breeding season, and 27 species were observed during the non-breeding season but not during the breeding season. During the breeding season, the majority of survey effort was along the Ranch transect, where 47 species were observed. This was far more than on the Grassland (22 species) or the Upland (16 species) transects (Appendices L, M). The chipping sparrow had the highest relative abundance on any transect during the breeding season. Though it was

found in large flocks and early in the season, it was not found to breed at the memorial. Other abundant species were the Mexican jay, mourning dove, and house finch. During the non-breeding season, the eastern meadowlark and vesper sparrow had the highest relative abundance on the Grassland transect and the northern flicker had the highest relative abundance on the Ranch transect (Appendix L).

### **Inventory Completeness**

Based on our surveys and a review of past studies and current projects, we believe that the inventory of birds that regularly use the memorial is complete. A look at the species accumulation curve for our work indicates that our effort alone was not sufficient to document all of the species that occur on the memorial because the cumulative number of new species was not approaching an asymptote (Fig. 5.4).

Based on the species list from the visitor center database, the number of new species being added to the list did not begin to reach an asymptote until year eight or nine (Fig. 5.4). We found six new species for the memorial, but most of the new species that we found were located in the semi-desert grassland area, which is rarely visited by birders because of safety concerns and the lack of trails in that area.

Because birds are highly mobile animals, it is difficult to compile a truly complete list, especially for a place like the Huachuca Mountains, which is well-known for the occurrence of species that have their northern-most

**Table 5.6. Mean relative abundance of birds observed during reconnaissance VCP surveys, Coronado NM, 2002–2004.**  
See Appendix K for additional information. See Appendix C for scientific names.

Species	Group					
	High elevation			Grassland		
	Ridge	Coronado Peak	Crest trail	Level	Upland	Ranch
	2004	2002	2004	2003	2004	2004
white-winged dove						0.2
mourning dove		0.3		0.2		0.4
black-chinned hummingbird					0.4	
ladder-backed woodpecker	0.1			0.2	0.2	
western wood-pewee	0.3	0.2				
Say's phoebe	0.1		0.2			0.4
dusky-capped flycatcher		0.3				
ash-throated flycatcher			0.2	0.2	0.2	0.2
Cassin's kingbird		0.3				
western kingbird					0.2	0.8
curve-billed thrasher	0.3					
crissal thrasher				0.2		
western scrub-jay	0.6		0.8	1.0		
Mexican jay	0.1			0.8		1.6
bridled titmouse				0.4	0.2	
verdin						0.6
bushtit			0.8	0.2	5.0	1.4
white-breasted nuthatch				0.2		
cactus wren	0.1	1.0	0.2	0.2		0.4
canyon wren	0.1					
Bewick's wren	0.3	0.3		0.6	1.8	1.2
blue-gray gnatcatcher				0.4		
Lucy's warbler						0.2
hepatic tanager	0.3	0.3				
western tanager			0.4			
spotted towhee	0.4		0.2	0.8		
canyon towhee	0.6		0.2		0.2	0.2
rufous-crowned sparrow	1.1	0.3	0.4	0.4	0.4	
lark sparrow				0.2		
black-throated sparrow				0.2	0.2	
blue grosbeak						0.4
eastern meadowlark						0.6
brown-headed cowbird	0.1			0.2		0.2
hooded oriole						0.2
Scott's oriole	0.6			0.4	0.2	
house finch	0.1					0.8

**Table 5.7. Relative abundance of birds observed during line-transect surveys by UA inventory personnel, Coronado NM, 2002–2003.** See Appendix C for scientific names.

Species	Riparian ( <i>n</i> = 18)			Wash ( <i>n</i> = 12)		
	Sum	Mean	SE	Sum	Mean	SE
Montezuma quail	1	0.06	0.056			
northern harrier	1	0.06	0.056	1	0.08	0.083
sharp-shinned hawk				1	0.08	0.083
red-naped sapsucker	1	0.06	0.056			
ladder-backed woodpecker				1	0.08	0.083
northern flicker	3	0.17	0.121	2	0.17	0.112
Say's phoebe				2	0.17	0.112
Hutton's vireo	2	0.11	0.076	1	0.08	0.083
crissal thrasher				1	0.08	0.083
Mexican jay	42	2.33	0.796	4	0.33	0.333
common raven	1	0.06	0.056	2	0.17	0.112
bridled titmouse	7	0.39	0.293	1	0.08	0.083
verdin				5	0.42	0.149
bushtit	24	1.33	0.840	29	2.42	2.006
white-breasted nuthatch	3	0.17	0.121	2	0.17	0.167
canyon wren	1	0.06	0.056			
Bewick's wren	11	0.61	0.200	3	0.25	0.131
house wren				1	0.08	0.083
ruby-crowned kinglet	16	0.89	0.196	14	1.17	0.297
western bluebird	12	0.67	0.370			
mountain bluebird				2	0.17	0.167
hermit thrush	3	0.17	0.121			
phainopepla	1	0.06	0.056	1	0.08	0.083
yellow-rumped warbler				2	0.17	0.112
spotted towhee	14	0.78	0.222	1	0.08	0.083
rufous-crowned sparrow	2	0.11	0.076	3	0.25	0.179
chipping sparrow	13	0.72	0.434			
vesper sparrow				1	0.08	0.083
white-crowned sparrow	1	0.06	0.056			
dark-eyed junco	12	0.67	0.370			
lesser goldfinch	2	0.11	0.076	1	0.08	0.083

**Table 5.8. Sum (total number of observations) and relative abundance (mean  $\pm$  SE) of birds detected during nocturnal surveys, Coronado NM, 2003.** Sample size for calculation of relative abundance was 15.

Species	Sum	Mean	SE
western screech owl	6	0.40	0.190
whiskered screech owl	10	0.67	0.252
great horned owl	2	0.13	0.091
northern pygmy-owl	2	0.13	0.133
elf owl	13	0.87	0.192
common poorwill	6	0.40	0.163
whip-poor-will	3	0.20	0.107

**Table 5.9. Number of observations for each breeding behavior for birds, Coronado NM, 2003 and 2004.** Breeding behaviors follow standards set by NAOAC (1990). See Appendix C for scientific names.

Common name	Nest				Adults carrying			Feeding recently fledged young	Recently fledged young	Totals
	Building	With eggs	With young	Occupied	Food	Nesting material	Distraction displays			
Cooper's hawk	1		1	1					1	4
red-tailed hawk			1							1
Anna's hummingbird				1						1
broad-tailed hummingbird							1			1
northern flicker				1						1
Say's phoebe								1		1
dusky-capped flycatcher							1			1
sulphur-bellied flycatcher		1				1		1		3
western kingbird									1	1
Hutton's vireo		1		1	1					3
Mexican jay	1			1		1		1		4
cactus wren								1		1
Bewick's wren					2					2
curve-billed thrasher					1					1
crissal thrasher								1		1
hepatic tanager	1									1
rufous-crowned sparrow		1		1		1				3
eastern meadowlark			1							1
house finch								1		1
<b>Totals</b>	<b>3</b>	<b>3</b>	<b>3</b>	<b>6</b>	<b>4</b>	<b>3</b>	<b>2</b>	<b>6</b>	<b>2</b>	<b>32</b>

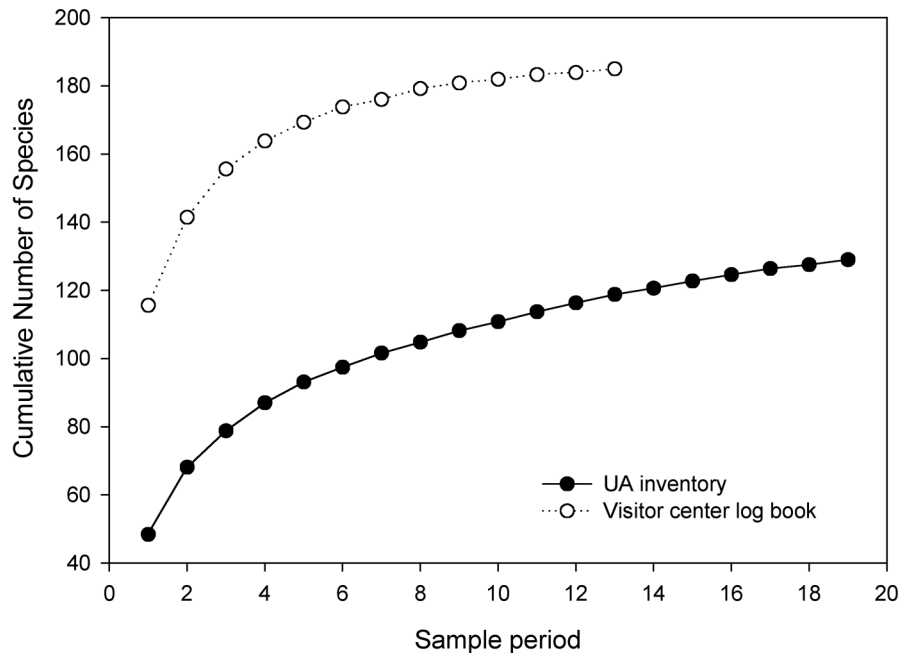
distribution in southern Arizona. Because of the variety of vegetation communities at the memorial, its location at the south end of the Huachuca Mountains, and the active birding community in the area, we believe that rare bird species will be added to the list for many years to come.

## Discussion

Results of our research and that of others indicate that Coronado NM has a very species-rich bird community. Collectively, there have been 196 bird species observed at the memorial, 2 (1%) of which are non-native. This richness results from two main factors. First, the location of the memorial at the south end of the Huachuca Mountains ensures that many regionally rare or unique species such as the Lucifer's, white-eared, and violet-crowned hummingbirds, elegant trogon, and streak-backed oriole are not uncommon. However, the memorial has no major drainages or springs, which precludes

the establishment of a large riparian area (as in nearby Carr or Ramsey canyons) and therefore fewer individuals of riparian-obligate birds are likely to stay at the memorial for long periods of time.

Second, the diversity of vegetation communities on the memorial, ranging from semi-desert grassland to oak savannah and woodland support a high diversity of birds. Although many environmental factors influence bird communities, vegetation characteristics are one of the most important predictors of avian community structure (James 1971). Important vegetation characteristics include vertical structure (MacArthur and MacArthur 1961, Cody 1981), horizontal patchiness (Roth 1976, Kotliar and Weins 1990), and floristics (Rice et al. 1984, Strong and Bock 1990). The changes in these resources at the memorial are exemplified in the gradient from the open semi-desert grassland with scattered trees in the Wash transect to dense vegetation along



**Figure 5.4. Species accumulation curves for the UA inventory effort (2002–2004) and data from the visitor center log book (1991–2003), Coronado NM.** Species accumulation data for UA effort include all observations from all survey types. Each sample period for UA inventory represents a randomized ordering of 103 observations ( $N = 1,952$ ; a completely randomized combination of the four survey types). Each sample period for the memorial's visitor center log book represents one year.

Montezuma Wash along the Riparian transect (see Fig. 5.1). The differences in bird community composition and relative abundance of the most common bird species (Tables 5.3, 5.4) between the Wash and Riparian transects (Appendix I) are pronounced but not surprising because of the differences in the vegetation characteristics between the two transects (Table 5.2, Appendix J). There were also significant within-transect changes in the bird communities between breeding and non-breeding seasons (Tables 5.5, 5.6).

The impact on the bird community from the conversion of the memorial's semi-desert grasslands from native species to one dominated by Lehmann lovegrass has not been established, but the loss of native grassland has been identified as a primary factor in population declines of grassland birds as a group (Herkert 1994, Knopf 1994, Peterjohn and Sauer 1999), including species of management concern such as the Botteri's and Cassin's sparrows. Since its introduction in the 1930s, Lehmann lovegrass has spread to occupy >400,000 ha in southern Arizona with little indication that its spread is complete (E. L. Geiger, unpublished data). We found the Cassin's sparrow

and Botteri's sparrow to be two of the most abundant species during breeding-season surveys along the Wash transect (Table 5.4). Unfortunately we were not able to establish whether these species nested at the memorial. Russell and Danforth (1979) considered the Cassin's sparrow common in Montezuma Wash, but Botteri's sparrow was not observed during their study. Initial studies indicate that relative abundance of birds and other taxa in these grasslands is lower in areas dominated by nonnative grasses (Bock et al. 1986). However, on the Fort Huachuca Military Reservation, north of Coronado NM, Albrecht and Steidl (In prep) found the density of Botteri's sparrow nests to be greater on plots dominated by Lehmann lovegrass than on native grassland plots, though nest success appeared to be lower on plots dominated by Lehmann lovegrass.

The native velvet mesquite has also increased in density and distribution in SE Arizona since the late 1800s, primarily due to disruption of historical fire regimes and grazing (Humphrey 1974, Brown 1994, Van Auken 2000). This encroachment has taken place at the memorial and has likely changed the bird community. Even in the



last 25 years, shrub-associated species, such as verdin and rufous-crowned sparrow, were some of the most abundant species in the memorial but were not found on the Wash transect in the late 1970s (Russell and Danforth 1979). Also, we observed the first record of northern cardinal at the memorial, a species that has presumably expanded its range due the increasing availability of dense vegetation in the shrub layer.

Sycamore trees line Montezuma Wash west of the visitor center. We found a nest of sulphur-bellied flycatchers in a sycamore tree there. Other species that prefer sycamore trees for nesting, and that we found along the Riparian transect, included the western wood pewee, painted redstart, and hooded oriole. These species were not common and we did not determine whether they nested in the memorial. Nevertheless, research in the southwestern U.S. has consistently shown that areas with riparian trees have bird communities that are more diverse than adjacent sites (Carothers et al. 1974, Szaro and Jakle 1985, Strong and Bock 1990), which is due, in part, to the variety of microhabitats that riparian vegetation provides for nesting, cover, and foraging (Powell and Steidl 2002). Riparian trees provide an abundance of nest substrates for primary- (i.e., mainly woodpeckers) and secondary-cavity-nesting species (e.g., elegant trogon, Lucy's warbler, and Bewick's wren). In addition to sycamore trees, the riparian area west of the visitor center also has a high density of oak trees. In this area, we observed nesting Cooper's hawks in 2003 and 2004, four wild turkeys, and a yellow-billed cuckoo.

#### *Comparison to Russell and Danforth (1979)*

The research by Russell and Danforth (1979) was the first comprehensive bird survey of the memorial and provides a valuable baseline for evaluating gross changes in the memorial's bird community. Because we did not use the same survey methods, and because the original data from that effort is lost, we are limited in our

comparisons. Yet some important patterns emerged. As mentioned earlier, we found the Botteri's sparrow to be among the most abundant species on the Wash transect, but it was not recorded by Russell and Danforth (1979). A look at the relative abundance rankings between our two studies also shows some interesting patterns. For the Riparian transect, most of the common species from each study were similarly common (Appendix N). For the Wash transect, however, most of the common species from each study were not found to be common by the other study. These comparisons may reflect actual changes in the bird community, be an artifact of different sampling intensities, or could simply be due to chance. However, Botteri's sparrow and other species recorded on the Wash transect that were not recorded by Russell and Danforth (1979), such as western bluebird and dark-eyed junco, are considered grassland species. The decrease in grazing in 1991 and the subsequent increase in grassy plant species may be the reason for these differences.

#### **Monitoring**

The bird inventory of Coronado NM is close to completion, though new species will likely continue to be found. We applaud the effort by memorial volunteers to maintain the observation database and suggest that they continue their effort. Perhaps the most important research-related activity that the park can undertake is to monitor the distribution, abundance, and species richness of birds in the memorial. To this end it appears that the Sonoran Desert Network will include landbirds in the core "vital signs" monitoring program (Powell et al. 2006). This program will provide managers with information on the changes to the bird community, and because other parameters will be monitored (e.g., vegetation and climate) this information may provide information about potential causes to the changes observed in the bird community.



## Chapter 6: Mammal Inventory

Don E. Swann, Cecil R. Schwalbe, Amy J. Kuenzi, Melanie Bucci, and Barbara N. Alberti

There have been few past studies of mammals at Coronado NM, though one notable exception is the study of coatis described in the popular book *Chulo* by Bill Gilbert (1973). Prior to 2000, most knowledge of mammals was compiled during a study by University of Arizona researchers in 1977–1978. Using trapping for nocturnal rodents, mist-netting for bats, and observations, Petryszyn and Cockrum (1979) confirmed 26 species of mammals (including 12 bat species) at the memorial. However, they were unable to confirm an additional 31 mammals (including 17 bat species) that they believed were probably present based on range maps and historic records. Petryszyn and Cockrum (1979) believed that the most significant gaps in knowledge of mammals at the memorial were of grassland species of nocturnal rodents, and most of the larger species they observed were not confirmed with voucher specimens or photographs. They also noted nine species that may have been present in the past but appeared to be at least locally extirpated, including such federally-listed species as the Mexican gray wolf, black-tailed prairie dog, jaguarundi, ocelot, and jaguar.

### Objectives

The purpose of the present study was to fill in gaps in our knowledge of terrestrial mammals at Coronado NM, particularly in grassland habitats, and to combine this information with historic data to produce a baseline inventory. Our primary goal was to use a variety of techniques to confirm as many of the memorial's terrestrial mammal species as possible. Our secondary goal was to fit these data into a geographic context, and produce range maps for the current distribution of mammals at the memorial. Although monitoring population abundances was not a goal of this study, we did measure relative abundance of nocturnal rodents. Specific objectives were:

1. To survey terrestrial mammals using techniques that have minimal impact on animals and the environment;
2. To use voucher photographs as a method of confirming easily identified species present on the memorial, and to collect as voucher specimens animals found dead or species difficult to identify from photographs;
3. To gather historic information on mammals at Coronado NM from published sources, museum collections, and historic reports;
4. To document relative abundance of selected species in different vegetation communities in the memorial;
5. To provide this information to the memorial in a format useful for management decisions and interpretation.

Two products are provided in the appendices of this report. Appendix D is a table of all species confirmed during this study and the Petryszyn and Cockrum (1979) study. Appendix F is a table of species that may be found at the memorial. For accounts of all confirmed and potential species, including historic information and current range maps see Swann et al. (2000) appendices C and D. Copies of field data sheets, database files, Global Positioning Systems (GPS) files, Trailmaster photographs, field notes, and voucher photographs were given to Coronado NM.

### Methods

Detecting presence and absence of mammals can be difficult due to their diverse lifestyles, including nocturnal and underground habits. In addition, small mammals may be very specific in their microhabitat requirements, while larger species, especially carnivores, may occur naturally at very low population densities. To detect as many species that occur in the memorial as possible, we used a wide variety of techniques as outlined below.

#### *Historic Records*

To gather information about the present and historic distribution of mammals at the memorial, we surveyed sources inside and outside the NPS system. For all mammal species of possible occurrence, we reviewed published records,

particularly Hoffmeister (1986), the primary source of such data for Arizona, and unpublished sources, particularly Petryszyn and Cockrum (1979). We also reviewed records from the University of Arizona mammal museum and other major U.S. museums. Records from wildlife observation cards at Coronado NM through November 2000 were summarized by volunteers from the memorial and Southern Arizona Office (SOAR). Additional historic records were provided by the memorial, the NPS Western Archaeological Conservation Center in Tucson, and the Heritage Database of the Arizona Game and Fish Department (AGFD 1996).

### *Small Mammal Trapping*

To verify species of nocturnal rodents and to determine species distribution and relative abundance, we trapped nocturnal rodents using “extra large” Sherman and Tomahawk brand live traps for small mammals (7.5 X 9 X 23.5 cm). Traps were baited with oatmeal mixed with a small amount of peanut butter (approximately one tbsp. peanut butter per 0.5 kg of oatmeal). All traps were covered by vegetation and/or soil for insulation, and a handful of polyester batting was placed inside for bedding. Traps were baited in late afternoon and checked early the following morning. Traps were usually closed during the day, except during cool weather in thick grassland and riparian areas.

Standard inventory grids consisted of 25 traps placed ten meters apart, usually arranged in a square grid of five rows (A–E) running north/south and five rows (1–5) running east/west; grids in riparian areas were arranged to conform to the riparian corridor. A few smaller arrays of five-ten traps were also set in special microhabitats and human-use areas. Each trap station was marked with surveyor’s tape. All traps were removed from grids at the end of each trapping session of one to four nights, and the southeast trap station (A–1) was flagged for later relocation for vegetation analysis and UTM coordinates. We did not attempt to randomly locate grids, but located them in areas that represented the geographic, topographic, and vegetative diversity of the memorial. These included burned and unburned oak woodland areas, wet seeps, cattle tanks, riparian corridors, high- and low-elevation grasslands, grazed and ungrazed semi-desert grassland areas, and areas altered by human activity.

Each captured animal was identified to species, age, sex, and reproductive condition, and the following measurements were taken: weight in grams; right hindfoot length, ear length, tail length, and body length in millimeters. Individuals were batch-marked with a colored permanent marker, with each color representing a unique day. Where animals were difficult to identify to species, we noted special characteristics (such as foot tubercle color in pocket mice, tail stripe width in harvest mice, etc.). Specimens or photographs of difficult-to-identify mammals were brought to the University of Arizona Mammal Collection and confirmed by Dr. Yar Petryszyn, Assistant Curator. All trapping data were entered into a database (Microsoft Access 97), and brought into the Geographic Information Systems (GIS) in Program ArcView (version 3.0) to generate range maps for each species.

### *Infrared-Triggered Photography*

Infrared-triggered photographs of large- and medium-sized mammals were obtained using the model 1500 Trailmaster camera system (Goodson and Associates, Inc., Lenexa, KS), where a single infrared beam is emitted by a transmitter and detected by a receiver; a photograph is taken when this beam is broken by an animal. Three units were operated continuously from October 1996 through December 1997. We did not randomly locate camera units, but placed them in vegetated areas that represented the geographic diversity of the memorial and were protected from direct sunlight and observation by visitors. Cameras were set out for intervals of two weeks or more at a natural water source, or baited with sardines, cat or other carnivore lure, a visual lure, or some combination of these. We recorded all changes of film and bait used, and map coordinates were obtained using GPS. We also recorded slope, aspect, and vegetation at each site. Animals in each photograph were identified to species if possible, and times and dates recorded; data were entered into a Microsoft Access database as for nocturnal rodents.

### *Observations*

To supplement records of mammal distribution and relative abundance, we recorded all mammals observed during this study, and during field trips associated with a study of reptiles and amphibians

which was initiated at the memorial in spring 1997. We looked for mammals while driving on a road transect between the east and west boundaries; while conducting trapping and habitat analysis; and under boards and other materials while searching for reptiles. We recorded date, time, species, and location associated with each animal observed. Observations and specimens were also collected by memorial staff through 2003.

### *Voucher Specimens and Photographs*

One specimen of each small mammal species not previously captured at the memorial, and all trap mortalities, were taken as vouchers. Live specimens were euthanized with carbon dioxide gas according to University of Arizona animal handling protocols (Institutional Animal Care and Use Committee (IACUC), Control #94-067). Voucher specimens of medium and large mammals usually were animals killed by cars on East Montezuma Canyon Road, but occasionally parts of animals (e.g., skulls or carcasses) found dead were also collected. All specimens were deposited in the University of Arizona mammal collection. In addition to photographs taken by infrared-triggered cameras, we also voucher-photographed live individuals of rodents and a desert shrew.

### *Monitoring Grids*

As a supplement to the small mammal inventory, two grids of 100 traps each were selected as test grids for monitoring abundance. The purpose of this monitoring was to evaluate the sample size and cost necessary for monitoring nocturnal rodents at the memorial. One grid (9A18) was randomly located in semi-desert grassland below 1,524 m; a second grid (2A6) was randomly located on a south-facing slope in oak savannah above 1,921 m, within 1.6 km of a road. Methods were the same as described for inventory, except individual animals were uniquely marked using permanent color pens in order to develop a capture history for each. Grids were trapped for 4–6 nights each year from 1997–2003 during October, November, or early December.

### *Determining Relative Abundance*

Data from small mammal trapping was used to provide an index of abundance for each species listed in Appendix P and Q. Comparing the abundance of one species relative to another is

problematic, because the number of individuals observed reflects not only the numbers of animals present, but also how easily they can be captured. For example, pocket gophers are certainly very common at the memorial, but are rarely observed directly because they spend so much of their time underground and are difficult to trap alive. Abundance comparisons should ideally be based on estimates of absolute abundance or density (number of individuals per unit area), obtained through unbiased methods, such as mark-recapture studies. However, these methods are extremely time-consuming, and usually cannot be obtained without intensive, species-specific study.

### *Nomenclature and Field Schedule*

In this report we follow the standard English and scientific nomenclature of Jones et al. (1997). Field work for this study began in September 1996 and continued through February 1998; however, with activities taking place in every month, but primarily in spring and fall. Monitoring of rodents continued each fall through 2003 (Swann et al. 2002). C. R. Schwalbe was the principle investigator. D. E. Swann conducted the majority of fieldwork, with primary assistance from A. J. Kuenzi, B. N. Alberti, M. Bucci, S. Wolf, and a number of volunteers.

## **Results**

Twenty-nine species of native terrestrial mammals were confirmed during this study by voucher specimens; an additional nine species not confirmed by specimens were confirmed by unambiguous voucher photographs. We did not collect specimens of three additional species that had previously been collected by Petryszyn and Cockrum (1979); however, except for one species of pocket gopher and all bats, we observed all of the species confirmed by Petryszyn and Cockrum (1979). An additional three species of mammals (black-tailed jackrabbit, feral cat, and mule deer) were confirmed by reliable sightings by us or memorial staff. The total number of mammal species confirmed for Coronado NM is 52 (Appendix D). Not included in this total are domestic cattle, but we do include two non-native species (feral dog and feral cat) that exist in the wild but probably would not survive without human assistance, and one non-native (house

mouse) that is probably established in the wild. Sightings were considered reliable only for species that are difficult to misidentify (e.g., black-tailed jackrabbit) and that were observed within their expected range and habitat. We also do not include eastern cottontail because a voucher specimen is needed to unambiguously identify this species; however, we are fairly certain that we observed and photographed this species and believe it will eventually be verified.

Of the 43 confirmed species, at least 12 may be considered “new”, because to our knowledge no voucher specimens or photographs had been taken in the past, and the species does not appear on previous species lists. For all but 12 species observed or captured during our study, voucher photographs were obtained. Slides of these species have been deposited in archives at Coronado NM and copies retained by CRS and DES.

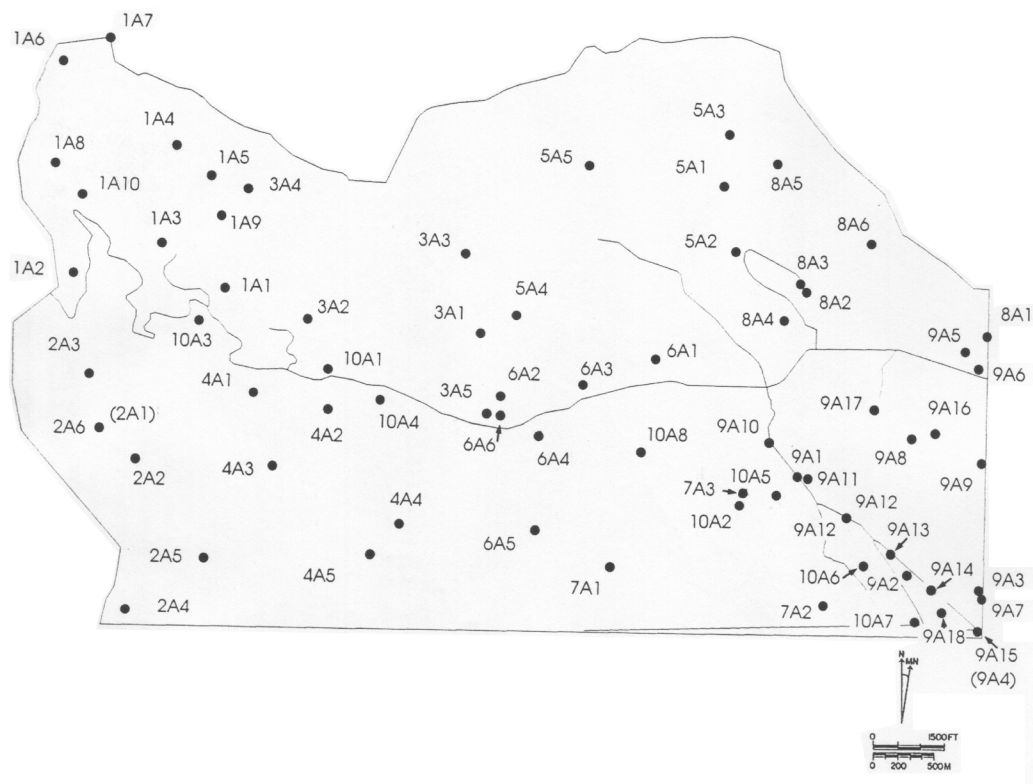
### *Small Mammal Trapping*

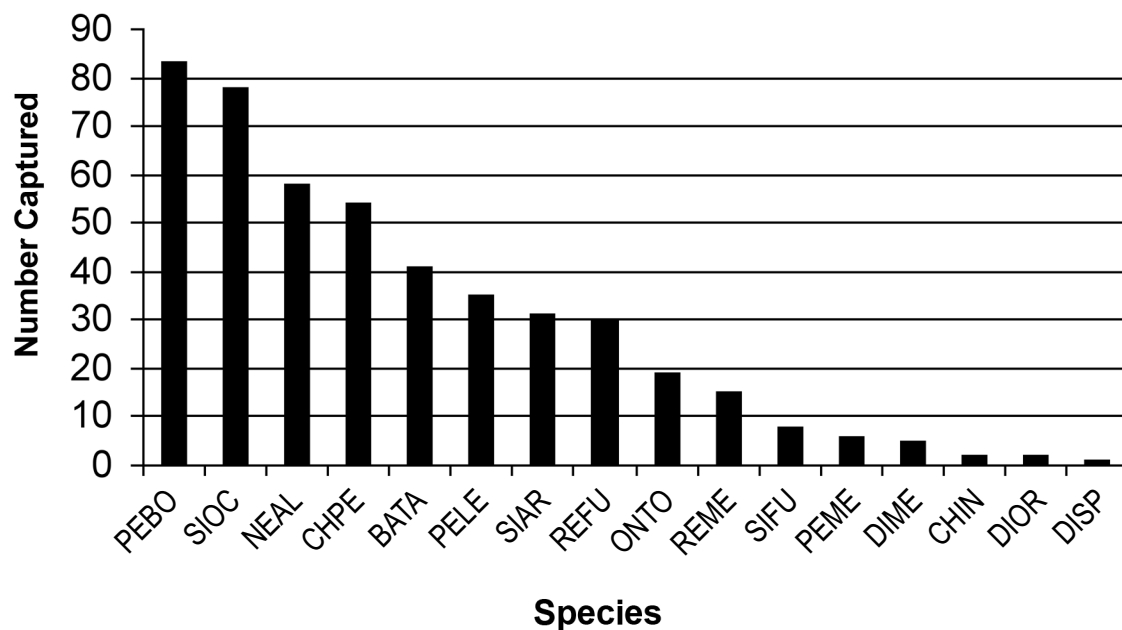
We trapped nocturnal rodents on inventory grids from September 1996 through December 1997 for

a total of 5,424 trap-nights. Locations of trapping grids are shown in Figure 6.1, and vegetation type and elevation for each grid are summarized in Swann et al. (2000) Appendix B. Photographs of each grid and data sheets with vegetation data have been provided to Coronado NM.

A total of 672 captures of 17 species were made, for a mean trap success of 12.4%. A total of 498 individuals were captured. The most common species captured (Figure 6.2) were the brush mouse, yellow-nosed cotton rat, white-throated woodrat, and Sonoran Desert pocket mouse.

A few species were very limited in their distribution in the memorial. The deer mouse was only captured at the “blue waterfall” seep north of the residences and two other locations. Merriam’s kangaroo rat and banner-tailed kangaroo rat were only captured near the east boundary fence approximately 2.0 km south of East Montezuma Canyon Road, and Ord’s kangaroo rat was only trapped at one location in the orchard area of Montezuma Ranch. The rock pocket mouse was captured rarely in rocky areas only. The tawny-bellied cotton rat and southern grasshopper mouse





**Figure 6.2. Number of individuals trapped (captures – recaptures) of nocturnal rodent species captured at Coronado NM, 1996–1997.** Based on 5,424 trap-nights. Acronyms are based on scientific names: first two letters of genus name, followed by first two letters of species name. See Appendix D for scientific names.

were only captured in semi-desert grasslands below 1,509 m. During monitoring from 1997–2003, we did slightly increase the number of locations for several species (Figure 6.1 and Appendix C).

Six species that were largely confined to grassy areas occurred in both lower-elevation semi-desert grassland as well as upper-elevation (above 1,982 m) oak savannah where grasses were abundant. These species included the Sonoran Desert pocket mouse, yellow-nosed cotton rat, Arizona cotton rat, fulvous harvest mouse, western harvest mouse, and northern pygmy mouse. Three species were nearly ubiquitous: the white-footed mouse was found at all elevations and vegetation associations except semi-desert grassland, and the brush mouse and white-throated woodrat were found in every vegetation association. Table 6.1 lists the number of each species captured in each of the four major vegetation communities plus residential areas and seeps, with additional categories of grazed vs. ungrazed, and tree cover > grass cover vs. tree cover < grass cover.

#### *Monitoring Grids*

On the 100-trap oak savannah grid near Coronado Peak, 405 individuals of 11 species were captured

during 1997–2003. On the lower grassland grid just north of Montezuma Canyon near the east boundary, 470 individuals of 14 species were captured. The number of individuals and species trapped on the two grids varied greatly among years (Appendices P, Q). For example, on the grassland grid the number of captured individuals ranged from a low of 27 in 1997 to a high of 119 in 2001. Appendices P and Q are summaries of raw capture data on the monitoring grids. Over seven years of effort we confirmed three species that had not been previously confirmed at the memorial (spotted ground squirrel, silky pocket mouse, and house mouse). In addition, we made additional captures of species in areas where they had not been previously trapped, including capture of several “grassland” species (such as pygmy mouse) on the oak savannah grid.

#### *Infrared-Triggered Photography*

Between September 1996 and December 1997, three Trailmaster cameras were used for approximately 1,142 nights at 25 locations (Figure 6.3). Cameras were often not operational because a roll of film had been completely exposed or an equipment malfunction had occurred; we estimate

**Table 6.1. Number of nocturnal rodents trapped per 100 trap-nights in selected vegetation types at Coronado NM.**

Vegetation types: 1 = in and around buildings, 2 = semi-desert grassland (grazed), 3 = mesquite riparian, 4 = oak riparian with less than 50% tree cover, 5 = oak riparian with more than 50% tree cover, 6 = oak woodland with less than 50% tree cover, 7 = oak woodland with more than 50% tree cover, 8 = seeps with permanent water or moisture, 9 = semi-desert grassland (ungrazed).

Species	Vegetation Type								
	1	2	3	4	5	6	7	8	9
Sonoran Desert pocket mouse	1.6	1.5	3.5	1.9	0.9	0.3	0.0	0.0	1.5
rock pocket mouse	0.0	0.2	0.0	0.0	0.0	0.0	0.1	0.0	1.1
hispid pocket mouse	0.0	0.2	1.0	0.0	0.0	0.1	0.0	0.0	0.0
banner-tailed kangaroo rat	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.1
Ord's kangaroo rat	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.1
Merriam's kangaroo rat	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.3
western harvest mouse	0.0	0.0	0.0	0.0	0.0	0.3	0.0	0.0	0.6
fulvous harvest mouse	1.6	0.0	2.0	0.0	0.0	0.8	0.0	0.6	0.5
deer mouse	0.0	0.0	0.0	0.0	0.1	0.0	0.0	2.3	0.0
white-footed mouse	0.0	0.0	0.2	0.0	0.9	1.4	0.7	2.3	0.0
brush mouse	1.6	0.5	0.8	1.9	0.4	2.5	2.7	2.8	0.1
northern pygmy mouse	0.0	0.0	3.2	0.0	0.0	0.2	0.0	0.0	0.2
southern grasshopper mouse	0.0	0.2	0.2	0.0	0.0	0.0	0.0	0.0	1.0
white-throated woodrat	4.2	0.2	1.1	1.9	1.3	2.0	0.7	2.8	0.0
tawny-bellied cotton rat	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.5
yellow-nosed cotton rat	1.6	0.0	0.3	1.9	0.0	2.6	0.1	0.6	1.1
Arizona cotton rat	0.8	0.0	1.0	0.0	0.0	0.1	0.0	2.3	1.1

the cameras were operational for a total of approximately 640 nights. During this period, 379 photographs of 18 native mammal species (and one photograph of a feral dog) were obtained (Figure 6.4). For the majority of species, these photographs were the first voucher photographs obtained for the memorial. In addition, we obtained photographs of great horned owls, red-tailed hawks, and greater roadrunners. Summary data for Trailmaster cameras is contained in Appendix O.

### Inventory Completeness

We believe that our inventory was fairly complete, given the large amount of time we spent using infrared-triggered cameras and trapping small mammals. We recorded 46 species of terrestrial mammals for the memorial, and strongly suspect that eastern cottontail occurs, even though we could not confirm this species. We included on the list the only species of terrestrial mammal confirmed by Petryszyn and Cockrum (1979) that we did not see, the Southern pocket gopher, because we did record pocket gopher sign in appropriate habitat for this common species. In

addition to confirmed native and non-native species, an additional 17 native species either occur or have occurred historically within the Huachuca Mountains (Hoffmeister 1986) or nearby valleys (Appendix F). These include a few species that have been sighted or confirmed very close to the memorial and are likely to occur (e.g., common porcupine); species that occur nearby but are unlikely due to lack of suitable habitat (e.g., round-tailed ground squirrel and Bailey's pocket mouse); species that are certainly not resident but range widely and may pass through the memorial from time to time (jaguar and ocelot); and a few species that are now certainly extirpated in the area (gray wolf and grizzly bear). Detailed species accounts for all known and potential species, including summaries of historic and museum records, are provided in Appendix C of Swann et al. (2000).

### Discussion

#### *Species Diversity*

Results of our inventory indicate that Coronado NM has a great diversity of terrestrial mammal



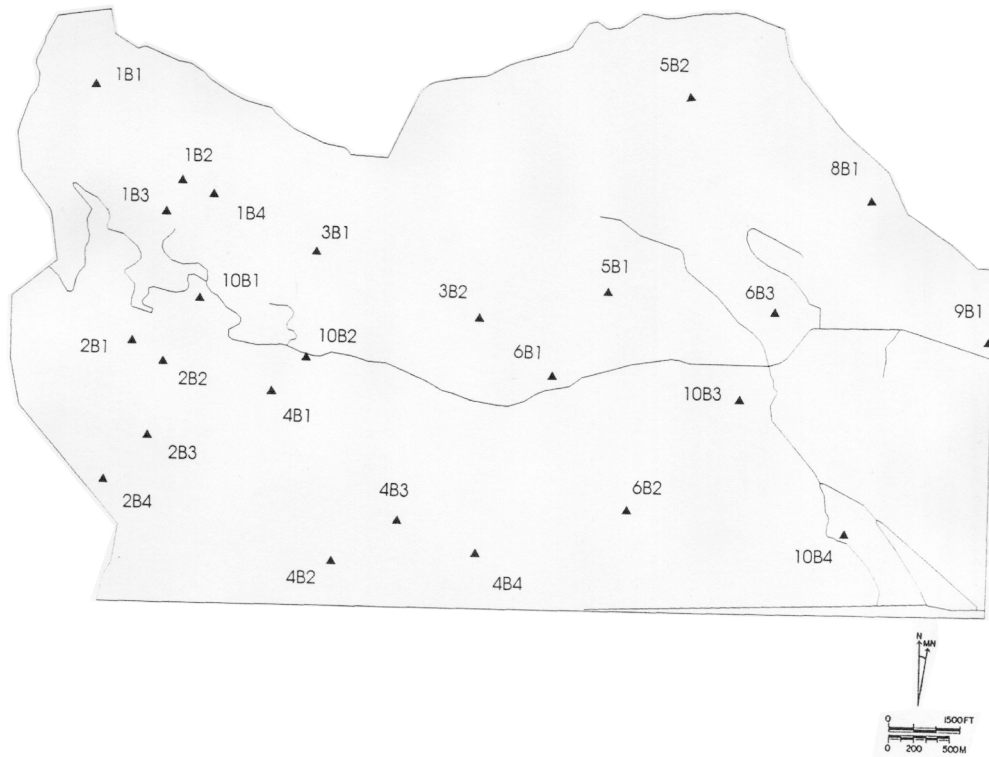


Figure 6.3. Map of Coronado NM, showing locations of infrared-triggered photograph stations.

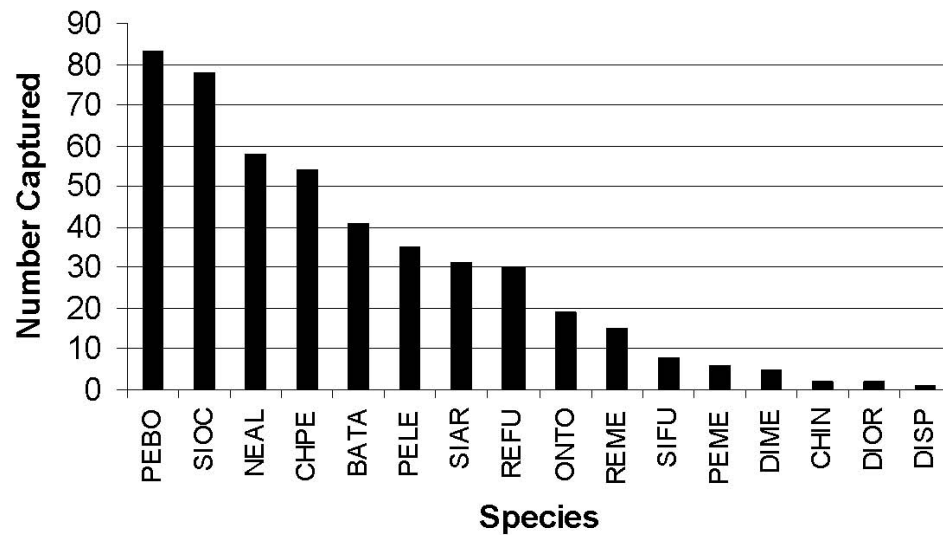


Figure 6.4. Number of individual photographs of 18 mammal species taken by infrared-triggered photography at Coronado NM, 1996–1997. Based on approximately 1,142 camera-nights. Acronyms are based on genus and species names, as in Appendix D.

species for its size compared to other parks in southern Arizona (although not as many as Tonto National Monument; Table 6.2). This is probably due to a number of factors, including the presence of thick grassland vegetation in ungrazed areas, which supports many species of rodents; the memorial's location in the Huachuca Mountains, which is itself very diverse for biogeographic reasons; and the memorial's connectivity to other natural areas nearby, including Coronado National Forest, the San Pedro River, and undeveloped areas in Mexico. In addition, small parks such as Coronado NM and Tonto NM may have higher diversity per unit area because their mammalian fauna include most of the habitat generalists (e.g., species like desert cottontail, mountain lion, and white-throated woodrat) that occur in larger parks. Preserving this diversity at the memorial is important, and some effort should be made to better understand why it occurs.

It is significant that this study confirmed a number of rodent species not confirmed by Petryszyn and Cockrum (1979) (Appendix D). Their study was shorter in duration than ours and involved far fewer trap-nights; we trapped a number of species in areas where they did not detect them. Indeed, the yellow-nosed cotton rat, currently the most abundant rodent in both higher- and lower-elevation grasslands, was not trapped by Petryszyn and Cockrum (1979).

The most plausible explanation for the difference between our results and those of the previous study is that grasses in the low-elevation grasslands south of East Montezuma Canyon Road, which were very robust during our study, were sparse in 1977–78. Much of this area was bare ground at that time (Y. Petryszyn, pers. comm.), probably due to the combined results of heavy cattle grazing and low rainfall. The beginning of our study coincided with the end of a very wet summer monsoon season, and no grazing had

occurred in this area for over eight years (E. Lopez, pers. comm.).

The other major change that has occurred at the memorial since 1978 has been the loss of trees (particularly conifers) and the resultant growth of high-elevation grasses since the severe Peak Fire of June 1988 (Ruffner and Johnson 1991). This vegetative change may also be related to the cessation of grazing at higher elevations. The resulting increase in grass seed crop has clearly been favorable to small rodents. It will be interesting to track changes in the species diversity of these oak savannahs if they become revegetated with oaks and pinyon pine.

As a small but significant block of protected land at the south end of the Huachuca Mountains, the memorial undoubtedly plays an important role as a refugium for many mammals, including not only grassland species, but also hunted animals such as deer (*Odocoileus* spp.) and predators such as mountain lions. Because of this, the memorial may also play a significant role in any return of species that are presently extirpated from the area. Black-tailed prairie dogs, which probably occurred in or nearby the memorial earlier in this century, apparently continue to persist over the border in Mexico less than 4.8 km south of the Huachuca Mountains (Ecological Center of Sonora 1994). It is not impossible that this species could naturally recolonize protected grassland areas of the memorial at some time in the future; there was recently an unconfirmed sighting of this species by a visiting ranger (Barbara Alberti, pers. comm.). Some scientists and ranchers now believe that prairie dogs are beneficial for cattle because of their role in suppressing mesquite (Weltzin 1997).

In addition, the memorial provides a potential refugium for jaguars, ocelots, and gray wolves moving northward, should these species increase in number. However, for most large

**Table 6.2. Number of species of native terrestrial mammals on species lists at four parks in southern Arizona, and density in number of species per 100 ha.**

Park unit	Area (Ha)	Number of mammals	Density
Organ Pipe Cactus NM	133,830	50	0.037
Saguaro NP	36,531	49	0.134
Coronado NM	1,900	43	2.263
Tonto NM	461	30	6.508

mammals, the memorial does not contain sufficient area to contain the entire home range for even a small population. If development and loss of habitat in the San Pedro Valley continues, it is very possible that more species will disappear from the memorial, as has been documented for other western national parks (Newmark 1995). Species with relatively large home ranges at the memorial include black bear, mountain lion, bobcat, white-nosed coati, and others.

## Monitoring

Monitoring vertebrate animals in national parks is essential if the NPS is to fulfill the important mission of preserving biodiversity on its lands that is presently mandated by the agency (NPS 1992); however, monitoring of vertebrates can be time-consuming and expensive. Many mammal species are difficult to observe and count, and their populations often fluctuate greatly due to natural causes. Monitoring is particularly difficult in small park areas where human and financial resources may be even more limited than in larger parks.

Although a number of monitoring programs for mammals have been developed in national parks in recent years (e.g., at Channel Islands National Park and Organ Pipe Cactus National Monument), such programs may be unrealistic for small park areas such as Coronado NM because of the cost involved (Swann 1999). In addition, many small park monitoring programs emphasize estimating population size of either threatened species such as Florida panthers, or common species such as deer mice, but are not concerned with monitoring other species. We believe that knowledge of changes in abundance of selected species must be coupled with knowledge of the presence and distribution of all species using the park. Loss of species from national parks and other natural areas due to human impacts is a major concern, yet one that few monitoring programs are designed to measure. In addition to tracking potential changes in abundance of common species, monitoring must provide information on species that may be in danger of extirpation because of their rarity or loss of specialized habitat.

For this reason, we provide information for conducting a multi-level approach to monitoring mammals at Coronado NM and suggest that it is in the best interest of management that park-wide

inventories of mammal species be conducted every five to ten years. Such inventories should take a systematic approach to sampling all species that occur in the memorial, and include trapping and use of infrared-triggered cameras or similar technology in all vegetation communities. By providing documentation of locations and effort associated with our study, we envision that all or portions of this effort could be replicated in future surveys.

Secondly, if additional monitoring activities can be scheduled, important information can be gained by small mammal trapping that is conducted on monitoring grids randomly located in selected habitat types. We established one grid each in oak savannah and in semi-desert grassland areas; additional grids could be established in Montezuma Canyon and in other areas. To understand the natural variability associated with different rodent species, which can be extreme, these grids should be trapped several times each year for four to eight nights each, for at least several years; later, it may be possible to reduce the frequency of trapping occasions. Program MONITOR (Gibbs 1995) enables researchers to determine, based on measured means and variances, the number of trap grids and trapping occasions necessary to detect a predetermined change in abundance (such as 3% annually) over a period of time. Although our study of two years was not adequate to determine the natural variation in abundance of rodent species, results from other southern Arizona studies (e.g., Petryszyn 1982) suggest variance may be extreme. For example, the estimated abundance of cotton rats on monitoring Grid 2A6 has ranged from 21 individuals in 1997 to five in 1999 (Bucci et al. 1999). High natural variability increases the number of grids that must be sampled in order to detect significant levels of change.

Our estimated costs for annual monitoring of large grids (\$800) should be considered a minimum estimate. Monitoring these grids in 1998 and 1999 was more expensive (Bucci et al. 1999) because we trapped for a longer period. The greater number of trap-nights provided more accurate population estimates, and the higher costs were at least partly off-set by a large volunteer effort.

Further valuable information is to be gained by monitoring selected mammal species intensively, measuring abundance as well as parameters such as reproduction and survival.

Determining which species are studied will depend upon financial resources and researcher interest. Long-term studies of species such as ringtails, skunks, or pygmy mice are extremely rare, and would benefit not only Coronado NM but also other wildlife conservation agencies. Species such as coatis and mountain lions are studied more frequently, but are of interest because of their

elusiveness, limited distribution, and public appeal. Long-term studies of single species (e.g., Turner 1991) have given tremendous insight into their natural history and endangerment factors. Any such studies at the memorial would significantly improve our ability to preserve the mammalian diversity of the memorial for the future.

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**Appendix A. Plant species observed at Coronado NM during our vegetation sampling at breeding-season bird stations in 2004.** Species observed or collected at Coronado NM based on: Ruffner and Johnson (R&J; 1991), Parfitt and Christy (P&C; 1992), Halvorson and Guertin (H&G; 2003), Ruyle (Ruy; 2002), NPS (unpublished data), and specimens in herbaria at Arizona State University (ASU), Desert Botanical Gardens (DBG), Northern Arizona University (NAU) and the University of Arizona (UA). Species in bold-faced type are non-native.

Family	Scientific name	Common name	Studies						Specimens in herbaria			
			R&J	P&C	H&G	Ruy	NPS	UA	ASU	DBG	NAU	UA
Acanthaceae												
	<i>Dyschoriste decumbens</i> (Gray) Kuntze	spreading snakeherb	X	X								
Agavaceae												
	<i>Agave palmeri</i> Engelm.	Palmer's century plant	X	X		X	X	X		X		
	<i>Agave parryi</i> Engelm. var. <i>parryi</i>	Parry's agave	X	X								
	<i>Agave schottii</i> Engelm. var. <i>schottii</i>	Schott's century plant	X	X								
	<i>Yucca schottii</i> Engelm.	Schott's yucca	X	X			X	X		X		
Amaranthaceae												
	<i>Amaranthus albus</i> L.	prostrate pigweed		X								
	<i>Amaranthus arenicola</i> I.M. Johnston	sandhill amaranth	X	X								
	<i>Amaranthus hybridus</i> L.	slim amaranth	X	X								
	<i>Amaranthus palmeri</i> S. Wats.	carelessweed		X								
	<i>Amaranthus powellii</i> S. Wats.	Powell's amaranth										X
	<i>Froelichia arizonica</i> Thomb. ex Standl.	Arizona snakecotton	X	X								
	<i>Gomphrena caespitosa</i> Torr.	tufted globe amaranth	X	X								X
	<i>Gomphrena nitida</i> Rothrock	pearly globe amaranth	X	X								
	<i>Gomphrena sonora</i> Torr.	Sonoran globe amaranth	X	X								
	<i>Guilleminea densa</i> (Humb. & Bonpl. ex Willd.) Moq.	small matweed	X	X								
	<i>Iresine heterophylla</i> Standl.	Standley's bloodleaf		X								
Anacardiaceae												
	<i>Rhus aromatica</i> Ait.	fragrant sumac					X					
	<i>Rhus glabra</i> L.	smooth sumac	X	X								
	<i>Rhus trilobata</i> Nutt.	skunkbush sumac					X	X			X	
	<i>Rhus trilobata</i> var. <i>racemulosa</i> (Greene) Barkl.	skunkbush sumac	X	X								
	<i>Rhus virens</i> Lindheimer ex Gray	evergreen sumac					X					
	<i>Rhus virens</i> var. <i>choriophylla</i> (Woot. & Standl.) L. Benson	evergreen sumac	X	X			X	X			X	X
	<i>Toxicodendron radicans</i> (L.) Kuntze	eastern poison ivy	X	X								
	<i>Toxicodendron radicans</i> ssp. <i>radicans</i> (L.) Kuntze	eastern poison ivy						X				
Apiaceae												
	<i>Spermolepis echinata</i> (Nutt. ex DC.) Heller	bristly scaleseed		X						X		X
	<i>Yabea microcarpa</i> (Hook. & Arn.) K.-Pol.	false carrot		X								
Apocynaceae												
	<i>Macrosiphonia brachysiphon</i> (Torr.) Gray	Huachuca Mountain rocktrumpet	X	X								
Araliaceae												
	<i>Aralia humilis</i> Cav.	Arizona spikenard		X								
Asclepiadaceae												
	<i>Asclepias asperula</i> (Dcne.) Woods. ssp. <i>asperula</i>	spider milkweed		X						X		

Family	Scientific name	Common name	Studies						Specimens in herbaria			
			R&J	P&C	H&G	Ruy	NPS	UA	ASU	DBG	NAU	UA
Asclepiadaceae												
	<i>Asclepias engelmanniana</i> Woods.	Engelmann's milkweed	X	X								
	<i>Asclepias glaucescens</i> Kunth	nodding milkweed	X	X								
	<i>Asclepias linaria</i> Cav.	pineneedle milkweed		X								
	<i>Asclepias macrotis</i> Torr.	longhood milkweed	X	X								
	<i>Asclepias nummularia</i> Torr.	tufted milkweed		X								
	<i>Asclepias nyctaginifolia</i> Gray	Mojave milkweed		X						X		
	<i>Funastrum crispum</i> (Benth.) Schlechter	wavyleaf twinevine		X						X		
	<i>Funastrum cynanchoides</i> ssp. <i>heterophyllum</i> (Vail) Kartesz, comb. nov. ined.	Hartweg's twinevine	X	X						X		
Asteraceae												
	<i>Acourtia thurberi</i> (Gray) Reveal & King	Thurber's desertpeony		X							X	X
	<i>Ageratina herbacea</i> (Gray) King & H.E. Robins.	fragrant snakeroot	X	X				X		X		
	<i>Ageratina paupercula</i> (Gray) King & H.E. Robins.	Santa Rita snakeroot		X								
	<i>Ambrosia confertiflora</i> DC.	weakleaf burr ragweed		X		X				X		
	<i>Ambrosia psilostachya</i> DC.	Cuman ragweed	X							X		
	<i>Artemisia ludoviciana</i> Nutt.	white sagebrush		X							X	
	<i>Baccharis bigelovii</i> Gray	Bigelow's false willow	X	X								
	<i>Baccharis neglecta</i> Britt.	Rooseveltweed	X	X								
	<i>Baccharis pteronioides</i> DC.	yerba de pasmo	X	X		X	X	X				
	<i>Baccharis salicifolia</i> (Ruiz & Pavón) Pers.	mule's fat	X	X								
	<i>Baccharis sarothroides</i> Gray	desertbroom		X		X	X	X				
	<i>Baccharis thesioides</i> Kunth	Arizona baccharis		X				X			X	
	<i>Bahia absinthifolia</i> Benth.	hairyseed bahia	X	X						X		
	<i>Bahia dissecta</i> (Gray) Britt.	ragleaf bahia		X								
	<i>Baileya multiradiata</i> Harvey & Gray ex Gray	desert marigold	X	X								
	<i>Bidens aurea</i> (Ait.) Sherff	Arizona beggarticks								X		
	<i>Bidens bigelovii</i> Gray	Bigelow's beggarticks		X						X		
	<i>Bidens leptcephala</i> Sherff	fewflower beggarticks		X								
	<i>Brickellia baccharidea</i> Gray	resinleaf brickellbush	X	X								
	<i>Brickellia betonicifolia</i> Gray	betonyleaf brickellbush								X	X	
	<i>Brickellia californica</i> (Torr. & Gray) Gray	California brickellbush		X				X				X
	<i>Brickellia californica</i> (Torr. & Gray) Gray var. <i>californica</i>	California brickellbush								X	X	
	<i>Brickellia eupatorioides</i> var. <i>chlorolepis</i> (Woot. & Standl.) B.L. Turner	false boneset	X	X						X	X	
	<i>Brickellia eupatorioides</i> (L.) Shinners var. <i>eupatorioides</i>	false boneset	X									
	<i>Brickellia grandiflora</i> (Hook.) Nutt.	tasselflower brickellbush	X	X								
	<i>Brickellia lemmonii</i> Gray	Lemmon's brickellbush		X				X				
	<i>Brickellia simplex</i> Gray	Sonoran brickellbush	X	X								
	<i>Brickellia venosa</i> (Woot. & Standl.) B.L. Robins.	veiny brickellbush		X						X	X	
	<i>Carminatia tenuiflora</i> DC.	plumeweed	X	X							X	
	<i>Carphochaete bigelovii</i> Gray	Bigelow's bristlehead		X								X
	<i>Chaetopappa ericoides</i> (Torr.) Nesom	rose heath	X	X							X	

Family	Scientific name	Common name	Studies						Specimens in herbaria			
			R&J	P&C	H&G	Ruy	NPS	UA	ASU	DBG	NAU	UA
Asteraceae												
	<i>Cirsium arizonicum</i> (Gray) Petrak	Arizona thistle	X	X						X		
	<i>Cirsium neomexicanum</i> Gray	New Mexico thistle	X	X								
	<i>Conyza canadensis</i> (L.) Cronq.	Canadian horseweed	X	X							X	
	<i>Cosmos parviflorus</i> (Jacq.) Pers.	southwestern cosmos	X	X					X	X		
	<i>Ericameria laricifolia</i> (Gray) Shinnery	turpentine bush	X	X			X					
	<i>Ericameria nauseosa</i> (Pallas ex Pursh) Nesom & Baird	rubber rabbitbrush				X						
	<i>Ericameria nauseosa</i> var. <i>nauseosa</i> (Pallas ex Pursh) Nesom & Baird	rubber rabbitbrush	X	X				X				
	<i>Erigeron concinnus</i> (Hook. & Arn.) Torr. & Gray	Navajo fleabane		X								
	<i>Erigeron divergens</i> Torr. & Gray	spreading fleabane	X	X		X					X	
	<i>Erigeron flagellaris</i> Gray	trailing fleabane		X								
	<i>Erigeron neomexicanus</i> Gray	New Mexico fleabane	X	X						X		
	<i>Erigeron oreophilus</i> Greenm.	chaparral fleabane		X								
	<i>Filago californica</i> Nutt.	California cottonrose		X						X		X
	<i>Fleischmannia pycnocephala</i> (Less.) King & H.E. Robins.	lavender thoroughwort		X								
	<i>Gamochaeta purpurea</i> (L.) Cabrera	spoonleaf purple everlasting		X								
	<i>Gnaphalium</i> L.	cudweed				X				X		
	<i>Guardiola platyphylla</i> Gray	Apache plant	X	X						X	X	
	<i>Gutierrezia microcephala</i> (DC.) Gray	threadleaf snakeweed	X	X			X					
	<i>Gutierrezia sarothrae</i> (Pursh) Britt. & Rusby	broom snakeweed		X		X	X					
	<i>Gutierrezia wrightii</i> Gray	Wright's snakeweed		X						X		
	<i>Gymnosperma glutinosum</i> (Spreng.) Less.	gumhead		X						X	X	
	<i>Helianthus annuus</i> L.	common sunflower		X						X		
	<i>Helianthus petiolaris</i> ssp. <i>fallax</i> Heiser	prairie sunflower	X	X								
	<i>Heliomeris longifolia</i> var. <i>annua</i> (M.E. Jones) Yates	longleaf false goldeneye		X		X				X	X	
	<i>Heliomeris longifolia</i> var. <i>longifolia</i> (Robins. & Greenm.) Cockerell	longleaf false goldeneye								X		
	<i>Heliomeris multiflora</i> var. <i>multiflora</i> Nutt.	showy goldeneye		X								
	<i>Heliopsis parvifolia</i> Gray	mountain oxeye		X						X		
	<i>Heterosperma pinnatum</i> Cav.	wingpetal	X	X						X	X	
	<i>Heterotheca subaxillaris</i> (Lam.) Britt. & Rusby	camphorweed	X	X	X	X				X	X	
	<i>Heterotheca villosa</i> var. <i>minor</i> (Hook.) Semple	hairy false goldenaster		X						X		
	<i>Hymenothrix wislizeni</i> Gray	TransPecos thimblehead	X	X						X		
	<i>Hymenothrix wrightii</i> Gray	Wright's thimblehead		X						X	X	
	<i>Isocoma tenuisecta</i> Greene	burroweed	X	X		X		X				
	<i>Iva ambrosiifolia</i> (Gray) Gray	ragged marshelder		X							X	
	<i>Lactuca serriola</i> L.	lettuce			X							
	<i>Laennecia coulteri</i> (Gray) Nesom	conyza	X	X						X		
	<i>Laennecia sopheriifolia</i> (Kunth) Nesom	leafy marshail	X	X						X	X	

Family			Studies						Specimens in herbaria			
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Asteraceae												
	<i>Lasianthaea podocephala</i> (Gray)											
	K. Becker	San Pedro daisy	X	X						X	X	
	<i>Machaeranthera canescens</i> (Pursh)											
	Gray var. <i>canescens</i>	hoary tansyaster									X	
	<i>Machaeranthera gracilis</i> (Nutt.) Shinnery	slender goldenweed	X	X						X		
	<i>Machaeranthera parviflora</i> Gray	smallflower tansyaster	X	X								
	<i>Machaeranthera pinnatifida</i> (Hook.) Shinnery	lacy tansyaster	X	X						X		
	<i>Machaeranthera tagetina</i> Greene	mesa tansyaster		X						X	X	
	<i>Machaeranthera tanacetifolia</i> (Kunth) Nees	tanseyleaf tansyaster	X	X								
	<i>Malacothrix fendleri</i> Gray	Fendler's desertdandelion	X	X						X		X
	<i>Melampodium leucanthum</i> Torr. & Gray	plains blackfoot		X						X		
	<i>Melampodium longicorne</i> Gray	Arizona blackfoot	X	X						X		
	<i>Melampodium strigosum</i> Stuessy	shaggy blackfoot	X	X						X	X	
	<i>Packera neomexicana</i> (Gray)											
	W.A. Weber & A. Löve	New Mexico groundsel								X		
	<i>Packera neomexicana</i> var. <i>neomexicana</i> (Gray) W.A. Weber & A. Löve	New Mexico groundsel		X								
	<i>Packera neomexicana</i> var. <i>toumeyii</i> (Greene) D.K. Trock & T.M. Barkl.	Toumey's groundsel								X		
	<i>Parthenium incanum</i> Kunth	mariola								X		
	<i>Pectis filipes</i> var. <i>subnuda</i> Fern.	fivebract cinchweed	X	X						X	X	
	<i>Pectis imberbis</i> Gray	beardless cinchweed	X	X						X		
	<i>Pectis longipes</i> Gray	longstalk cinchweed		X						X		
	<i>Pectis prostrata</i> Cav.	spreading cinchweed									X	
	<i>Perityle coronopifolia</i> Gray	crowfoot rockdaisy								X		
	<i>Porophyllum ruderales</i> ssp. <i>macrocephalum</i> (DC.) R.R. Johnson	yerba porosa		X						X		X
	<i>Pseudognaphalium arizonicum</i> (Gray) A. Anderb.	Arizona cudweed		X		X						
	<i>Pseudognaphalium canescens</i> ssp. <i>canescens</i> (DC.) W.A. Weber	Wright's cudweed	X	X								
	<i>Pseudognaphalium leucocephalum</i> (Gray) A. Anderb.	white cudweed									X	
	<i>Sanvitalia abertii</i> Gray	Albert's creeping zinnia	X	X						X	X	
	<i>Schkuhria anthemioidea</i> var. <i>wrightii</i> (Gray) Heiser	Wright's false threadleaf										X
	<i>Schkuhria pinnata</i> (Lam.) Kuntze ex Thell.	pinnate false threadleaf								X		
	<i>Schkuhria pinnata</i> var. <i>wislizenii</i> (Gray) B.L. Turner	Wislizenus' false threadleaf		X								
	<i>Senecio flaccidus</i> Less.	threadleaf ragwort								X		
	<i>Senecio flaccidus</i> var. <i>flaccidus</i> Less.	threadleaf ragwort	X	X						X	X	
	<i>Solidago canadensis</i> L.	Canada goldenrod								X		
	<i>Solidago canadensis</i> var. <i>scabra</i> Torr. & Gray	Canada goldenrod	X	X								
	<i>Solidago velutina</i> DC.	threenerve goldenrod		X								

Family	Studies							Specimens in herbaria				
	Scientific name	Common name	R&J	P&C	H&G	Ruy	NPS	UA	ASU	DBG	NAU	UA
Asteraceae												
	<i>Solidago wrightii</i> Gray	Wright's goldenrod		X						X		
	<b><i>Sonchus asper</i> (L.) Hill</b>	<b>spiny sowthistle</b>		<b>X</b>						<b>X</b>		
	<b><i>Sonchus oleraceus</i> L.</b>	<b>common sowthistle</b>		<b>X</b>						<b>X</b>		
	<i>Stephanomeria exigua</i> Nutt.	small wirelettuce	X	X								
	<i>Stephanomeria pauciflora</i> (Torr.) A. Nels.	brownplume wirelettuce		X						X		
	<i>Stephanomeria thurberi</i> Gray	Thurber's wirelettuce		X						X		
	<i>Stevia serrata</i> Cav.	sawtooth candyleaf		X								
	<i>Tagetes lemmonii</i> Gray	Lemmon's marigold		X			X			X		
	<i>Tagetes micrantha</i> Cav.	licorice marigold								X		
	<i>Thelesperma longipes</i> Gray	longstalk greenthread		X								
	<i>Thelesperma megapotamicum</i> (Spreng.) Kuntze	Hopi tea greenthread		X						X		
	<i>Thymophylla acerosa</i> (DC.) Strother	pricklyleaf dogweed	X	X								
	<i>Thymophylla pentachaeta</i> (DC.) Small	fiveneedle pricklyleaf								X		
	<i>Thymophylla pentachaeta</i> var. <i>pentachaeta</i> (DC.) Small	fiveneedle pricklyleaf		X								
	<i>Trixis californica</i> Kellogg	American threefold	X	X								
	<i>Verbesina encelioides</i> (Cav.) Benth. & Hook. f. ex Gray	golden crownbeard	X	X						X		
	<i>Verbesina longifolia</i> (Gray) Gray	longleaf crownbeard		X								
	<i>Verbesina rothrockii</i> Robins. & Greenm.	Rothrock's crownbeard		X						X		
	<i>Viguiera cordifolia</i> Gray	heartleaf goldeneye	X	X						X		
	<i>Viguiera dentata</i> (Cav.) Spreng.	toothleaf goldeneye	X	X						X		
	<i>Viguiera dentata</i> (Cav.) Spreng. var. <i>dentata</i>	toothleaf goldeneye									X	X
	<i>Xanthium strumarium</i> L.	rough cocklebur	X	X	X							
	<i>Zinnia acerosa</i> (DC.) Gray	desert zinnia				X						
	<i>Zinnia grandiflora</i> Nutt.	Rocky Mountain zinnia	X	X						X		X
	<b><i>Zinnia peruviana</i> (L.) L.</b>	<b>Peruvian zinnia</b>		<b>X</b>							<b>X</b>	
Bignoniaceae												
	<i>Chilopsis linearis</i> (Cav.) Sweet	desert willow	X	X				X		X		
Boraginaceae												
	<i>Cryptantha cinerea</i> var. <i>jamesii</i> Cronq.	James' cryptantha		X								
	<i>Cryptantha pusilla</i> (Torr. & Gray) Greene	low cryptantha		X						X		X
	<i>Lithospermum cobrense</i> Greene	smooththroat stoneseed		X						X		
	<i>Plagiobothrys arizonicus</i> (Gray) Greene ex Gray	Arizona popcornflower		X						X		
Brassicaceae												
	<b><i>Brassica tournefortii</i> Gouan Asian</b>	<b>mustard</b>		<b>X</b>								
	<i>Descurainia pinnata</i> (Walt.) Britt.	western tansymustard	X	X						X		
	<b><i>Descurainia sophia</i> (L.) Webb ex Prantl</b>	<b>herb sophia</b>		<b>X</b>	<b>X</b>							<b>X</b>
	<i>Draba cuneifolia</i> Nutt. ex Torr. & Gray	wedgeleaf draba										X
	<i>Draba cuneifolia</i> Nutt. ex Torr. & Gray var. <i>cuneifolia</i>	wedgeleaf draba		X								
	<i>Erysimum capitatum</i> (Dougl. ex Hook.) Greene	sanddune wallflower									X	



Family			Studies						Specimens in herbaria			
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Brassicaceae												
	<i>Erysimum capitatum</i> var. <i>capitatum</i> (Dougl. ex Hook.) Greene	sanddune wallflower		X								
	<i>Lepidium lasiocarpum</i> Nutt.	shaggyfruit pepperweed		X								X
	<i>Lepidium oblongum</i> Small	veiny pepperweed								X		
	<i>Lepidium thurberi</i> Woot.	Thurber's pepperweed	X	X								
	<i>Lesquerella gordonii</i> (Gray) S. Wats.	Gordon's bladderpod		X								
	<i>Lesquerella tenella</i> A. Nels.	Moapa bladderpod										X
	<i>Schoenocrambe linearifolia</i> (Gray) Rollins	slimleaf plainsmustard	X	X			X			X	X	
	<b><i>Sisymbrium irio</i> L.</b>	<b>London rocket</b>	<b>X</b>	<b>X</b>	<b>X</b>					<b>X</b>		<b>X</b>
	<i>Thysanocarpus curvipes</i> Hook.	sand fringe-pod		X						X		
Cactaceae												
	<i>Echinocereus coccineus</i> var. <i>arizonicus</i> (Rose ex Orcutt) Ferguson	Arizona hedgehog cactus								X		
	<i>Echinocereus fendleri</i> (Engelm.) F. Seitz	pinkflower hedgehog cactus								X		
	<i>Echinocereus fendleri</i> var. <i>rectispinus</i> (Peebles) L. Benson	pinkflower hedgehog cactus		X								X
	<i>Echinocereus pectinatus</i> (Scheidw.) Engelm.	rainbow cactus		X			X					
	<i>Echinocereus pectinatus</i> (Scheidw.) Engelm. var. <i>pectinatus</i>	rainbow cactus	X									
	<i>Echinocereus polyacanthus</i> Engelm.	Mojave mound cactus										X
	<i>Echinocereus rigidissimus</i> (Engelm.) Haage f.	rainbow hedgehog cactus	X	X			X					
	<i>Echinocereus triglochidiatus</i> Engelm.	kingcup cactus		X						X		
	<i>Escobaria vivipara</i> var. <i>bisbeeana</i> (Orcutt) D.R. Hunt	Bisbee spinystar		X								X
	<i>Escobaria vivipara</i> var. <i>vivipara</i> (Nutt.) Buxbaum	spinystar	X				X					
	<i>Mammillaria heyderi</i> var. <i>macdougalii</i> (Rose) L. Benson	Macdougal's nipple cactus	X	X								
	<i>Mammillaria wrightii</i> var. <i>wilcoxii</i> (Toumey ex K. Schum.) W.T. Marsh.	Wilcox's nipple cactus		X						X		
	<i>Opuntia arbuscula</i> Engelm.	Arizona pencil cholla				X						
	<i>Opuntia chlorotica</i> Engelm. & Bigelow	dollarjoint pricklypear	X	X						X	X	
	<i>Opuntia engelmannii</i> Salm-Dyck	cactus apple				X	X	X				
	<i>Opuntia engelmannii</i> Salm-Dyck var. <i>engelmannii</i>	cactus apple	X	X						X		
	<i>Opuntia macrocentra</i> Engelm.	purple pricklypear		X						X		
	<i>Opuntia macrorhiza</i> Engelm. var. <i>macrorhiza</i>	twistspine pricklypear	X	X						X		
	<i>Opuntia phaeacantha</i> Engelm.	tulip pricklypear	X				X					
	<i>Opuntia spinosior</i> (Engelm.) Toumey	walkingstick cactus	X	X			X	X				X
	<i>Opuntia versicolor</i> Engelm. ex Coult.	staghorn cholla				X						
	<i>Sclerocactus intertextus</i> var. <i>intertextus</i> (Engelm.) N.P. Taylor	white fishhook cactus		X								X
Campanulaceae												
	<i>Lobelia cardinalis</i> L.	cardinalflower								X		
	<i>Triodanis perfoliata</i> (L.) Nieuwl.	clasping Venus' looking-glass		X						X		

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			R&J	P&C	H&G	Ruy	NPS	UA	ASU	DBG	NAU	UA
Campanulaceae												
	<i>Triodanis perfoliata</i> var. <i>biflora</i> (Ruiz & Pavón) Bradley	clasping Venus' looking-glass		X								
Caprifoliaceae												
	<i>Lonicera albiflora</i> Torr. & Gray	western white honeysuckle	X	X						X		
	<i>Sambucus nigra</i> L.	European black elderberry								X		
	<i>Sambucus nigra</i> ssp. <i>canadensis</i> (L.) R. Bolli	common elderberry		X								
	<i>Sambucus nigra</i> ssp. <i>cerulea</i> (Raf.) R. Bolli	blue elderberry						X				
Caryophyllaceae												
	<i>Arenaria lanuginosa</i> (Michx.) Rohrb.	spreading sandwort								X		
	<i>Arenaria lanuginosa</i> ssp. <i>saxosa</i> (Gray) Maguire	spreading sandwort		X								
	<i>Cerastium texanum</i> Britt.	Texas chickweed		X						X		X
	<i>Drymaria molluginea</i> (Lag.) Didr.	slimleaf drymary		X						X		X
	<i>Silene antirrhina</i> L.	sleepy silene		X						X		X
	<i>Silene laciniata</i> Cav.	cardinal catchfly								X		
	<i>Silene laciniata</i> ssp. <i>greggii</i> (Gray) C.L. Hitchc. & Maguire	cardinal catchfly		X								
Chenopodiaceae												
	<i>Atriplex elegans</i> (Moq.) D. Dietr. var. <i>elegans</i>	wheelscale saltbush	X	X								
	<i>Chenopodium fremontii</i> S. Wats.	Fremont's goosefoot	X	X						X		X
	<i>Chenopodium graveolens</i> Willd.	fetid goosefoot	X	X							X	
	<i>Chenopodium neomexicanum</i> Standl.	New Mexico goosefoot		X						X		
	<b><i>Salsola kali</i> L.</b>	<b>Russian thistle</b>					X					
	<b><i>Salsola tragus</i> L.</b>	<b>prickly Russian thistle</b>		X						X		
Commelinaceae												
	<i>Commelina dianthifolia</i> Delile	birdbill dayflower	X	X						X	X	
	<i>Tradescantia pinetorum</i> Greene	pinewoods spiderwort		X								
Convolvulaceae												
	<i>Calystegia longipes</i> (S. Wats.) Brummitt	Paiute false bindweed		X								
	<i>Convolvulus equitans</i> Benth.	Texas bindweed	X	X						X		
	<i>Evolvulus alsinoides</i> (L.) L.	slender dwarf morning-glory	X	X								
	<i>Evolvulus arizonicus</i> Gray	wild dwarf morning-glory	X	X		X				X	X	
	<i>Evolvulus nuttallianus</i> J.A. Schultes	shaggy dwarf morning-glory		X						X		
	<i>Evolvulus sericeus</i> Sw.	silver dwarf morning-glory	X	X						X		
	<i>Ipomoea capillacea</i> (Kunth) G. Don	purple morning-glory	X	X						X		
	<b><i>Ipomoea coccinea</i> L.</b>	<b>redstar</b>	<b>X</b>									
	<i>Ipomoea costellata</i> Torr.	crestrub morning-glory	X	X						X	X	
	<i>Ipomoea cristulata</i> Hallier f.	Transpecos morning-glory		X							X	
	<b><i>Ipomoea hederacea</i> Jacq.</b>	<b>ivy leaf morning-glory</b>								<b>X</b>		
	<i>Ipomoea longifolia</i> Benth.	pinkthroat morning-glory	X	X		X				X		
	<b><i>Ipomoea purpurea</i> (L.) Roth</b>	<b>tall morning-glory</b>	<b>X</b>	<b>X</b>	<b>X</b>					<b>X</b>	<b>X</b>	
Cucurbitaceae												
	<i>Cucurbita digitata</i> Gray	fingerleaf gourd	X	X								
	<i>Cucurbita foetidissima</i> Kunth	Missouri gourd	X	X								
	<i>Cucurbita palmata</i> S. Wats.	coyote gourd				X						

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			R&J	P&C	H&G	Ruy	NPS	UA	ASU	DBG	NAU	UA
Cucurbitaceae												
	<i>Sicyos ampelophyllus</i> Woot. & Standl.	streamside burr cucumber	X	X								
Cupressaceae												
	<i>Cupressus arizonica</i> Greene											
	ssp. <i>arizonica</i>	Arizona cypress		X								
	<i>Juniperus coahuilensis</i> (Martinez) Gaussen ex R.P. Adams	redberry juniper	X	X							X	
	<i>Juniperus deppeana</i> Steud.	alligator juniper	X	X			X	X				
Cuscutaceae												
	<i>Cuscuta appianata</i> Engelm.	Gila River dodder		X								
Cyperaceae												
	<i>Bulbostylis capillaris</i> (L.) Kunth ex C.B. Clarke	densetuft hairsedge		X						X		X
	<i>Bulbostylis funckii</i> (Steud.) C.B. Clarke	Funck's hairsedge		X								
	<i>Cyperus aggregatus</i> (Willd.) Endl.	inflatedscale flatsedge	X	X								
	<i>Cyperus dipsaceus</i> Liebmamann	Wright's flatsedge										X
	<i>Cyperus fendlerianus</i> Boeckl.	Fendler's flatsedge	X	X						X	X	
	<i>Cyperus mutisii</i> (Kunth) Griseb.	Mutis' flatsedge										X
	<i>Cyperus niger</i> Ruiz & Pavón	black flatsedge	X	X								
	<i>Cyperus pallidicolor</i> (Kükenth.) G. Tucker	pallid flatsedge								X		
	<i>Cyperus spectabilis</i> Link	spectacular flatsedge		X						X		
	<i>Cyperus sphaerolepis</i> Boeckl.	Rusby's flatsedge		X						X		
	<i>Cyperus squarrosus</i> L.	bearded flatsedge										X
	<i>Lipocarpa micrantha</i> (Vahl) G. Tucker	smallflower halfchaff sedge										X
	<i>Schoenoplectus acutus</i> var. <i>acutus</i> (Muhl. ex Bigelow) A. & D. Löve	hardstem bulrush		X								X
Dryopteridaceae												
	<i>Woodsia</i> R. Br.	cliff fern				X						
Equisetaceae												
	<i>Equisetum ×ferrissii</i> Clute (pro sp.)	ferris horsetail		X								X
	<i>Equisetum</i> L.	horsetail								X		
Ericaceae												
	<i>Arbutus arizonica</i> (Gray) Sarg.	Arizona madrone	X	X				X				X
	<i>Arctostaphylos pringlei</i> Parry	Pringle manzanita	X	X								
	<i>Arctostaphylos pungens</i> Kunth	pointleaf manzanita	X	X		X	X	X		X		
Euphorbiaceae												
	<i>Acalypha neomexicana</i> Muell.-Arg.	New Mexico copperleaf	X	X						X		X
	<i>Acalypha ostryifolia</i> Riddell	pineland threeseed mercury		X						X		
	<i>Chamaesyce dioica</i> (Kunth) Millsp.	royal sandmat	X	X						X		
	<i>Chamaesyce hirta</i> (L.) Millsp.	pillpod sandmat		X						X		X
	<i>Chamaesyce hyssopifolia</i> (L.) Small	hyssopleaf sandmat	X	X						X		
	<i>Chamaesyce prostrata</i> (Ait.) Small	prostrate sandmat	X	X						X		
	<i>Chamaesyce revoluta</i> (Engelm.) Small	threadstem sandmat		X						X		
	<i>Chamaesyce serpyllifolia</i> (Pers.) Small	thymeleaf sandmat								X		
	<i>Chamaesyce serpyllifolia</i> ssp. <i>serpyllifolia</i> (Pers.) Small	thymeleaf sandmat		X								
	<i>Croton pottsii</i> (Klotzsch) Muell.-Arg.	leatherweed	X			X				X		
	<i>Croton pottsii</i> var. <i>pottsii</i> (Klotzsch) Muell.-Arg.	leatherweed		X								

Family	Studies								Specimens in herbaria			
	Scientific name	Common name	R&J	P&C	H&G	Ruy	NPS	UA	ASU	DBG	NAU	UA
Euphorbiaceae												
	<i>Euphorbia bilobata</i> Engelm.	blackseed spurge		X						X		X
	<i>Euphorbia brachycera</i> Engelm.	horned spurge	X	X						X		
	<i>Euphorbia cyathophora</i> Murr.	fire on the mountain										X
	<b><i>Euphorbia dentata</i> Michx.</b>	<b>toothed spurge</b>	<b>X</b>	<b>X</b>								
	<i>Euphorbia exstipulata</i> Engelm.	squareseed spurge		X								X
	<i>Euphorbia heterophylla</i> L.	Mexican fireplant	X	X							X	X
	<i>Euphorbia incisa</i> Engelm.	Mojave spurge		X								
	<i>Jatropha macrorhiza</i> Benth.	ragged nettlespurge										X
	<i>Jatropha macrorhiza</i> var. <i>septemfida</i> Engelm.	ragged nettlespurge	X	X							X	
	<i>Tragia nepetifolia</i> Cav.	catnip noseburn		X								X
	<i>Tragia ramosa</i> Torr.	branched noseburn		X								
Fabaceae												
	<i>Acacia angustissima</i> (P. Mill.) Kuntze	prairie acacia	X			X	X	X			X	
	<i>Acacia angustissima</i> var. <i>suffrutescens</i> (Rose) Isely	prairie acacia		X								
	<i>Acacia constricta</i> Benth.	whitethorn acacia	X	X								
	<i>Acacia greggii</i> Gray	catclaw acacia	X	X								
	<i>Amorpha fruticosa</i> L.	desert false indigo	X	X		X						
	<i>Astragalus allochrous</i> var. <i>playanus</i> Isely	halfmoon milkvetch		X								X
	<i>Astragalus hypoxylus</i> S. Wats.	Huachuca Mountain milkvetch	X	X								
	<i>Astragalus lentiginosus</i> var. <i>australis</i> Barneby	freckled milkvetch	X	X								
	<i>Astragalus nothoxys</i> Gray	sheep milkvetch		X						X		X
	<i>Astragalus nuttallianus</i> DC.	smallflowered milkvetch		X								
	<i>Astragalus thurberi</i> Gray	Thurber's milkvetch		X								X
	<b><i>Caesalpinia gilliesii</i> (Hook.) Wallich ex D. Dietr.</b>	<b>bird-of-paradise shrub</b>	<b>X</b>	<b>X</b>								
	<i>Calliandra eriophylla</i> Benth.	fairyduster	X	X		X	X	X				
	<i>Calliandra humilis</i> var. <i>reticulata</i> (Gray) L. Benson	dwarf stickpea		X								
	<i>Chamaecrista nictitans</i> (L.) Moench	partridge pea					X					
	<i>Chamaecrista nictitans</i> var. <i>leptadenia</i> (Greenm.) Gandhi & Hatch	partridge pea		X								
	<i>Clitoria mariana</i> L.	Atlantic pigeonwings	X	X								
	<i>Cologania angustifolia</i> Kunth	longleaf cologania	X	X								
	<i>Coursetia caribaea</i> var. <i>caribaea</i> (Jacq.) Lavin	anil falso	X	X								
	<i>Crotalaria pumila</i> Ortega	low rattlebox	X	X							X	
	<i>Crotalaria sagittalis</i> L.	arrowhead rattlebox	X	X								
	<i>Dalea albiflora</i> Gray	whiteflower prairie clover	X	X							X	X
	<i>Dalea brachystachya</i> Gray	Fort Bowie prairie clover		X							X	
	<i>Dalea exigua</i> Barneby	Chihuahuan prairie clover	X	X								X
	<i>Dalea filiformis</i> Gray	Sonoran prairie clover		X								
	<i>Dalea grayi</i> (Vail) L.O. Williams	Gray's prairie clover		X								
	<i>Dalea nana</i> var. <i>canescens</i> Kearney & Peebles	dwarf prairie clover		X								

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			R&J	P&C	H&G	Ruy	NPS	UA	ASU	DBG	NAU	UA
Fabaceae												
	<i>Dalea pogonathera</i> Gray	bearded prairie clover		X								
	<i>Dalea pulchra</i> H.C. Gentry	Santa Catalina prairie clover					X			X		
	<i>Dalea versicolor</i> var. <i>sessilis</i> (Gray) Barneby	oakwoods prairie clover	X	X						X		
	<i>Desmanthus cooleyi</i> (Eat.) Trel.	Cooley's bundleflower	X	X								
	<i>Desmodium batocaulon</i> Gray	San Pedro ticktrefoil	X	X								
	<i>Desmodium cinerascens</i> Gray	spiked ticktrefoil		X								X
	<i>Desmodium neomexicanum</i> Gray	New Mexico ticktrefoil	X	X								
	<i>Desmodium retinens</i> Schlecht.	Santa Rita Mountain ticktrefoil	X	X								
	<i>Desmodium rosei</i> Schub.	Rose's ticktrefoil	X	X						X	X	
	<i>Erythrina flabelliformis</i> Kearney	coralbean	X	X		X						
	<i>Eysenhardtia orthocarpa</i> (Gray) S. Wats.	Tahitian kidneywood	X	X				X			X	X
	<i>Galactia wrightii</i> var. <i>mollissima</i> Kearney & Peebles			X								
	<i>Galactia wrightii</i> Gray var. <i>wrightii</i>	Wright's milkpea		X								
	<i>Hoffmannseggia glauca</i> (Ortega) Eifert	Indian rushpea				X						
	<i>Lathyrus graminifolius</i> (S. Wats.) White	grassleaf pea	X	X								
	<i>Lotus greenei</i> Ottley ex Kearney & Peebles	Greene's bird's-foot trefoil	X	X						X		
	<i>Lotus humistratus</i> Greene	foothill deervetch		X								
	<i>Lotus plebeius</i> (Brand) Barneby	New Mexico bird's-foot trefoil	X	X								
	<i>Lotus wrightii</i> (Gray) Greene	Wright's deervetch	X	X							X	
	<i>Lupinus concinnus</i> J.G. Agardh	scarlet lupine		X								
	<i>Lupinus palmeri</i> S. Wats.	bluebonnet lupine	X	X								
	<b><i>Macroptilium gibbosifolium</i> (Ortega) A. Delgado</b>	<b>variableleaf bushbean</b>	<b>X</b>	<b>X</b>						<b>X</b>	<b>X</b>	
	<b><i>Medicago lupulina</i> L.</b>	<b>black medick</b>		<b>X</b>						<b>X</b>		
	<b><i>Melilotus indicus</i> (L.) All.</b>	<b>annual yellow sweetclover</b>		<b>X</b>						<b>X</b>		
	<i>Mimosa aculeaticarpa</i> Ortega	catclaw mimosa					X					
	<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i> (Benth.) Barneby	catclaw mimosa	X	X		X	X	X				
	<i>Mimosa dysocarpa</i> Benth.	velvetpod mimosa	X	X			X					
	<i>Mimosa grahamii</i> Gray	Graham's mimosa	X	X				X				
	<i>Nissolia wislizeni</i> (Gray) Gray	Arizona yellowhood		X								
	<i>Phaseolus acutifolius</i> var. <i>tenuifolius</i> Gray	teparty bean		X								
	<i>Phaseolus maculatus</i> Scheele	spotted bean		X								
	<i>Phaseolus ritensis</i> M.E. Jones	Santa Rita Mountain bean	X	X								
	<i>Prosopis glandulosa</i> Torr.	honey mesquite		X		X						
	<b><i>Prosopis juliflora</i> (Sw.) DC.</b>	<b>mesquite</b>	<b>X</b>									
	<i>Prosopis velutina</i> Woot.	velvet mesquite					X	X				X
	<i>Psoralidium tenuiflorum</i> (Pursh) Rydb.	slimflower scurfpea		X								
	<i>Rhynchosia senna</i> var. <i>texana</i> (Torr. & Gray) M.C. Johnston	Texas snoutbean		X								
	<i>Robinia neomexicana</i> Gray	New Mexico locust	X	X								
	<i>Senna hirsuta</i> var. <i>glaberrima</i> (M.E. Jones) Irwin & Barneby	woolly senna	X	X								

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			R&J	P&C	H&G	Ruy	NPS	UA	ASU	DBG	NAU	UA
Fabaceae												
	<i>Senna lindheimeriana</i> (Scheele)											
	Irwin & Barneby	velvet leaf senna	X	X								
	<i>Tephrosia tenella</i> Gray	red hoarypea		X								
	<i>Tephrosia thurberi</i> (Rydb.) C.E. Wood	Thurber's hoarypea		X								
	<i>Vicia ludoviciana</i> Nutt.	Louisiana vetch		X						X		
Fagaceae												
	<i>Quercus ajoensis</i> C.H. Muller	Ajo Mountain scrub oak						X				
	<i>Quercus arizonica</i> Sarg.	Arizona white oak	X					X	X			
	<i>Quercus dunni</i> Kellogg	Palmer oak	X	X								
	<i>Quercus emoryi</i> Torr.	Emory oak	X	X		X	X	X		X		
	<i>Quercus gambelii</i> Nutt.	Gambel oak	X	X								
	<i>Quercus grisea</i> Liebm.	gray oak		X						X	X	
	<i>Quercus hypoleucoides</i> A. Camus	silverleaf oak	X	X			X	X		X		
	<i>Quercus oblongifolia</i> Torr.	Mexican blue oak	X	X		X	X	X		X		
	<i>Quercus pungens</i> Liebm.	pungent oak	X	X								
	<i>Quercus rugosa</i> Née	netleaf oak	X	X								
	<i>Quercus toumey</i> Sarg.	Toumey oak	X	X								X
	<i>Quercus turbinella</i> Greene	Sonoran scrub oak	X	X			X					
Fouquieriaceae												
	<i>Fouquieria splendens</i> Engelm.	ocotillo	X	X		X						
Fumariaceae												
	<i>Corydalis aurea</i> Willd.	scrambled eggs		X								
	<i>Corydalis curvisiliqua</i> ssp. <i>occidentalis</i>											
	(Engelm. ex Gray) W.A. Weber	curvepod fumewort										X
Garryaceae												
	<i>Garrya flavescens</i> S. Wats.	ashy silktassel						X				
	<i>Garrya wrightii</i> Torr.	Wright's silktassel	X	X				X	X		X	
Gentianaceae												
	<i>Centaurium calycosum</i> (Buckl.) Fern.	Arizona centaury	X	X						X		
Geraniaceae												
	<b><i>Erodium cicutarium</i> (L.) L'Hér. ex Ait.</b>	<b>redstem stork's bill</b>		<b>X</b>	<b>X</b>					<b>X</b>		
	<i>Geranium caespitosum</i> var. <i>parryi</i>											
	(Engelm.) W.A. Weber	Parry's geranium		X						X		
Grossulariaceae												
	<i>Ribes</i> L.	currant		X								
Hydrangeaceae												
	<i>Fendlera rupicola</i> Gray	cliff fendlerbush		X						X		
	<i>Philadelphus microphyllus</i> Gray	littleleaf mock orange	X	X						X	X	X
Hydrophyllaceae												
	<i>Nama dichotomum</i> (Ruiz & Pavón)											
	Choisy	wishbone fiddleleaf		X						X		
	<i>Phacelia arizonica</i> Gray	Arizona phacelia		X						X		
	<i>Phacelia caerulea</i> Greene	skyblue phacelia		X						X		
Juglandaceae												
	<i>Juglans major</i> (Torr.) Heller	Arizona walnut	X	X				X				X
Juncaceae												
	<i>Juncus bufonius</i> L.	toad rush		X								

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Juncaceae												
	<i>Juncus ensifolius</i> Wikstr.	swordleaf rush										X
	<i>Juncus saximontanus</i> A. Nels.	Rocky Mountain rush		X						X		
	<i>Juncus tenuis</i> Willd.	poverty rush		X						X		
Krameriaceae												
	<i>Krameria erecta</i> Willd. ex J.A. Schultes	littleleaf ratany	X	X		X				X		
Lamiaceae												
	<i>Agastache wrightii</i> (Greenm.) Woot. & Standl.	Sonoran giant hyssop	X	X								
	<i>Hedeoma dentata</i> Torr.	dentate false pennyroyal	X	X						X	X	
	<i>Hedeoma nana</i> (Torr.) Briq.	dwarf false pennyroyal								X		
	<i>Hedeoma nana</i> (Torr.) Briq. ssp. <i>nana</i>	dwarf false pennyroyal		X								
	<i>Salvia lemmonii</i> Gray	Lemmon's sage	X	X						X		X
	<i>Salvia parryi</i> Gray	Parry's sage	X	X						X		
	<i>Salvia subincisa</i> Benth.	sawtooth sage	X	X						X	X	
	<i>Stachys coccinea</i> Ortega	scarlet hedgenettle	X	X								
	<i>Trichostema arizonicum</i> Gray	Arizona bluecurls	X	X						X	X	
Liliaceae												
	<i>Allium macropetalum</i> Rydb.	largeflower onion				X						
	<b>Asparagus officinalis L.</b>	<b>garden asparagus</b>	<b>X</b>	<b>X</b>								
	<i>Dasylirion wheeleri</i> S. Wats.	common sotol	X	X		X	X	X		X		
	<i>Dichelostemma capitatum</i> (Benth.) Wood ssp. <i>capitatum</i>	bluedicks	X	X		X						
	<i>Dichelostemma capitatum</i> ssp. <i>pauciflorum</i> (Torr.) G. Keator	bluedicks								X		X
	<i>Echeandia flavescens</i> (J.A. & J.H. Schultes) Cruden	Torrey's craglily	X	X							X	X
	<i>Milla biflora</i> Cav.	Mexican star	X	X						X	X	
	<i>Nolina microcarpa</i> S. Wats.	sacahuista	X	X		X	X	X		X	X	
Linaceae												
	<i>Linum puberulum</i> (Engelm.) Heller	plains flax	X	X						X		X
Loasaceae												
	<i>Mentzelia albicaulis</i> (Dougl. ex Hook.) Dougl. ex Torr. & Gray	whitestem blazingstar		X						X		X
	<i>Mentzelia asperula</i> Woot. & Standl.	Organ Mountain blazingstar								X		X
	<i>Mentzelia isolata</i> H.C. Gentry	isolated blazingstar								X		
	<i>Mentzelia montana</i> (A. Davids.) A. Davids.	variegated-bract blazingstar								X		
	<i>Mentzelia multiflora</i> (Nutt.) Gray	Adonis blazingstar	X	X								X
	<i>Mentzelia texana</i> Urban & Gilg	Texas blazingstar	X	X								
Lythraceae												
	<i>Cuphea wrightii</i> Gray	Wright's waxweed	X	X						X	X	X
	<i>Lythrum californicum</i> Torr. & Gray	California loosestrife								X		
Malpighiaceae												
	<i>Aspicarpa hirtella</i> L.C. Rich.	chaparral aspehead		X						X		X
Malvaceae												
	<i>Abutilon parvulum</i> Gray	dwarf Indian mallow		X						X		
	<i>Anoda cristata</i> (L.) Schlecht.	crested anoda	X	X						X	X	
	<b>Malva parviflora L.</b>	<b>cheeseweed mallow</b>		<b>X</b>	<b>X</b>							
	<b>Sida abutifolia P. Mill.</b>	<b>spreading fanpetals</b>	<b>X</b>	<b>X</b>		<b>X</b>				<b>X</b>	<b>X</b>	<b>X</b>

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Malvaceae												
	<i>Sida neomexicana</i> Gray	New Mexico fanpetals	X	X						X		
	<i>Sida spinosa</i> L.	prickly fanpetals		X						X		
	<i>Sphaeralcea angustifolia</i> (Cav.) G. Don	copper globemallow		X						X		
Molluginaceae												
	<i>Mollugo verticillata</i> L.	green carpetweed	X	X						X		
Moraceae												
	<i>Morus microphylla</i> Buckl.	Texas mulberry	X	X								
Nyctaginaceae												
	<i>Allionia incarnata</i> L.	trailing windmills	X	X						X		
	<i>Boerhavia coccinea</i> P. Mill.	scarlet spiderling	X	X	X					X	X	
	<i>Boerhavia erecta</i> L.	erect spiderling	X	X						X		X
	<i>Boerhavia purpurascens</i> Gray	purple spiderling	X	X						X	X	
	<i>Mirabilis albida</i> (Walt.) Heimerl	white four o'clock		X						X		
	<i>Mirabilis coccinea</i> (Torr.) Benth. & Hook. f.	scarlet four o'clock	X	X						X		
	<i>Mirabilis comata</i> (Small) Standl.	hairy-tuft four o'clock								X		X
	<i>Mirabilis linearis</i> (Pursh) Heimerl	narrowleaf four o'clock	X	X						X	X	
	<i>Mirabilis longiflora</i> L.	sweet four o'clock	X	X							X	
	<i>Mirabilis pumila</i> (Standl.) Standl.	dwarf four o'clock		X								
Oleaceae												
	<i>Fraxinus velutina</i> Torr.	velvet ash	X	X						X		
Onagraceae												
	<i>Camissonia chamaenerioides</i> (Gray) Raven	longcapsule suncup		X								X
	<i>Gaura coccinea</i> Nutt. ex Pursh	scarlet beeblossom		X						X		
	<i>Gaura hexandra</i> ssp. <i>gracilis</i> (Woot. & Standl.) Raven & Gregory	harlequinbush									X	
	<i>Oenothera albicaulis</i> Pursh	whitest evening-primrose	X	X						X		X
	<i>Oenothera brachycarpa</i> Gray	shortfruit evening-primrose	X	X							X	
	<i>Oenothera caespitosa</i> Nutt.	tufted evening-primrose		X								X
	<i>Oenothera primiveris</i> Gray	desert evening-primrose		X								X
Oxalidaceae												
	<i>Oxalis alpina</i> (Rose) Rose ex R. Knuth	alpine woodsorrel		X								
	<i>Oxalis corniculata</i> L.	creeping woodsorrel		X						X		
	<i>Oxalis decaphylla</i> Kunth	tenleaf woodsorrel		X						X		
Papaveraceae												
	<i>Argemone pleiakantha</i> Greene ssp. <i>pleiakantha</i>	southwestern pricklypoppy	X	X								X
Pedaliaceae												
	<i>Proboscidea parviflora</i> (Woot.) Woot. & Standl.	doubleclaw								X		
	<i>Proboscidea parviflora</i> (Woot.) Woot. & Standl. ssp. <i>parviflora</i>	doubleclaw	X	X						X		
Phytolaccaceae												
	<i>Phytolacca americana</i> L.	American pokeweed		X								
	<i>Phytolacca icosandra</i> L.									X		
Pinaceae												
	<i>Pinus cembroides</i> Zucc.	Mexican pinyon						X		X		
	<i>Pinus discolor</i> D.K. Bailey & Hawsworth	border pinyon	X	X			X					



Family	Scientific name	Common name	Studies						Specimens in herbaria			
			R&J	P&C	H&G	Ruy	NPS	UA	ASU	DBG	NAU	UA
Pinaceae												
	<b><i>Pinus pinea</i> L.</b>	<b>Italian stone pine</b>					<b>X</b>					
	<i>Pseudotsuga menziesii</i> (Mirbel) Franco	Douglas-fir								X		
Plantaginaceae												
	<i>Plantago patagonica</i> Jacq.	woolly plantain	X	X						X		
Platanaceae												
	<i>Platanus wrightii</i> S. Wats.	Arizona sycamore	X	X				X		X		
Poaceae												
	<i>Achnatherum eminens</i> (Cav.) Barkworth	southwestern needlegrass		X						X		X
	<i>Aristida adscensionis</i> L.	sixweeks threeawn	X	X			X			X		X
	<i>Aristida divaricata</i> Humb. & Bonpl. ex Willd.	poverty threeawn	X	X			X			X		
	<i>Aristida havardii</i> Vasey	Havard's threeawn					X					
	<i>Aristida pansa</i> Woot. & Standl.	Wooton's threeawn	X	X								
	<i>Aristida purpurea</i> Nutt.	purple threeawn					X					
	<i>Aristida purpurea</i> var. <i>nealleyi</i> (Vasey) Allred	blue threeawn		X								X
	<i>Aristida schiedeana</i> var. <i>orcuttiana</i> (Vasey) Allred & Valdés-Reyna	Orcutt's threeawn	X	X			X			X		
	<i>Aristida temipes</i> Cav.	spidergrass	X	X			X			X	X	
	<i>Aristida temipes</i> var. <i>gentilis</i> (Henr.) Allred	spidergrass	X	X								
	<i>Aristida temipes</i> Cav. var. <i>temipes</i>	spidergrass					X					
	<i>Blepharoneuron tricholepis</i> (Torr.) Nash	pine dropseed	X	X								
	<i>Bothriochloa barbinodis</i> (Lag.) Herter	cane bluestem	X	X		X	X			X		
	<i>Bouteloua aristidoides</i> (Kunth) Griseb.	needle grama	X	X								
	<i>Bouteloua barbata</i> Lag.	sixweeks grama	X	X								
	<i>Bouteloua chondrosioides</i> (Kunth) Benth. ex S. Wats.	sprucetop grama	X	X		X	X			X		
	<i>Bouteloua curtipendula</i> (Michx.) Torr.	sideoats grama		X		X	X			X	X	
	<i>Bouteloua curtipendula</i> var. <i>caespitosa</i> Gould & Kapadia	sideoats grama	X									
	<i>Bouteloua eludens</i> Griffiths	Santa Rita Mountain grama		X						X		X
	<i>Bouteloua eriopoda</i> (Torr.) Torr.	black grama	X	X		X	X					X
	<i>Bouteloua gracilis</i> (Willd. ex Kunth) Lag. ex Griffiths	blue grama	X	X		X	X			X	X	
	<i>Bouteloua hirsuta</i> Lag.	hairy grama		X		X	X			X		
	<i>Bouteloua hirsuta</i> Lag. var. <i>hirsuta</i>	hairy grama	X									
	<i>Bouteloua radicata</i> (Fourn.) Griffiths	purple grama	X	X		X				X	X	
	<i>Bouteloua repens</i> (Kunth) Scribn. & Merr.	slender grama	X	X		X	X			X		
	<i>Bouteloua rothrockii</i> Vasey	Rothrock's grama	X	X						X		X
	<i>Bromus anomalus</i> Rupr. ex Fourn.	nodding brome	X	X			X			X		
	<b><i>Bromus catharticus</i> Vahl</b>	<b>rescuegrass</b>		<b>X</b>	<b>X</b>		<b>X</b>			<b>X</b>		<b>X</b>
	<i>Bromus ciliatus</i> L.	fringed brome		X						X		X
	<i>Cenchrus spinifex</i> Cav.	coastal sandbur	X	X						X		
	<i>Chloris virgata</i> Sw.	feather fingergrass	X	X	X		X					
	<b><i>Cynodon dactylon</i> (L.) Pers.</b>	<b>Bermudagrass</b>	<b>X</b>	<b>X</b>	<b>X</b>					<b>X</b>		
	<i>Dasyochloa pulchella</i> (Kunth) Willd. ex Rydb.	low woollygrass	X	X								

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			R&J	P&C	H&G	Ruy	NPS	UA	ASU	DBG	NAU	UA
Poaceae												
	<i>Digitaria californica</i> (Benth.) Henr.	Arizona cottontop	X	X		X				X		
	<i>Digitaria cognata</i> (J.A. Schultes) Pilger	Carolina crabgrass								X		
	<i>Digitaria sanguinalis</i> (L.) Scop.	hairy crabgrass	X	X	X					X		
	<b><i>Echinochloa colona</i> (L.) Link</b>	<b>jungle rice</b>		<b>X</b>	<b>X</b>					<b>X</b>		
	<i>Elionurus barbiculmis</i> Hack.	woolyspike balsamscale										X
	<i>Elymus elymoides</i> (Raf.) Swezey	squirreltail		X		X				X		X
	<i>Elymus elymoides</i> ssp. <i>elymoides</i> (Raf.) Swezey	squirreltail	X	X								
	<i>Elyonurus barbiculmus</i> Hack.		X	X			X			X		
	<i>Enneapogon desvauxii</i> Desv. ex Beauv.	nineawn pappusgrass				X	X					
	<b><i>Eragrostis cilianensis</i> (All.) Vign. ex Janchen</b>	<b>stinkgrass</b>	<b>X</b>	<b>X</b>	<b>X</b>		<b>X</b>			<b>X</b>		
	<b><i>Eragrostis curvula</i> (Schrud.) Nees</b>	<b>weeping lovegrass</b>		<b>X</b>	<b>X</b>		<b>X</b>			<b>X</b>		
	<i>Eragrostis intermedia</i> A.S. Hitchc.	plains lovegrass	X	X		X	X			X	X	X
	<b><i>Eragrostis lehmanniana</i> Nees</b>	<b>Lehmann lovegrass</b>	<b>X</b>	<b>X</b>	<b>X</b>	<b>X</b>	<b>X</b>			<b>X</b>		<b>X</b>
	<i>Eragrostis mexicana</i> (Hornem.) Link	Mexican lovegrass	X							X		
	<i>Eragrostis mexicana</i> ssp. <i>mexicana</i> (Hornem.) Link	Mexican lovegrass		X		X				X		
	<i>Eragrostis pectinacea</i> (Michx.) Nees ex Steud.	tufted lovegrass				X						
	<i>Eragrostis pectinacea</i> var. <i>miserrima</i> (Fourn.) J. Reeder	desert lovegrass								X		
	<i>Eriochloa acuminata</i> (J. Presl) Kunth	tapertip cupgrass		X			X					
	<i>Eriochloa acuminata</i> var. <i>acuminata</i> (J. Presl) Kunth	tapertip cupgrass	X	X								
	<i>Eriochloa lemmonii</i> Vasey & Scribn.	canyon cupgrass	X	X								
	<i>Erioneuron avenaceum</i> (Kunth) Tateoka	shortleaf woollygrass		X			X			X		
	<b><i>Hackelochloa granularis</i> (L.) Kuntze</b>	<b>pitscale grass</b>					<b>X</b>					
	<i>Hesperostipa neomexicana</i> (Thurb. ex Coult.) Barkworth	New Mexico feathergrass		X						X		
	<i>Heteropogon contortus</i> (L.) Beauv. ex Roemer & J.A. Schultes	tanglehead	X	X		X	X			X		X
	<i>Hilaria belangeri</i> (Steud.) Nash	curly-mesquite	X	X		X						
	<b><i>Hordeum murinum</i> ssp. <i>glaucom</i> (Steud.) Tzvelev</b>	<b>smooth barley</b>		<b>X</b>								
	<i>Koeleria macrantha</i> (Ledeb.) J.A. Schultes	prairie Junegrass				X						
	<i>Leptochloa dubia</i> (Kunth) Nees	green sprangletop	X	X		X	X				X	
	<b><i>Lolium perenne</i> L.</b>	<b>perennial ryegrass</b>								<b>X</b>		
	<i>Lycurus phleoides</i> Kunth	common wolfstail				X						
	<i>Lycurus setosus</i> (Nutt.) C.G. Reeder	bristly wolfstail	X	X			X				X	X
	<i>Muhlenbergia arizonica</i> Scribn.	Arizona muhly	X	X						X		X
	<i>Muhlenbergia emersleyi</i> Vasey	bullgrass	X	X		X	X			X		X
	<i>Muhlenbergia fragilis</i> Swallen	delicate muhly		X			X					
	<i>Muhlenbergia glauca</i> (Nees) B.D. Jackson	desert muhly		X						X		X
	<i>Muhlenbergia longiligula</i> A.S. Hitchc.	longtongue muhly		X								
	<i>Muhlenbergia minutissima</i> (Steud.) Swallen	annual muhly	X	X								

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			R&J	P&C	H&G	Ruy	NPS	UA	ASU	DBG	NAU	UA
Poaceae												
	<i>Muhlenbergia montana</i> (Nutt.) A.S. Hitchc.	mountain muhly	X	X		X						
	<i>Muhlenbergia pauciflora</i> Buckl.	New Mexico muhly		X						X		
	<i>Muhlenbergia porteri</i> Scribn. ex Beal	bush muhly	X	X		X						
	<i>Muhlenbergia repens</i> (J. Presl) A.S. Hitchc.	creeping muhly				X						
	<i>Muhlenbergia richardsonis</i> (Trin.) Rydb.	mat muhly				X						
	<i>Muhlenbergia rigens</i> (Benth.) A.S. Hitchc.	deergrass	X	X						X		
	<i>Muhlenbergia rigida</i> (Kunth) Trin.	purple muhly	X	X								X
	<i>Muhlenbergia sinuosa</i> Swallen	marshland muhly										X
	<i>Muhlenbergia tenuifolia</i> (Kunth) Trin.	slimflower muhly		X			X			X		X
	<i>Panicum bulbosum</i> Kunth	bulb panicgrass		X			X			X	X	
	<i>Panicum capillare</i> L.	witchgrass	X	X								
	<i>Panicum hallii</i> Vasey	Hall's panicgrass	X	X			X			X		
	<i>Panicum hirticaule</i> J. Presl	Mexican panicgrass	X	X			X					
	<i>Panicum hirticaule</i> var. <i>hirticaule</i> J. Presl	Mexican panicgrass								X		
	<i>Panicum obtusum</i> Kunth	vine mesquite	X	X		X				X		X
	<i>Pappophorum</i> Schreb.	pappusgrass				X						
	<i>Paspalum setaceum</i> Michx.	thin paspalum								X		
	<i>Piptochaetium fimbriatum</i> (Kunth) A.S. Hitchc.	pinyon ricegrass	X	X		X	X			X		
	<i>Piptochaetium pringlei</i> (Beal) Parodi	Pringle's speargrass									X	
	<i>Pleuraphis mutica</i> Buckl.	tobosagrass	X	X								
	<i>Poa fendleriana</i> (Steud.) Vasey	muttongrass								X		X
	<i>Poa fendleriana</i> (Steud.) Vasey ssp. <i>fendleriana</i>	muttongrass		X						X		
	<b><i>Polypogon monspeliensis</i> (L.) Desf</b>	<b>annual rabbitsfoot grass</b>			X							
	<i>Schizachyrium cirratum</i> (Hack.) Woot. & Standl.	Texas bluestem	X	X		X	X			X		X
	<i>Schizachyrium sanguineum</i> (Retz.) Alston	crimson bluestem		X			X			X		
	<i>Schizachyrium scoparium</i> (Michx.) Nash var. <i>scoparium</i>	little bluestem	X	X								
	<i>Scleropogon brevifolius</i> Phil.	burrograss	X	X								
	<i>Setaria grisebachii</i> Fourn.	Grisebach's bristlegrass	X	X			X			X	X	X
	<i>Setaria leucopila</i> (Scribn. & Merr.) K. Schum.	streambed bristlegrass		X						X		
	<b><i>Sorghum halepense</i> (L.) Pers.</b>	<b>Johnsongrass</b>	X	X	X					X		
	<i>Sporobolus airoides</i> (Torr.) Torr.	alkali sacaton	X	X								
	<i>Sporobolus cryptandrus</i> (Torr.) Gray	sand dropseed	X	X			X					
	<i>Sporobolus wrightii</i> Munro ex Scribn.	big sacaton	X	X						X		
	<i>Trachypogon spicatus</i> (L.) Kuntze	spiked crinkleawn	X	X		X	X			X		
	<i>Tridens muticus</i> (Torr.) Nash	slim tridens	X			X						
	<i>Tridens muticus</i> var. <i>muticus</i> (Torr.) Nash	slim tridens		X						X		
	<i>Tripsacum lanceolatum</i> Rupr. ex Fourn.	Mexican gamagrass	X	X			X			X		X
	<b><i>Triticum aestivum</i> L.</b>	<b>common wheat</b>		X						X		
	<i>Urochloa arizonica</i> (Scribn. & Merr.) O. Morrone & F. Zuloaga	Arizona signalgrass	X	X			X			X		

Family	Studies							Specimens in herbaria				
	Scientific name	Common name	R&J	P&C	H&G	Ruy	NPS	UA	ASU	DBG	NAU	UA
Poaceae												
	<i>Vulpia octoflora</i> var. <i>hirtella</i> (Piper) Henr.	sixweeks fescue		X						X		
	<i>Vulpia octoflora</i> var. <i>octoflora</i> (Walt.) Rydb.	sixweeks fescue		X						X		X
Polemoniaceae												
	<i>Eriastrum diffusum</i> (Gray) Mason	miniature woollystar		X						X		
	<i>Gilia flavocincta</i> A. Nels.	lesser yellowthroat gilia		X								
	<i>Gilia flavocincta</i> ssp. <i>australis</i> (A. & V. Grant) Day & V. Grant	lesser yellowthroat gilia								X		
	<i>Gilia mexicana</i> A. & V. Grant	El Paso gilia		X						X		
	<i>Ipomopsis longiflora</i> (Torr.) V. Grant	flaxflowered ipomopsis	X	X								
	<i>Ipomopsis thurberi</i> (Torr. ex Gray) V. Grant	El Paso skyrocket	X	X						X	X	
	<i>Phlox stansburyi</i> (Torr.) Heller ssp. <i>stansburyi</i>	cold-desert phlox	X	X								
Polygalaceae												
	<i>Polygala hemipterocarpa</i> Gray	winged milkwort										X
	<i>Polygala obscura</i> Benth.	velvetseed milkwort	X	X								
Polygonaceae												
	<i>Eriogonum abertianum</i> Torr.	Abert's buckwheat	X	X							X	
	<i>Eriogonum palmerianum</i> Reveal	Palmer's buckwheat	X	X								
	<i>Eriogonum polycladon</i> Benth.	sorrel buckwheat		X								
	<i>Eriogonum wrightii</i> Torr. ex Benth.	bastardsage	X	X		X						
	<i>Polygonum aviculare</i> L.	prostate knotweed			X							
Portulacaceae												
	<i>Calandrinia ciliata</i> (Ruiz & Pavón) DC.	fringed redmaids								X		
	<i>Cistanthe ambigua</i> (S. Wats.) Hershkovitz	desert pussypaws		X								
	<i>Portulaca oleracea</i> L.	little hogweed	X	X								
	<i>Portulaca pilosa</i> L.	kiss me quick	X	X								
	<i>Portulaca suffrutescens</i> Engelm.	shrubby purslane	X	X								
	<i>Portulaca umbraticola</i> Kunth	wingpod purslane	X	X								
	<i>Talinum aurantiacum</i> Engelm.	orange fameflower	X	X								X
	<i>Talinum brevicaule</i> S. Wats.	dwarf fameflower		X								
	<i>Talinum paniculatum</i> (Jacq.) Gaertn.	jewels of Opar		X								
	<i>Talinum parviflorum</i> Nutt.	sunbright		X						X		
Pteridaceae												
	<i>Argyroschisma limitanea</i> (Maxon) Windham	southwestern false cloak fern								X		
	<i>Argyroschisma limitanea</i> ssp. <i>limitanea</i> (Maxon) Windham	southwestern false cloakfern		X								
	<i>Astrolepis cochisensis</i> ssp. <i>cochisensis</i> (Goodding) Benham & Windham	Cochise scaly cloakfern								X		
	<i>Astrolepis integerrima</i> (Hook.) Benham & Windham	hybrid cloakfern		X						X		
	<i>Astrolepis sinuata</i> (Lag. ex Sw.) Benham & Windham	wavy scaly cloakfern								X		
	<i>Astrolepis sinuata</i> (Lag. ex Sw.) Benham & Windham ssp. <i>sinuata</i>	wavy scaly cloakfern	X	X								
	<i>Bommeria hispida</i> (Mett. ex Kuhn) Underwood	copper fern		X						X		

Family	Studies							Specimens in herbaria				
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Pteridaceae												
	<i>Cheilanthes bonariensis</i> (Willd.) Proctor	golden lipfern		X						X		
	<i>Cheilanthes eatonii</i> Baker	Eaton's lipfern		X						X		
	<i>Cheilanthes fendleri</i> Hook.	Fendler's lipfern		X						X		
	<i>Cheilanthes lendigera</i> (Cav.) Sw.	nitbearing lipfern		X								
	<i>Cheilanthes lindheimeri</i> Hook.	fairyswords		X						X		X
	<i>Cheilanthes tomentosa</i> Link	woolly lipfern		X						X		
	<i>Cheilanthes villosa</i> Davenport ex Maxon	villous lipfern		X						X		
	<i>Cheilanthes wootonii</i> Maxon	beaded lipfern	X	X						X		X
	<i>Cheilanthes wrightii</i> Hook.	Wright's lipfern		X						X		
	<i>Notholaena grayi</i> Davenport	Gray's cloak fern		X						X		
	<i>Notholaena standleyi</i> Maxon	star cloak fern	X	X						X		
	<i>Pellaea atropurpurea</i> (L.) Link	purple cliffbrake		X						X		
Ranunculaceae												
	<i>Anemone tuberosa</i> Rydb.	tuber anemone		X						X		X
	<i>Thalictrum fendleri</i> Engelm. ex Gray	Fendler's meadow-rue	X	X							X	X
Rhamnaceae												
	<i>Ceanothus fendleri</i> Gray	Fendler's ceanothus	X	X						X		
	<i>Ceanothus greggii</i> Gray	desert ceanothus	X	X								
	<i>Ziziphus obtusifolia</i> var. <i>canescens</i> (Gray) M.C. Johnston	lotebush	X	X								
Rosaceae												
	<i>Cercocarpus montanus</i> Raf.	alderleaf mountain mahogany					X					
	<i>Cercocarpus montanus</i> var. <i>glaber</i> (S. Wats.) F.L. Martin	birchleaf mountain mahogany	X	X								
	<i>Cercocarpus montanus</i> var. <i>paucidentatus</i> (S. Wats.) F.L. Martin	hairy mountain mahogany	X	X								X
	<b><i>Prunus armeniaca</i> L.</b>	<b>apricot</b>	<b>X</b>	<b>X</b>						<b>X</b>		
	<i>Prunus serotina</i> Ehrh.	black cherry		X						X		
	<i>Prunus serotina</i> var. <i>rufula</i> (Woot. & Standl.) McVaugh	black cherry										X
	<i>Prunus serotina</i> var. <i>virens</i> (Woot. & Standl.) McVaugh	black cherry	X									
	<i>Purshia stansburiana</i> (Torr.) Henrickson	Stansbury cliffrose	X	X						X		
	<b><i>Pyracantha koidzumii</i> (Hayata) Rehd.</b>	<b>Formosa firethorn</b>	<b>X</b>	<b>X</b>						<b>X</b>		
	<i>Vauquelinia californica</i> (Torr.) Sarg.	Arizona rosewood	X	X								
Rubiaceae												
	<i>Bouvardia ternifolia</i> (Cav.) Schlecht.	firecrackerbush	X	X			X			X	X	
	<i>Crusea diversifolia</i> (Kunth) W.A. Anderson	mountain saucerflower		X							X	
	<i>Diodia teres</i> Walt.	poorjoe		X						X		
	<i>Galium aparine</i> L.	stickywilly	X									
	<i>Galium mexicanum</i> ssp. <i>asperrimum</i> (Gray) Dempster	Mexican bedstraw		X						X		
	<i>Galium microphyllum</i> Gray	bracted bedstraw	X	X						X		
	<i>Galium wrightii</i> Gray	Wright's bedstraw		X						X	X	
	<i>Houstonia wrightii</i> Gray	pygmy bluet								X		
	<i>Mitracarpus breviflorus</i> Gray	white girdlepod	X	X						X		

Family	Scientific name	Common name	Studies						Specimens in herbaria			
			R&J	P&C	H&G	Ruy	NPS	UA	ASU	DBG	NAU	UA
Rutaceae												
	<i>Ptelea trifoliata</i> ssp. <i>angustifolia</i> (Benth.) V. Bailey	common hoptree		X								
Salicaceae												
	<i>Populus fremontii</i> S. Wats. ssp. <i>fremontii</i>	Fremont cottonwood	X	X								
	<i>Salix gooddingii</i> Ball	Goodding's willow	X	X				X		X		
Santalaceae												
	<i>Comandra umbellata</i> ssp. <i>pallida</i> (A. DC.) Piehl	pale bastard toadflax		X						X		
Sapindaceae												
	<i>Sapindus saponaria</i> L.	wingleaf soapberry						X				
	<i>Sapindus saponaria</i> var. <i>drummondii</i> (Hook. & Arn.) L. Benson	western soapberry	X	X						X		
Saxifragaceae												
	<i>Heuchera sanguinea</i> Engelm.	coralbelles	X	X						X		
Scrophulariaceae												
	<i>Brachystigma wrightii</i> (Gray) Pennell	Arizona desert foxglove	X	X								
	<i>Castilleja integra</i> Gray	wholeleaf Indian paintbrush		X							X	
	<i>Castilleja lanata</i> Gray	Sierra woolly Indian paintbrush		X								
	<i>Castilleja tenuiflora</i> Benth.	Santa Catalina Indian paintbrush	X	X						X		
	<i>Maurandella antirrhiniflora</i> (Humb. & Bonpl. ex Willd.) Rothm.	roving sailor		X						X	X	
	<i>Mimulus guttatus</i> DC.	seep monkeyflower	X	X						X		
	<i>Mimulus rubellus</i> Gray	little redstem monkeyflower		X						X		
	<i>Nuttallanthus texanus</i> (Scheele) D.A. Sutton	Texas toadflax		X						X		
	<i>Penstemon barbatus</i> (Cav.) Roth	beardlip penstemon		X						X		
	<i>Penstemon dasyphyllus</i> Gray	Cochise beardtongue	X	X								
	<i>Penstemon parryi</i> (Gray) Gray	Parry's beardtongue	X	X		X				X		
	<i>Penstemon stenophyllus</i> (Gray) T.J. Howell	Sonoran beardtongue		X						X		
	<i>Penstemon superbus</i> A. Nels.	superb beardtongue		X								
	<i>Penstemon virgatus</i> Gray	upright blue beardtongue	X	X								
	<i>Schistophragma intermedia</i> (Gray) Pennell	harlequin spiralseed									X	
	<b><i>Verbascum thapsus</i> L.</b>	<b>common mullein</b>			X							
	<i>Veronica peregrina</i> ssp. <i>xalapensis</i> (Kunth) Pennell	hairy purslane speedwell		X						X		
Simaroubaceae												
	<b><i>Ailanthus altissima</i> (P. Mill.) Swingle</b>	<b>tree of heaven</b>		X						X		
Solanaceae												
	<b><i>Datura quercifolia</i> Kunth</b>	<b>Chinese thorn-apple</b>		X						X		
	<b><i>Datura stramonium</i> L.</b>	<b>jimsonweed</b>	X	X								
	<i>Datura wrightii</i> Regel	sacred thorn-apple	X	X						X		
	<i>Margaranthus solanaceus</i> Schlecht.	netted globecherry		X						X		
	<i>Nicotiana obtusifolia</i> Mertens & Galeotti	desert tobacco								X		
	<i>Nicotiana obtusifolia</i> var. <i>obtusifolia</i> Mertens & Galeotti	desert tobacco		X								
	<i>Physalis hederifolia</i> var. <i>fendleri</i> (Gray) Cronq.	Fendler's groundcherry				X						

Family	Scientific name	Common name	Studies						Specimens in herbaria			
			R&J	P&C	H&G	Ruy	NPS	UA	ASU	DBG	NAU	UA
Solanaceae												
	<i>Physalis longifolia</i> Nutt. var. <i>longifolia</i>	longleaf groundcherry		X						X		
	<i>Solanum adscendens</i> Sendtner	sonoita nightshade	X	X						X		
	<i>Solanum americanum</i> P. Mill.	American black nightshade		X						X		
	<i>Solanum douglasii</i> Dunal	greenspot nightshade		X						X		
	<i>Solanum elaeagnifolium</i> Cav.	silverleaf nightshade	X	X						X		
	<i>Solanum jamesii</i> Torr.	wild potato										X X
	<i>Solanum rostratum</i> Dunal	buffalobur nightshade		X						X		
Sterculiaceae												
	<i>Ayenia compacta</i> Rose	California ayenia	X	X								
	<i>Ayenia filiformis</i> S. Wats.	TransPecos ayenia	X							X		
Ulmaceae												
	<i>Celtis laevigata</i> var. <i>reticulata</i> (Torr.) L. Benson	netleaf hackberry	X	X								
Urticaceae												
	<i>Parietaria hespera</i> Hinton	rillita pellitory		X								X
Valerianaceae												
	<i>Valeriana arizonica</i> Gray	Arizona valerian	X	X								
	<i>Valeriana sorbifolia</i> Kunth	pineland valerian		X						X		
Verbenaceae												
	<i>Aloysia wrightii</i> Heller ex Abrams	Wright's beebrush	X	X		X	X			X		
	<i>Bouchea prismatica</i> (L.) Kuntze	prism bouchea	X	X							X	
	<i>Glandularia bipinnatifida</i> (Nutt.) Nutt.	Dakota mock vervain	X									
	<i>Glandularia bipinnatifida</i> var. <i>bipinnatifida</i> (Nutt.) Nutt.	Dakota mock vervain		X						X	X	
	<i>Glandularia bipinnatifida</i> var. <i>ciliata</i> (Benth.) B.L. Turner	Dakota mock vervain		X								
	<i>Tetradlea coulteri</i> Gray	Coulter's wrinklefruit		X						X		
	<i>Verbena bracteata</i> Lag. & Rodr.	bigbract verbena	X	X								
Viscaceae												
	<i>Phoradendron capitellatum</i> Torr. ex Trel.	downy mistletoe	X	X								
	<i>Phoradendron coryae</i> Trel.	Cory's mistletoe	X	X							X	
	<i>Phoradendron juniperinum</i> Engelm. ex Gray	juniper mistletoe	X	X								
	<i>Phoradendron villosum</i> (Nutt.) Nutt.	Pacific mistletoe					X					
Vitaceae												
	<i>Vitis arizonica</i> Engelm.	canyon grape	X	X								X
Zygophyllaceae												
	<i>Kallstroemia parviflora</i> J.B.S. Norton	warty caltrop	X	X								
	<i>Tribulus terrestris</i> L.	puncturevine		X	X							

**Appendix B. Species list of amphibians and reptiles for Coronado NM.** All species listed have been confirmed present at the memorial either through a voucher specimen or photograph.

Taxon	Family	Scientific name	Common name
Amphibian	Ambystomatidae	<i>Ambystoma mavortium</i>	barred tiger salamander
	Leptodactylidae	<i>Eleutherodactylus augusti</i> <sup>a</sup>	barking frog
	Pelobatidae	<i>Spea multiplicata</i>	New Mexico spadefoot
	Bufonidae	<i>Bufo woodhousii</i>	Woodhouse's toad
		<i>Bufo punctatus</i>	red-spotted toad
		<i>Bufo cognatus</i>	Great Plains toad
	Hylidae	<i>Hyla arenicolor</i>	canyon treefrog
Reptile	Emydidae	<i>Terrapene ornata</i>	ornate box turtle
	Crotaphytidae	<i>Crotaphytus collaris</i>	eastern collared lizard
	Phrynosomatidae	<i>Holbrookia maculata</i>	lesser earless lizard
		<i>Sceloporus slevini</i>	bunch grass lizard
		<i>Sceloporus jarrovi</i>	Yarrow's spiny lizard
		<i>Sceloporus clarkii</i>	Clark's spiny lizard
		<i>Sceloporus undulatus</i>	prairie lizard
		<i>Urosaurus ornatus</i>	tree lizard
		<i>Phrynosoma douglasii</i>	short-horned lizard
	Scincidae	<i>Eumeces obsoletus</i>	Great Plains skink
		<i>Eumeces callicephalus</i>	mountain skink
	Teiidae	<i>Cnemidophorus uniparens</i>	desert grassland whiptail
		<i>Cnemidophorus sonora</i>	Sonoran spotted whiptail
	Anguidae	<i>Elgaria kingii</i>	Madrean alligator lizard
	Helodermatidae	<i>Heloderma suspectum</i>	Gila monster
	Leptotyphlopidae	<i>Leptotyphlops dulcis</i>	Texas blind snake
	Colubridae	<i>Diadophis punctatus</i>	ringneck snake
		<i>Heterodon nasicus</i>	western hognose snake
		<i>Masticophis flagellum</i>	coachwhip
		<i>Masticophis bilineatus</i>	Sonoran whipsnake
		<i>Salvadora deserticola</i>	Big Bend patchnose snake
		<i>Salvadora grahamiae</i>	mountain patchnose snake
		<i>Pituophis catenifer</i>	gopher snake
		<i>Lampropeltis getula</i>	common kingsnake
		<i>Lampropeltis pyromelana</i>	Sonoran mountain kingsnake
		<i>Rhinocheilus lecontei</i>	longnose snake
		<i>Thamnophis cyrtopsis</i>	blackneck garter snake
		<i>Gyalopion canum</i>	western hooknose snake
		<i>Tantilla wilcoxi</i>	Chihuahuan blackhead snake
		<i>Trimorphodon biscutatus</i>	lyre snake
		<i>Hypsiglena torquata</i>	night snake
	Elapidae	<i>Micruroides euryxanthus</i>	western coral snake
	Viperidae	<i>Crotalus atrox</i>	western diamondback rattlesnake
		<i>Crotalus lepidus</i>	rock rattlesnake
		<i>Crotalus molossus</i>	blacktail rattlesnake
		<i>Crotalus scutulatus</i>	Mojave rattlesnake

<sup>a</sup> USFS = U.S. Forest Service: Sensitive species; Arizona Game and Fish Department: Wildlife Species of Concern (HDMS 2003).



**Appendix C. Number of observations, by bird species and detection type, at Coronado NM by University of Arizona (UA) Inventory personnel, 2002–2004.** Numbers of individuals recorded are not scaled by search effort and should not be used for comparison among species. List also includes species reported to the visitor center log book, those observed by Russell and Danforth (R&D; 1979) and those observed by NPS Monitoring personnel in 2005 (unpublished data). The species list at the visitor center includes data from the MAPS effort in the oak woodland (see text for more details). Underlined species are neotropical migrants (Rappole 1995). Species in bold-faced type are non-native.

Order				UA Detection type		Other efforts			Conservation Designations					
	Family							NPS						
	Scientific name	Common name	VCP	Winter	Incidental	Nocturnal	Visitor center	R&D	Monitoring	ESA <sup>a</sup>	US FS <sup>b</sup>	AZ G&F <sup>c</sup>	APF <sup>d</sup>	US FWS <sup>e</sup>
<b>Galliformes</b>														
Phasianidae														
	<i>Meleagris gallopavo</i>	wild turkey			4			X						
Odontophoridae														
	<i>Callipepla squamata</i>	scaled quail			2		X	X						
	<i>Callipepla gambelii</i>	Gambel's quail	1				X	X						
	<i>Cyrtonyx montezumae</i>	Montezuma quail	1	1	6		X	X	X					
<b>Ciconiiformes</b>														
Cathartidae														
	<i>Coragyps atratus</i>	<u>black vulture</u>					X							
	<i>Cathartes aura</i>	<u>turkey vulture</u>	84		7		X	X	X					
<b>Falconiformes</b>														
Accipitridae														
	<i>Haliaeetus leucocephalus</i>	bald eagle					X			LT	S	WSC		
	<i>Circus cyaneus</i>	<u>northern harrier</u>		2	2		X	X						
	<i>Elanus leucurus</i>	<u>white-tailed kite</u>					X							
	<i>Accipiter striatus</i>	<u>sharp-shinned hawk</u>		1	1		X	X			S			
	<i>Accipiter cooperii</i>	<u>Cooper's hawk</u>	9		2		X	X	X					
	<i>Accipiter gentilis</i>	northern goshawk					X			SC	S	WSC		
	<i>Buteogallus anthracinus</i>	<u>common black-hawk</u>					X				S	WSC	P	
	<i>Parabuteo unicinctus</i>	Harris's hawk					X							
	<i>Buteo swainsoni</i>	<u>Swainson's hawk</u>			1		X							
	<i>Buteo albonotatus</i>	zone-tailed hawk	1		1		X		X					
	<i>Buteo jamaicensis</i>	<u>red-tailed hawk</u>	4		2		X	X	X					
	<i>Aquila chrysaetos</i>	golden eagle	1				X	X						
Falconidae														
	<i>Falco sparverius</i>	<u>American kestrel</u>			2		X	X						
	<i>Falco columbarius</i>	<u>merlin</u>					X							
	<i>Falco peregrinus</i>	<u>peregrine falcon</u>			1		X			SC		WSC		BCC
	<i>Falco mexicanus</i>	<u>prairie falcon</u>	4		1		X	X						
<b>Charadriiformes</b>														
Scolopacidae														
	<i>Gallinago gallinago</i>	<u>common snipe</u>					X							
<b>Columbiformes</b>														
Columbidae														
	<b><i>Columba livia</i></b>	<b>rock pigeon</b>			1									
	<i>Patagioenas fasciata</i>	<u>band-tailed pigeon</u>					X	X	X					
	<i>Zenaida asiatica</i>	<u>white-winged dove</u>	36		1		X	X	X					
	<i>Zenaida macroura</i>	<u>mourning dove</u>	37		1		X	X	X					
	<i>Columbina inca</i>	Inca dove					X							
	<i>Columbina passerina</i>	common ground-dove			1		X							

Order				UA Detection type			Other efforts			Conservation Designations				
	Family								NPS					
	Scientific name	Common name	VCP	Winter	dental	Noc- turnal	Visitor center	R&D	Moni- toring	ESA <sup>a</sup>	US FS <sup>b</sup>	AZ G&F <sup>c</sup>	APF <sup>d</sup>	US FWS <sup>e</sup>
<b>Cuculiformes</b>														
Cuculidae														
	<i>Coccyzus americanus</i>	yellow-billed cuckoo	1							C	S	WSC	P	BCC
	<i>Geococcyx californianus</i>	greater roadrunner	13		1		X	X	X					
<b>Strigiformes</b>														
Tytonidae														
	<i>Tyto alba</i>	barn owl					X							
Strigidae														
	<i>Otus flammeolus</i>	flamulated owl					X							
	<i>Megascops kennicottii</i>	western screech-owl			1	1	X	X	X					
	<i>Megascops trichopsis</i>	whiskered screech-owl			2	1	X	X						
	<i>Bubo virginianus</i>	great horned owl			1	1	X	X						
	<i>Glaucidium gnoma</i>	northern pygmy-owl				1	X							
	<i>Strix occidentalis lucida</i>	Mexican spotted owl							X	T	S	WSC		
	<i>Micrathene whitneyi</i>	elf owl			1	1	X	X	X					BCC
<b>Caprimulgiformes</b>														
Caprimulgidae														
	<i>Chordeiles minor</i>	common nighthawk					X	X	X					
	<i>Phalaenoptilus nuttallii</i>	common poorwill			1	1	X	X	X					
	<i>Caprimulgus vociferus</i>	whip-poor-will				1	X	X	X					
<b>Apodiformes</b>														
Apodidae														
	<i>Aeronautes saxatalis</i>	white-throated swift	92				X	X	X					
Trochilidae														
	<i>Heliomaster constantii</i>	plain-capped starthroat					X							
	<i>Cynanthus latirostris</i>	broad-billed hummingbird	1		1		X							
	<i>Hylocharis leucotis</i>	white-eared hummingbird					X							
	<i>Lampornis clemenciae</i>	blue-throated hummingbird					X							
	<i>Eugenes fulgens</i>	magnificent hummingbird			1		X	X						
	<i>Calothorax lucifer</i>	Lucifer's hummingbird			1		X	X						
	<i>Archilochus alexandri</i>	black-chinned hummingbird	7		2		X	X						
	<i>Calypte anna</i>	Anna's hummingbird	5		1		X	X	X					
	<i>Calypte costae</i>	Costa's hummingbird					X						P	
	<i>Amazilia violiceps</i>	violet-crowned hummingbird					X			S	WSC			
	<i>Stellula calliope</i>	calliope hummingbird			1		X							
	<i>Selasphorus platycercus</i>	broad-tailed hummingbird	6		1		X	X	X					
	<i>Selasphorus sasin</i>	Allen's hummingbird					X							
	<i>Selasphorus rufus</i>	rufous hummingbird	2		1		X	X						
<b>Trogoniformes</b>														
Trogonidae														
	<i>Trogon elegans</i>	elegant trogon			1		X	X				WSC		
<b>Piciformes</b>														
Picidae														
	<i>Melanerpes lewis</i>	Lewis's woodpecker					X							
	<i>Melanerpes formicivorus</i>	acorn woodpecker	1		1		X	X	X					
	<i>Melanerpes uropygialis</i>	Gila woodpecker			1		X	X						BCC
	<i>Sphyrapicus thyroideus</i>	Williamson's sapsucker						X						

Order	UA Detection type				Other efforts			Conservation Designations						
	Family						NPS							
	Scientific name	Common name	VCP	Winter	Inci- dental	Noc- turnal	Visitor center	R&D	Moni- toring	ESA <sup>a</sup>	US FS <sup>b</sup>	AZ G&F <sup>c</sup>	APF <sup>d</sup>	US FWS <sup>e</sup>
Piciformes														
Picidae														
	<i>Sphyrapicus nuchalis</i>	red-naped sapsucker		1	1		X							
	<i>Sphyrapicus ruber</i>	red-breasted sapsucker					X							
	<i>Sphyrpicus varius</i>	yellow-bellied sapsucker					X	X						
	<i>Picoides scalaris</i>	ladder-backed woodpecker	12	1	2		X		X					
	<i>Picoides villosus</i>	hairy woodpecker			1									
	<i>Picoides arizonae</i>	Arizona woodpecker	13		1		X	X						
	<i>Colaptes auratus</i>	northern flicker	13	5	1		X	X	X					
	<i>Colaptes chrysoides</i>	gilded flicker					X						P	BCC
Passeriformes														
Tyrannidae														
	<i>Contopus pertinax</i>	<u>greater pewee</u>			1		X							
	<i>Contopus sordidulus</i>	<u>western wood-pewee</u>	3		1			X	X					
	<i>Empidonax traillii</i>	<u>willow flycatcher</u>					X					WSC		
	<i>Empidonax hammondii</i>	<u>Hammond's flycatcher</u>	9		1		X	X	X					
	<i>Empidonax wrightii</i>	<u>gray flycatcher</u>	3		1		X	X	X					
	<i>Empidonax oberholseri</i>	<u>dusky flycatcher</u>			1		X	X						
	<i>Empidonax occidentalis</i> or <i>difficilis</i>	<u>western flycatcher</u>	2		2		X	X						
	<i>Empidonax occidentalis</i>	<u>cordilleran flycatcher</u>	1											
	<i>Sayornis nigricans</i>	black phoebe	1				X							
	<i>Sayornis saya</i>	<u>Say's phoebe</u>		2	3		X	X	X					
	<i>Pyrocephalus rubinus</i>	<u>vermillion flycatcher</u>					X		X					
	<i>Myiarchus tuberculifer</i>	<u>dusky-capped flycatcher</u>	59		2		X	X	X					
	<i>Myiarchus cinerascens</i>	<u>ash-throated flycatcher</u>	82		1		X	X	X					
	<i>Myiarchus tyrannulus</i>	<u>brown-crested flycatcher</u>					X	X						
	<i>Myiodynastes luteiventris</i>	<u>sulphur-bellied flycatcher</u>	14		3		X	X	X					
	<i>Tyrannus vociferans</i>	<u>Cassin's kingbird</u>	21		4		X	X	X					
	<i>Tyrannus verticalis</i>	<u>western kingbird</u>	3		3		X	X						
Laniidae														
	<i>Lanius ludovicianus</i>	<u>loggerhead shrike</u>	1		2		X	X		SC	S			
Vireonidae														
	<i>Vireo bellii</i>	<u>Bell's vireo</u>	1		1		X				S			BCC
	<i>Vireo plumbeus</i>	<u>plumbeous vireo</u>	7		1									
	<i>Vireo cassinii</i>	<u>Cassin's vireo</u>			1									
	<i>Vireo plumbeus</i> or <i>V. cassinii</i>	<u>solitary vireo type</u>			1		X	X						
	<i>Vireo huttoni</i>	Hutton's vireo	24	3	2		X	X						
	<i>Vireo gilvus</i>	<u>warbling vireo</u>			1		X	X	X					
Corvidae														
	<i>Cyanocitta stelleri</i>	Steller's jay			1		X	X						
	<i>Aphelocoma californica</i>	western scrub-jay	1				X	X						
	<i>Aphelocoma ultramarina</i>	Mexican jay	201	46	5		X	X	X					
	<i>Gymnorhinus cyanocephalus</i>	pinyon Jay			1		X							
	<i>Corvus</i> sp.	unknown raven	39	3	1				X					
	<i>Corvus cryptoleucus</i>	Chihuahuan raven					X	X						
	<i>Corvus corax</i>	common raven						X	X					

Order	Family	Scientific name	Common name	UA Detection type				Other efforts		Conservation Designations						
				VCP	Winter	Incidental	Noc-turnal	Visitor center	R&D	NPS Monitoring	ESA <sup>a</sup>	US FS <sup>b</sup>	AZ G&F <sup>c</sup>	APF <sup>d</sup>	US FWS <sup>e</sup>	
Passeriformes																
	Alaudidae															
		<i>Eremophila alpestris</i>	horned lark						X							
	Hirundinidae															
		<i>Tachycineta thalassina</i>	<u>violet-green swallow</u>	3				X	X							
		<i>Hirundo rustica</i>	<u>barn swallow</u>	1				X								
	Paridae															
		<i>Baeolophus wollweberi</i>	bridled titmouse	56	8	2		X	X	X						
	Remizidae															
		<i>Auriparus flaviceps</i>	verdin	18	5			X	X	X						
	Aegithalidae															
		<i>Psaltiriparus minimus</i>	bushtit	50	53	10		X	X	X						
	Sittidae															
		<i>Sitta carolinensis</i>	white-breasted nuthatch	3	5	1		X	X	X						
	Certhiidae															
		<i>Certhia americana</i>	brown creeper					X								
	Troglodytidae															
		<i>Campylorhynchus</i>														
		<i>brunneicapillus</i>	cactus wren	11		2		X	X	X						
		<i>Salpinctes obsoletus</i>	rock wren			1		X	X	X						
		<i>Catherpes mexicanus</i>	canyon wren	11	1	1		X	X							
		<i>Thryomanes bewickii</i>	Bewick's wren	177	14	4		X	X	X						
		<i>Troglodytes aedon</i>	<u>house wren</u>	1	1	1		X	X							
	Regulidae															
		<i>Regulus calendula</i>	<u>ruby-crowned kinglet</u>	19	30	1		X	X							
	Sylviidae															
		<i>Poliophtila caerulea</i>	<u>blue-gray gnatcatcher</u>	1				X	X							
		<i>Poliophtila melanura</i>	black-tailed gnatcatcher							X						
	Turdidae															
		<i>Sialia sialis</i>	<u>eastern bluebird</u>			2		X	X							
		<i>Sialia mexicana</i>	<u>western bluebird</u>		12	12		X	X							
		<i>Sialia currucoides</i>	<u>mountain bluebird</u>		2	4		X								
		<i>Myadestes townsendi</i>	<u>Townsend's solitaire</u>			1		X								
		<i>Catharus ustulatus</i>	<u>Swainson's thrush</u>					X	X							
		<i>Catharus guttatus</i>	<u>hermit thrush</u>	3	3	2		X	X							
		<i>Ixoreus naevius</i>	<u>varied thrush</u>					X								
		<i>Turdus migratorius</i>	<u>American robin</u>			1		X	X							
	Mimidae															
		<i>Dumetella carolinensis</i>	<u>gray catbird</u>					X					WSC			
		<i>Mimus polyglottos</i>	northern mockingbird	3		1		X	X	X						
		<i>Toxostoma crissale</i>	crissal thrasher	3	1	3		X	X	X						BCC
		<i>Toxostoma curvirostre</i>	curve-billed thrasher			2		X	X							
		<i>Toxostoma bendirei</i>	Bendire's thrasher					X								
	Bombycillidae															
		<i>Bombycilla cedrorum</i>	<u>cedar waxwing</u>			15		X	X							
	Ptilonotidae															
		<i>Phainopepla nitens</i>	phainopepla	3	2	2		X	X	X						

Order	Family	Scientific name	Common name	UA Detection type				Other efforts			Conservation Designations					
				VCP	Winter	Incidental	Nocturnal	Visitor center	R&D	NPS Monitoring	ESA <sup>a</sup>	US FS <sup>b</sup>	AZ G&F <sup>c</sup>	APF <sup>d</sup>	US FWS <sup>e</sup>	
Passeriformes																
Parulidae																
		<i>Vermivora celata</i>	orange-crowned warbler	1		1		X	X	X						
		<i>Vermivora ruficapilla</i>	Nashville warbler					X	X							
		<i>Vermivora virginiae</i>	Virginia's warbler	1		1		X	X	X						
		<i>Vermivora luciae</i>	Lucy's warbler	19		1		X	X	X					P	
		<i>Dendroica petechia</i>	yellow warbler			1		X								
		<i>Dendroica coronata</i>	yellow-rumped warbler	11	2	2		X	X							
		<i>Dendroica nigrescens</i>	black-throated gray warbler	44		1		X	X	X						
		<i>Dendroica virens</i>	black-throated green warbler					X								
		<i>Dendroica townsendi</i>	Townsend's warbler	2		2		X	X							
		<i>Dendroica occidentalis</i>	hermit warbler					X	X							
		<i>Seiurus aurocapilla</i>	ovenbird					X								
		<i>Oporornis tolmiei</i>	MacGillivray's warbler					X								
		<i>Wilsonia pusilla</i>	Wilson's warbler	4		3		X	X	X						
		<i>Cardellina rubrifrons</i>	red-faced warbler					X								
		<i>Myioborus pictus</i>	painted redstart	1				X	X							
		<i>Icteria virens</i>	yellow-breasted chat					X								
Thraupidae																
		<i>Piranga flava</i>	hepatic tanager	12		1		X	X	X						
		<i>Piranga rubra</i>	summer tanager					X	X							
		<i>Piranga ludoviciana</i>	western tanager	18		2		X	X	X						
Emberizidae																
		<i>Pipilo chlorurus</i>	green-tailed towhee	3		1		X	X	X						
		<i>Pipilo maculatus</i>	spotted towhee	41	15	1		X		X						
		<i>Pipilo fuscus</i>	canyon towhee	26		2		X	X	X						
		<i>Pipilo aberti</i>	Abert's towhee					X								
		<i>Aimophila carpalis</i>	rufous-winged sparrow					X						P	BCC	
		<i>Aimophila cassinii</i>	Cassin's sparrow	18		3		X	X							
		<i>Aimophila texana</i> or <i>botterri</i>	Botteri's sparrow	35						X						
		<i>Aimophila ruficeps</i>	rufous-crowned sparrow	55	5	4		X	X	X						
		<i>Spizella passerina</i>	chipping sparrow	11	13	40		X	X	X						
		<i>Spizella breweri</i>	Brewer's sparrow					X								
		<i>Spizella atrogularis</i>	black-chinned sparrow					X								
		<i>Pooecetes gramineus</i>	vesper sparrow		1	10		X	X							
		<i>Chondestes grammacus</i>	lark sparrow	4		1		X	X	X						
		<i>Passerculus sandwichensis</i>	savannah sparrow					X								
		<i>Amphispiza bilineata</i>	black-throated sparrow	5		2		X	X	X						
		<i>Calcarius ornatus</i>	chestnut-collared longspur						X							
		<i>Calamospiza melanocorys</i>	lark bunting					X	X							
		<i>Passerella iliaca</i>	fox sparrow					X								
		<i>Melospiza melodia</i>	song sparrow					X								
		<i>Ammodramus savannarum</i>	grasshopper sparrow						X							
		<i>Melospiza georgiana</i>	swamp sparrow					X								
		<i>Melospiza lincolni</i>	Lincoln's sparrow						X							

Order				UA Detection type		Other efforts			Conservation Designations					
	Family				Inci-	Noc-	Visitor		NPS		US	AZ		US
	Scientific name	Common name	VCP	Winter	dental	turnal	center	R&D	Moni-	ESA <sup>a</sup>	FS <sup>b</sup>	G&F <sup>c</sup>	APF <sup>d</sup>	FWS <sup>e</sup>
Passeriformes														
Emberizidae														
	<i>Zonotrichia albicollis</i>	white-throated sparrow					X							
	<i>Zonotrichia leucophrys</i>	white-crowned sparrow		1	1		X							
	<i>Junco hyemalis</i>	dark-eyed junco	1	10	2		X	X						
	<i>Junco hyemalis mearnsi</i>	pink-sided junco <sup>f</sup>					X							
	<i>Junco hyemalis dorsalis</i>	gray-headed junco <sup>f</sup>			3		X							
	<i>Junco hyemalis oreganus</i>	Oregon junco <sup>f</sup>		1	1		X							
	<i>Junco phaeonotus</i>	yellow-eyed junco					X							
Cardinalidae														
	<i>Cardinalis cardinalis</i>	northern cardinal			1									
	<i>Cardinalis sinuatus</i>	pyrrhuloxia			3		X	X						
	<i>Pheucticus ludovicianus</i>	rose-breasted grosbeak					X							
	<i>Pheucticus melanocephalus</i>	black-headed grosbeak	25				X	X	X					
	<i>Passerina caerulea</i>	blue grosbeak	23		1		X	X	X					
	<i>Passerina amoena</i>	lazuli bunting			1		X							
	<i>Passerina cyanea</i>	indigo bunting							X					
Icteridae														
	<i>Sturnella magna lilianae</i>	eastern meadowlark	18		10		X	X						
	<i>Sturnella neglecta</i>	western meadowlark	1		1		X	X	X					
	<i>Molothrus aeneus</i>	bronzed cowbird					X							
	<i>Molothrus ater</i>	brown-headed cowbird	27		1		X	X	X					
	<i>Icterus cucullatus</i>	hooded oriole	2		2		X							
	<i>Icterus bullockii</i>	Bullock's oriole					X	X	X					
	<i>Icterus parisorum</i>	Scott's oriole	57		1		X	X	X					
	<i>Icterus pustulatus</i>	streak-backed oriole					X							
Fringillidae														
	<i>Carpodacus cassinii</i>	Cassin's finch					X							
	<i>Carpodacus mexicanus</i>	house finch	31		1		X	X	X					
	<i>Carpodacus purpureus</i>	purple finch					X							
	<i>Carduelis pinus</i>	pine siskin					X	X	X					
	<i>Carduelis tristis</i>	American goldfinch					X							
	<i>Carduelis psaltria</i>	lesser goldfinch	2	3	1		X	X	X					
Passeridae														
	<i>Passer domesticus</i>	house sparrow					X							

<sup>a</sup> ESA = Endangered Species Act: T = Threatened, SC = Species of Concern, C = Candidate for listing (HDMS 2004).

<sup>b</sup> USFS = U.S. Forest Service: Sensitive species (HDMS 2004).

<sup>c</sup> AZG&F = Arizona Game and Fish Department: Wildlife Species of Concern (HDMS 2004).

<sup>d</sup> APF = Arizona Partners in Flight; Priority species (Latta et al. 1999).

<sup>e</sup> USFWS = U.S. Fish and Wildlife Service: Species of conservation concern (HDMS 2004).

<sup>f</sup> We include observations of these subspecies in the appendix because field crew members occasionally made this distinction.

**Appendix D. Mammal species observed or collected at Coronado National Memorial based on 1977-1978 study (Petryszyn and Cockrum 1979), this study, and wildlife sightings database compiled by memorial staff.**

Order	Family	Scientific name	Common name	Confirmed (specimen) 1977–1978	Observed only 1977–1978	Confirmed (specimen) this study	Photo this study	Observed only this study	Observed CORO database
<b>Didelphimorphia</b>									
	Didelphidae	<i>Didelphis virginiana</i>	Virginia opossum				X <sup>a</sup>		X
<b>Insectivora</b>									
	Soricidae	<i>Notiosorex crawfordi</i>	desert shrew			X	X		
<b>Chiroptera</b>									
	Vespertilionidae	<i>Myotis auricolus</i>	southwestern myotis	X					
		<i>Myotis thysanodes</i> <sup>c,d</sup>	fringed myotis	X					
		<i>Myotis californicus</i>	California myotis	X					
		<i>Myotis leibii</i>	eastern small-footed myotis	X					
		<i>Lasiurus cinereus</i>	hoary bat	X					
		<i>Corynorhinus townsendii</i> <sup>c</sup>	Townsend's big-eared bat		X				
<b>Carnivora</b>									
	Ursidae	<i>Ursus americanus</i>	black bear				X		X
	Procyonidae	<i>Procyon lotor</i>	common raccoon				X		X
		<i>Nasua narica</i>	white-nosed coati			X	X		X
		<i>Bassariscus astutus</i>	ringtail		X		X		X
	Mustelidae	<i>Taxidea taxus</i>	American badger						X
	Mephitidae	<i>Spilogale gracilis</i>	western spotted skunk			X	X		X
		<i>Mephitis mephitis</i>	striped skunk				X		X
		<i>Mephitis macroura</i>	hooded skunk			X	X		X
		<i>Conepatus mesoleucus</i>	common hog-nosed skunk				X		X
	Canidae	<i>Canis familiaris</i>	feral dog				X		X
		<i>Canis latrans</i>	coyote		X	X	X		X
		<i>Urocyon cinereoargenteus</i>	common gray fox		X		X		X
	Felidae	<i>Felis catus</i>	feral cat					X	
		<i>Puma concolor</i>	mountain lion				X		X
		<i>Lynx rufus</i>	bobcat			X	X		X
<b>Rodentia</b>									
	Sciuridae	<i>Spermophilus variegatus</i>	rock squirrel		X	X	X		X
		<i>Spermophilus spilosoma</i>	spotted ground squirrel			X <sup>b</sup>			
		<i>Sciurus arizonensis</i> <sup>c</sup>	Arizona gray squirrel		X		X <sup>a</sup>	X	X
	Geomyidae	<i>Thomomys bottae</i>	Botta's pocket gopher	X		X			
		<i>Thomomys umbrinus</i>	southern pocket gopher	X					

Order	Family	Scientific name	Common name	Confirmed (specimen) 1977–1978	Observed only 1977–1978	Confirmed (specimen) this study	Photo this study	Observed only this study	Observed CORO database
<b>Rodentia</b>									
	Heteromyidae								
		<i>Perognathus flavus</i>	silky pocket mouse			X <sup>b</sup>			
		<i>Chaetodipus penicillatus</i>	Sonoran Desert pocket mouse	X		X	X		
		<i>Chaetodipus intermedius</i> <sup>c</sup>	rock pocket mouse	X				X	
		<i>Chaetodipus hispidus</i>	hispid pocket mouse	X		X	X		
		<i>Dipodomys spectabilis</i>	banner-tailed kangaroo rat			X	X		
		<i>Dipodomys ordii</i>	Ord's kangaroo rat			X	X		
		<i>Dipodomys merriami</i>	Merriam's kangaroo rat			X	X		
	Muridae								
		<i>Reithrodontomys megalotis</i> <sup>c</sup>	western harvest mouse	X		X			
		<i>Reithrodontomys fulvescens</i>	fulvous harvest mouse	X		X	X		
		<i>Peromyscus maniculatus</i>	deer mouse	X		X			
		<i>Peromyscus leucopus</i>	white-footed mouse	X		X			
		<i>Peromyscus boylii</i>	brush mouse	X		X	X		
		<i>Baiomys taylori</i>	northern pygmy mouse			X	X		
		<i>Onychomys torridus</i>	southern grasshopper mouse	X			X		
		<i>Neotoma albigula</i>	white-throated woodrat	X		X	X		
		<i>Sigmodon fulviventer</i>	tawny-bellied cotton rat			X			
		<i>Sigmodon ochrognathus</i> <sup>c</sup>	yellow-nosed cotton rat			X	X		
		<i>Sigmodon arizonae</i>	Arizona cotton rat			X	X		
		<b><i>Mus musculus</i></b>	<b>house mouse</b>			<b>X<sup>b</sup></b>			
<b>Lagomorpha</b>									
	Leporidae								
		<i>Lepus californicus</i>	black-tailed jackrabbit		X			X	X
		<i>Sylvilagus audubonii</i>	desert cottontail	X		X			X
<b>Artiodactyla</b>									
	Tayassuidae								
		<i>Pecari tajacu</i>	collared peccary		X	X	X		X
	Cervidae								
		<i>Odocoileus hemionus</i>	mule deer						X
		<i>Odocoileus virginianus</i>	white-tailed deer		X	X	X		X

<sup>a</sup> Recent photographs exist in Coronado NM photo archives.

<sup>b</sup> Species confirmed during trapping in long-term monitoring program, 1997-2003. Species in bold-faced type are non-native.

<sup>c</sup> ESA = Endangered Species Act; SC = Species of Concern (HDMS 2003).

<sup>d</sup> BLM = Bureau of Land Management; Sensitive species (HDMS 2003).



**Appendix E. Species of reptiles and amphibians not reported for CORO NM that could occur based on geographic and elevational range.** All species listed occur in the Huachuca Mountains or San Pedro River Valley between the U.S./Mexico border and northern Sierra Vista. Elevation (Elev.) is given in meters. Species in bold-faced type are non-native.

Taxon	Family	Scientific name Common name	Elev. (m)	Notes	ESA <sup>d</sup>	BLM <sup>e</sup>	USFS <sup>f</sup>
Amphibian	Pelobatidae	<i>Scaphiopus couchii</i> Couch's spadefoot	900-1800 <sup>b</sup>	Confirmed off-site by Johnson and Lowe (1979).			
	Bufonidae	<i>Bufo alvarius</i> Sonoran Desert toad	1250-1387 <sup>b</sup>	Southern Huachucas included in range map of Stebbins (1985).			
	Hylidae	<i>Hyla eximia</i> mountain treefrog	900-2900 <sup>b</sup>	Occurs in Huachucas <16 km (10 mi) to north and west of CORO (AGFD 1996).			
	Ranidae	<i>Rana chiricahuensis</i> Chiricahua leopard frog	1070-2410 <sup>c</sup>	Historic records for San Pedro River just east of CORO (AGFD 1996). Little suitable habitat on site.	LT		X
		<i>Rana subaquavocalis</i> Ramsey Canyon leopard frog <sup>d</sup>	5500-5800	Occurs about 16 km (10 mi) north of CORO in Ramsey, Tinker, and Brown canyons (AGFD 1996); little suitable habitat on-site.			
		<b><i>Rana catesbeiana</i></b> <b>bullfrog</b>	<b>861-2100<sup>b</sup></b>	<b>Confirmed 3.5 km (2.2 mi) west of Montezuma Pass by Johnson and Lowe (1979). Little suitable habitat at CORO.</b>			
Reptile	Kinosternidae	<i>Kinosternon sonoriense</i> Sonoran mud turtle	1200-2000 <sup>b</sup>	Specimen in UAZ collected 8 km (5 mi) east of CORO. Little suitable habitat occurs on-site, but may eventually be found, particularly in Yaqui drainage.			
	Testudinidae	<i>Gopherus agassizii</i> desert tortoise	224-1606 <sup>a</sup>	A single record exists for the east side of the Huachucas in Miller Canyon (<16 km and 10 mi north of CORO; AGFD 1996); very unlikely at CORO.	SC		
	Eublepharidae	<i>Coleonyx variegatus</i> western banded gecko	0-1485 <sup>b</sup>	CORO included in range map of Stebbins (1985). We believe this species occurs at CORO.			
	Crotaphytidae	<i>Gambelia wislizenii</i> longnose leopard lizard	2970-6930 <sup>b</sup>	CORO included in range map of Stebbins (1985). Usually found at lower elevations.			
	Phrynosomatidae	<i>Cophosaurus texanus</i> greater earless lizard	250-1545 <sup>b</sup>	CORO included in range map of Stebbins (1985). We are not sure why this species does not occur at CORO, but none were observed during our study.			
		<i>Callisaurus draconoides</i> zebratail lizard	1180-1250 <sup>b</sup>	CORO included in range map of Stebbins (1985), but generally found at lower elevations.			
		<i>Sceloporus magister</i> desert spiny lizard	900-1980 <sup>b</sup>	CORO included on Stebbins (1985) range map, but usually found at lower elevations than CORO.			
		<i>Uta stansburiana</i> side-blotched lizard	0-2750 <sup>c</sup>	CORO included on Stebbins (1985) range map, but usually found at lower elevations than CORO.			
		<i>Phrynosoma cornutum</i> Texas horned lizard	0-1387 <sup>b</sup>	Confirmed near CORO by Johnson and Lowe (1979). Fairly common in San Pedro grasslands, and may occur in low numbers.	SC		X

Taxon	Family	Scientific name Common name	Elev. (m)	Notes	ESA <sup>d</sup>	BLM <sup>e</sup>	USFS <sup>f</sup>
Reptile	Phrynosomatidae	<i>Phrynosoma modestum</i> round-tailed horned lizard	900-2200 <sup>b</sup>	CORO is just on the western edge of the range of this species (Stebbins 1985).			
		<i>Phrynosoma solare</i> regal horned lizard	900-2100 <sup>b</sup>	CORO included in Stebbins (1985). Usually found at lower elevations; not expected at CORO.			
	Teiidae	<i>Cnemidophorus burti</i> canyon spotted whiptail	1321-1387 <sup>b</sup>	CORO included in range map of Stebbins (1985), but distribution in southern Arizona is spotty.	SC	X	X
		<i>Cnemidophorus tigris</i> western whiptail (tiger whiptail)	900-1675 <sup>b</sup>	Occurs in San Pedro Valley at lower elevations; unlikely to occur on-site.			
	Leptotyphlopidae	<i>Leptotyphlops humilis</i> western blind snake	900-1425 <sup>b</sup>	Records exist from local area (AGFD 1996). Johnson and Lowe (1979) listed this species as of possible occurrence. We believe it may eventually be found on-site at lower elevations.			
	Colubridae	<i>Senticolis triaspis</i> green rat snake	0-2200 <sup>b</sup>	This species has not been documented in the Huachuclas. Johnson and Lowe (1979) believed that green rat snakes occurred at CORO based on sight records, but we believe these were misidentifications. The nearest records are from the Nogales area (AGFD 1996).			
		<i>Arizona occidentalis</i> western glossy snake	900-2200 <sup>b</sup>	Johnson and Lowe (1979) listed this species as of possible occurrence; CORO is within the range, but it is more often found at lower elevations.			
		<i>Lampropeltis triangulum</i> milk snake	1417-1417 <sup>a</sup>	Milk snakes are a very elusive and rare grassland species. The habitat is suitable but they have not been found in the San Pedro Valley.			
		<i>Thamnophis eques</i> Mexican garter snake	1125-1650 <sup>b</sup>	Found in area, but generally prefers larger water sources than occur at CORO.	SC		X
		<i>Thamnophis marcianus</i> checkered garter snake	1133-1864 <sup>a</sup>	Confirmed near CORO by Johnson and Lowe (1979). Common in San Pedro grasslands and may occur on-site as occasional or in low numbers.			
		<i>Sonora semiannulata</i> ground snake	900-1600 <sup>b</sup>	Within the range of Stebbins (1985), but usually found at lower elevations.			
		<i>Tantilla hobartsmithi</i> southwestern blackhead snake	900-1600 <sup>b</sup>	CORO is within range of this species (Stebbins 1985) but it is more common at lower elevations.			
		<i>Tantilla yaquia</i> Yaqui blackhead snake	1170-1576 <sup>a</sup>	CORO is within range of this species (Stebbins 1985), but the nearest specimens are from approximately 40 km (25 mi) north.			

Taxon	Family	Scientific name Common name	Elev. (m)	Notes	ESA <sup>d</sup>	BLM <sup>e</sup>	USFS <sup>f</sup>
	Viperidae	<i>Sistrurus catenatus</i> massasauga	1239-1535 <sup>a</sup>	This species occurred historically in the San Pedro grasslands, but is now very rare in Arizona. The AGFD Heritage Database (AGFD 1996) contains a 1975 record from Route 92 less than 9 km (5 mi) north of CORO.			
		<i>Crotalus pricei</i> twin-spotted rattlesnake	2012-2788 <sup>a</sup>	Records for this species are from Miller Canyon and Carr Peak. Some suitable habitat exists at CORO and the species may occur in low numbers, but we found none despite extensive searches.			
		<i>Crotalus willardi</i> ridgenosed rattlesnake	1600-2750 <sup>b</sup>	Records for this species are from Ramsey Canyon, Carr Peak. Generally prefers wet canyons and John Porter, a local expert, does not believe they occur on-site (J. Porter, <i>personal communication</i> ).			X

<sup>a</sup> Arizona Game and Fish Department (1996) for Cochise County.

<sup>b</sup> Degenhardt et al. (1996) for New Mexico.

<sup>c</sup> Stebbins (1985).

<sup>d</sup> ESA = Endangered Species Act: LT = Listed Threatened, SC = Species of Concern (HDMS 2003).

<sup>e</sup> BLM = Bureau of Land Management: Sensitive species (HDMS 2003).

<sup>f</sup> USFS = U.S. Forest Service: Sensitive species (HDMS 2003).

**Appendix F. Mammal species that might occur at Coronado NM including those believed to be possible by Petryszyn and Cockrum (1979), E = presently extirpated from Arizona; R = very rare in Arizona, occasional sightings only; U = status unknown, never confirmed in Arizona.**

Order	Family	Scientific name	Common name	Status	ESA <sup>a</sup>	BLM <sup>b</sup>	USFS <sup>c</sup>	AG&F <sup>d</sup>
Insectivora	Soricidae	<i>Sorex arizonae</i>	Arizona shrew		SC		X	X
Chiroptera	Mormoopidae	<i>Mormoops megalophylla</i>	ghost-faced bat					
	Phyllostomidae	<i>Choeronycteris mexicana</i>	Mexican long-tongued bat		SC	X		X
		<i>Leptonycteris curasoae</i>	southern long-nosed bat		LE		X	X
	Vespertilionidae	<i>Myotis velifer</i>	cave myotis		SC	X		
		<i>Myotis volans</i>	long-legged myotis		SC	X		
		<i>Lasionycteris noctivagans</i>	silver-haired bat					
		<i>Pipistrellus hesperus</i>	western pipistrelle					
		<i>Eptesicus fuscus</i>	big brown bat					
		<i>Euderma maculatum</i>	spotted bat		SC		X	X
		<i>Lasiurus borealis</i>	eastern red bat					
		<i>Lasiurus ega</i>	southern yellow bat					X
		<i>Idionycteris phyllotis</i>	Allen's big-eared bat		SC	X		
		<i>Antrozous pallidus</i>	pallid bat					
	Molossidae	<i>Tadarida brasiliensis</i>	Brazilian free-tailed bat					
		<i>Nyctinomops femorosaccus</i>	pocketed free-tailed bat			X		
		<i>Nyctinomops macrotis</i>	big free-tailed bat		SC	X		
		<i>Eumops perotis</i>	western bonneted bat		SC			
Carnivora	Ursidae	<i>Ursus arctos</i>	grizzly bear	E				
	Canidae	<i>Canis lupus baileyi</i>	Mexican gray wolf	E	LE		X	X
		<i>Vulpes macrotis</i>	kit fox					
	Felidae	<i>Panthera onca</i>	jaguar	R	LE		X	X
		<i>Leopardus pardalis</i>	ocelot	R	LE			X
		<i>Herpailurus yaguarondi</i>	jaguarundi	U	LE	X		
Rodentia	Sciuridae	<i>Spermophilus tereticaudus</i>	round-tailed ground squirrel					
		<i>Cynomys ludovicianus</i>	black-tailed prairie dog	E	SC			X
	Heteromyidae	<i>Chaetodipus baileyi</i>	Bailey's pocket mouse					
	Muridae	<i>Reithrodontomys montanus</i>	Plains harvest mouse					
		<i>Peromyscus eremicus</i>	cactus mouse		SC		X	
		<i>Onychomys leucogaster</i>	northern grasshopper mouse					
		<i>Neotoma mexicana</i>	Mexican woodrat		SC			
	Erethizontidae	<i>Erethizon dorsatum</i>	common porcupine					
Lagomorpha	Leporidae	<i>Lepus alleni</i>	antelope jackrabbit					
		<i>Sylvilagus floridanus</i>	eastern cottontail					

<sup>a</sup> ESA = Endangered Species Act: LE = Listed Endangered, SC = Species of Concern (HDMS 2003).

<sup>b</sup> BLM = Bureau of Land Management: Sensitive species (HDMS 2003).

<sup>c</sup> USFS = U.S. Forest Service: Sensitive species (HDMS 2003).

<sup>d</sup> AZG&F = Arizona Game and Fish Department: Wildlife Species of Concern (HDMS 2003).

**Appendix G. Incidental reptile and mammal observations collected during bird inventory ,  
Coronado NM 2003–2004.**

Taxa	Order	Family	Scientific name	Common name
Reptile	Squamata	Helodermatidae	<i>Heloderma suspectum</i>	Gila monster
		Colubridae	<i>Masticophis bilineatus</i>	Sonoran whipsnake
			<i>Pituophis catenifer</i>	gopher snake
		Viperidae	<i>Crotalus molossus</i>	black-tailed rattlesnake
Mammal	Didelphimorphia	Didelphidae	<i>Didelphis virginiana</i>	Virginia opossum
	Carnivora	Procyonidae	<i>Nasua narica</i>	white-nosed coati
		Mephitidae	<i>Spilogale gracilis</i>	western spotted skunk
		Canidae	<i>Canis latrans</i>	coyote
			<i>Urocyon cinereoargenteus</i>	common gray fox
	Rodentia	Sciuridae	<i>Spermophilus variegatus</i>	rock squirrel
	Lagomorpha	Leporidae	<i>Sylvilagus audubonii</i>	desert cottontail
	Artiodactyla	Tayassuidae	<i>Pecari tajacu</i>	collared peccary
		Cervidae	<i>Odocoileus virginianus</i>	white-tailed deer

**Appendix H. List of voucher specimens collected from Coronado NM and located in various collections.** See Table 1.1 for list of collections that were queried for specimens with “Coronado National Memorial” in the location field. See Appendices B, C, and D for scientific names.

Taxon	Scientific name	Common name	Museum <sup>a</sup>	Collection #	Collection date	Collector
Amphibian	<i>Bufo woodhousii</i>	Woodhouse's toad	UK	6084, 6085		
			NHMLA	88446, 88447		
Reptile	<i>Crotaphytus collaris</i>	eastern collared lizard	CAS	16904-11187	10/21/1935	F. N. Blanchard and H. K. Gloyd
	<i>Holbrookia maculata</i>	lesser earless lizard	CAS	17128-11407	10/21/1935	F. N. Blanchard and H. K. Gloyd
			UI	17540	08/23/1950	D. F. Hoffmeister
			UM	69785	07/28/1930	H. K. Gloyd
	<i>Sceloporus clarkii</i>	Clark's spiny lizard	CAS	17167-11446	10/21/1935	F. N. Blanchard and H. K. Gloyd
			UM	53981	09/01/1919	C. R. Biederman
	<i>Sceloporus virgatus</i>	striped plateau lizard	UM	69911	07/28/1930	H. K. Gloyd
	<i>Urosaurus ornatus</i>	ornate tree lizard	BYU	32441	06/03/1969	J. R. McMorris
			UM	69839	07/28/1930	H. K. Gloyd
	<i>Eumeces obsoletus</i>	Great Plains skink	MCZ	29029, 29027		
			UM	53987	09/01/1919	C. R. Biederman
	<i>Elgaria kingii</i>	Madrean alligator lizard	UM	53988	09/01/1919	C. R. Biederman
	<i>Masticophis flagellum</i>	coachwhip	MCZ	29028		
	<i>Salvadora hexalepis</i>	western patch-nosed snake	CAS	8096-12198	10/21/1935	F. N. Blanchard and H. K. Gloyd
	<i>Crotalus molossus</i>	black-tailed rattlesnake	UC	40921	07/27/1969	R. L. Holland
Bird	<i>Selasphorus rufus</i>	rufous hummingbird	UA	12063		
	<i>Pyrocephalus rubinus</i>	vermillion flycatcher	UA	17493		
	<i>Thryomanes bewickii</i>	Bewick's wren	UA	17494		
	<i>Pipilo maculatus</i>	spotted towhee	UA	17489		
			UA	17490		
	<i>Aimophila cassinii</i>	Cassin's sparrow	UA	17486		
			UA	17487		
	<i>Aimophila ruficeps</i>	rufous-crowned sparrow	UA	17483		
			UA	17484		
			UA	17485		
	<i>Amphispiza bilineata</i>	black-throated sparrow	UA	17488		
	<i>Cardinalis sinuatus</i>	pyrrhuloxia	UA	17492		
	<i>Sturnella magna</i>	eastern meadowlark	UA	17491		
Mammal	<i>Myotis auriculus</i>	southwestern myotis	UA	23766, 23767	06/16/1978	Y. Petryszyn
	<i>Myotis thysanodes</i>	fringed myotis	UA	23770	06/16/1978	Y. Petryszyn
	<i>Myotis californicus</i>	California myotis	UA	23768	06/16/1978	Y. Petryszyn
	<i>Myotis leibii</i>	eastern small-footed myotis	UA	23769	06/21/1978	Y. Petryszyn
	<i>Lasiurus cinereus</i>	hoary bat	UA	23756	06/18/1978	Y. Petryszyn
	<i>Nasua narica</i>	white-nosed coati	UA	26614	06/06/1998	T. A. Edwards
	<i>Spilogale gracilis</i>	western spotted skunk	UA	26615	06/06/1998	T. A. Edwards
	<i>Conepatus mesoleucus</i>	white-backed hog-nosed skunk	UA	26616	05/02/1997	D. Swann
	<i>Thomomys bottae</i>	Botta's pocket gopher	UA	23772, 26635	06/18/1978, 04/12/1997	Y. Petryszyn, D. E. Swann
	<i>Thomomys umbrinus</i>	southern pocket gopher	UA	23771	06/21/1978	Y. Petryszyn

Taxon	Scientific name	Common name	Museum <sup>a</sup>	Collection #	Collection date	Collector
Mammal	<i>Chaetodipus penicillatus</i>	Sonoran Desert pocket mouse	UA	23765	06/16/1978	Y. Petryszyn
	<i>Chaetodipus hispidus</i>	hispid pocket mouse	UA	23755	06/22/1978	Y. Petryszyn
	<i>Dipodomys spectabilis</i>	banner-tailed kangaroo rat	UA	26638	11/10/1996	D. E. Swann
	<i>Dipodomys ordii</i>	Ord's kangaroo rat	UA	26636	04/11/1997	D. E. Swann
	<i>Reithrodontomys megalotis</i>	western harvest mouse	UA	23760, 26637	06/18/1978, 11/25/1997	Y. Petryszyn, D. E. Swann
	<i>Reithrodontomys fulvescens</i>	fulvous harvest mouse	UA	26611	12/16/1996	T. A. Edwards
	<i>Peromyscus maniculatus</i>	deer mouse	UA	23764, 26609, 26639	06/18/1978, 04/17/1997, 06/18/1977	M. Courtney, D. Swann
	<i>Peromyscus boylii</i>	brush mouse	UA	23758, 23759, 23761, 23762, 23763, 26610	06/16/1978, 06/18/1978, 12/16/1996	Y. Petryszyn, M. Courtney, T. A. Edwards
	<i>Onychomys torridus</i>	southern grasshopper mouse	UA	23757	6/22/1978	Y. Petryszyn
	<i>Neotoma albigula</i>	western white-throated woodrat	UA	23773, 23775, 26613	06/16/1978, 06/21/1978, 10/07/1996	Y. Petryszyn, T. A. Edwards
	<i>Sigmodon ochrognathus</i>	yellow-nosed cotton rat	UA	26612	10/06/1996	D. Swann

<sup>a</sup> BYU = Brigham Young University; CAS = Chicago Academy of Sciences; MCZ = Museum of Comparative Zoology, Harvard University; NHMLA = Natural History Museum of Los Angeles County; UA = University of Arizona Collections; UC = University of Colorado Museum; UI = University of Illinois, Museum of Natural History; UK = University of Kansas, Museum of Natural History; UM = University of Michigan.

**Appendix I. Total number of observations, by transect and VCP survey type, Coronado NM, 2002–2004.** Observations include flyovers and birds seen at unlimited distances from stations. Numbers should not be used as a measure of relative abundance (see Table 5.3–5.5 for results expressed as relative abundance). See Appendix C for scientific names.

Species	Transect type							
	Repeat visit		Reconnaissance					
	Riparian	Wash	Coronado Peak	Crest Trail	Level	Ranch	Ridge	Uplands
Gambel's quail	1							
Montezuma quail		1						
turkey vulture	49	35		5	4	8		1
Cooper's hawk	9							
zone-tailed hawk		1						
red-tailed hawk	3	1						
golden eagle	1							
American kestrel				1				
prairie falcon	2	2				1		
white-winged dove	16	20	1			1		1
mourning dove	6	31	1	3		2	2	1
yellow-billed cuckoo	1							
greater roadrunner	6	7	2	1	1		1	
white-throated swift	76	16		3	6			3
broad-billed hummingbird	1							
black-chinned hummingbird	4	3						2
Anna's hummingbird	4	1						
broad-tailed hummingbird	6							
rufous hummingbird	2							
acorn woodpecker	1							
ladder-backed woodpecker	4	8		1			1	1
Arizona woodpecker	10	3						
northern flicker	13							
western wood-pewee	2	1		1			3	
Hammond's flycatcher	5	4						
gray flycatcher	1	2						
western flycatcher	2							
cordilleran flycatcher	1							
black phoebe	1							
Say's phoebe					1		2	1
dusky-capped flycatcher	58	1	1					
ash-throated flycatcher	28	54	1	2	1	2	1	1
sulphur-bellied flycatcher	14							
Cassin's kingbird	10	11	1				2	2
western kingbird		3				5		1
loggerhead shrike		1						
Bell's vireo		1						
plumbeous vireo	6	1						
Hutton's vireo	24			1				
curve-billed thrasher				2			4	
crissal thrasher	1	2		1	2			
western scrub-jay	1		1	9			4	
Mexican jay	148	53			8	10	4	4
unknown raven	6	19						



Species	Transect type							
	Repeat visit		Reconnaissance					
	Riparian	Wash	Coronado Peak	Crest Trail	Level	Ranch	Ridge	Uplands
violet-green swallow		3	3	6			15	
barn swallow		1						
bridled titmouse	42	14			5			3
verdin		18			1	3		
bushtit	29	21		5	39	7		
white-breasted nuthatch	3				1			
cactus wren		11	3	3		2	2	
rock wren			1				1	
canyon wren	11						2	
Bewick's wren	120	57	1	6	10	2	3	8
house wren		1						
ruby-crowned kinglet	10	9						
blue-gray gnatcatcher	1			1	2			
hermit thrush	3							
northern mockingbird		3						2
phainopepla		3						
orange-crowned warbler		1						
Virginia's warbler		1						
Lucy's warbler	4	15				1		
yellow-rumped warbler	11							
black-throated gray warbler	43	1						
Townsend's warbler	1	1						
Wilson's warbler	4							
painted redstart	1							
hepatic tanager	11	1	1				2	
western tanager	5	13		2				
green-tailed towhee		3						
spotted towhee	39	2	2	7			3	
canyon towhee	7	19		2		2	7	5
Cassin's sparrow		18						
Botteri's sparrow		35						
rufous-crowned sparrow	33	22	1	4		2	11	5
chipping sparrow	1	10						
lark sparrow		4			1			1
black-throated sparrow	4	1			1			1
dark-eyed junco	1							
black-headed grosbeak	24	1						1
blue grosbeak	1	22				2		
eastern meadowlark		18				5		
western meadowlark		1						
brown-headed cowbird	17	10			2	1	1	
hooded oriole	2					1		
Scott's oriole	43	14	2	1	4		8	4
house finch	15	16	1	2	1	9	1	
lesser goldfinch	2							
<b>Number of observations</b>	<b>1011</b>	<b>652</b>	<b>23</b>	<b>70</b>	<b>91</b>	<b>67</b>	<b>79</b>	<b>47</b>
<b>Species richness</b>	<b>64</b>	<b>60</b>	<b>16</b>	<b>24</b>	<b>19</b>	<b>19</b>	<b>22</b>	<b>19</b>

**Appendix J. Summary of vegetation characteristics measured at bird survey stations, Coronado NM, 2004.** See Appendix A for list of common names.

Transect			
Station	Category	Species	Mean density
<b>Riparian</b>			
1	Subshrub	<i>Agave palmeri</i>	32.91
		<i>Rhus choriophylla</i>	19.74
		<i>Toxicodendron radicans</i> ssp. <i>radicans</i>	6.58
		<i>Opuntia engelmannii</i>	6.58
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	19.74
		<i>Mimosa grahamii</i>	6.58
		<i>Dasyllirion wheeleri</i>	32.91
		<i>Nolina microcarpa</i>	6.58
	Shrub	<i>Agave palmeri</i>	70.02
		<i>Yucca schottii</i>	35.01
		<i>Rhus choriophylla</i>	105.03
		<i>Rhus trilobata</i>	17.51
		<i>Arctostaphylos pungens</i>	35.01
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	17.51
		<i>Dasyllirion wheeleri</i>	17.51
		<i>Nolina microcarpa</i>	35.01
		<i>Pinus cembroides</i>	17.51
	Tree	<i>Juniperus deppeana</i>	6.95
		<i>Quercus arizonica</i>	31.27
		<i>Quercus emoryi</i>	24.32
		<i>Pinus cembroides</i>	3.48
		<i>Platanus wrightii</i>	3.48
	Cavity	<i>Quercus arizonica</i>	18.38
		<i>Quercus emoryi</i>	28.88
		<i>Platanus wrightii</i>	5.25
2	Subshrub	<i>Agave palmeri</i>	9.61
		<i>Yucca schottii</i>	9.61
		<i>Rhus choriophylla</i>	38.43
		<i>Toxicodendron radicans</i> ssp. <i>radicans</i>	9.61
		<i>Baccharis pteronioides</i>	9.61
		<i>Baccharis thesioides</i>	9.61
		<i>Dalea</i> sp.	9.61
		<i>Eysenhardtia orthocarpa</i>	19.22
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	28.82
		<i>Mimosa grahamii</i>	9.61
		<i>Quercus emoryi</i>	9.61
		<i>Dasyllirion wheeleri</i>	19.22
		<i>Nolina microcarpa</i>	9.61
	Shrub	<i>Rhus choriophylla</i>	157.09
		<i>Baccharis sarothroides</i>	14.28
		<i>Arctostaphylos pungens</i>	42.84
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	14.28
		<i>Prosopis velutina</i>	14.28
		<i>Dasyllirion wheeleri</i>	14.28
		<i>Nolina microcarpa</i>	14.28
		<i>Pinus cembroides</i>	14.28

Transect			
Station	Category	Species	Mean density
	Tree	<i>Juniperus deppeana</i>	4.75
		<i>Quercus arizonica</i>	42.74
		<i>Quercus emoryi</i>	42.74
		<i>Platanus wrightii</i>	4.75
	Cavity	<i>Quercus arizonica</i>	29.26
		<i>Quercus emoryi</i>	23.94
3	Subshrub	<i>Agave palmeri</i>	63.66
		<i>Baccharis thesioides</i>	63.66
		<i>Opuntia spinosior</i>	63.66
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	763.94
		<i>Mimosa grahamii</i>	63.66
		<i>Dasyllirion wheeleri</i>	190.99
		<i>Nolina microcarpa</i>	63.66
	Shrub	<i>Yucca schottii</i>	58.15
		<i>Rhus choriophylla</i>	87.23
		<i>Rhus trilobata</i>	58.15
		<i>Juniperus deppeana</i>	29.08
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	203.54
	Tree	<i>Quercus emoryi</i>	58.15
		<i>Dasyllirion wheeleri</i>	58.15
		<i>Nolina microcarpa</i>	29.08
	Cavity	<i>Quercus arizonica</i>	27.20
		<i>Quercus emoryi</i>	95.19
		<i>Quercus oblongifolia</i>	6.80
		<i>Pinus cembroides</i>	6.80
		<i>Quercus arizonica</i>	10.15
		<i>Quercus emoryi</i>	22.00
		<i>Quercus oblongifolia</i>	1.69
4	Subshrub	<i>Yucca schottii</i>	22.82
		<i>Rhus trilobata</i>	68.47
		<i>Baccharis thesioides</i>	159.76
		<i>Opuntia spinosior</i>	22.82
		<i>Prosopis velutina</i>	22.82
		<i>Garrya wrightii</i>	22.82
		<i>Dasyllirion wheeleri</i>	91.29
		<i>Nolina microcarpa</i>	22.82
		<i>Pinus cembroides</i>	22.82
	Shrub	<i>Yucca schottii</i>	72.32
		<i>Rhus choriophylla</i>	72.32
		<i>Rhus trilobata</i>	72.32
		<i>Prosopis velutina</i>	18.08
		<i>Quercus arizonica</i>	54.24
		<i>Quercus emoryi</i>	36.16
		<i>Dasyllirion wheeleri</i>	18.08
	Tree	<i>Nolina microcarpa</i>	18.08
		<i>Quercus arizonica</i>	91.38
		<i>Quercus emoryi</i>	74.77

Transect			
Station	Category	Species	Mean density
<b>Riparian</b>			
4	Cavity	<i>Juniperus deppeana</i>	3.00
		<i>Quercus arizonica</i>	33.01
		<i>Quercus emoryi</i>	21.01
		<i>Quercus sp.</i>	3.00
5	Subshrub	<i>Yucca schottii</i>	15.08
		<i>Rhus choriophylla</i>	60.33
		<i>Toxicodendron radicans</i> ssp. <i>radicans</i>	30.16
		<i>Rhus trilobata</i>	15.08
		<i>Baccharis thesioides</i>	15.08
		<i>Opuntia spinosior</i>	15.08
		<i>Sambucus cerulea</i>	15.08
		<i>Juniperus deppeana</i>	15.08
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	75.41
		<i>Quercus arizonica</i>	15.08
		<i>Dasyllirion wheeleri</i>	15.08
		<i>Nolina microcarpa</i>	15.08
	Shrub	<i>Yucca schottii</i>	27.65
		<i>Rhus choriophylla</i>	110.59
		<i>Rhus trilobata</i>	13.82
		<i>Opuntia spinosior</i>	13.82
	Tree	<i>Arctostaphylos pungens</i>	13.82
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	13.82
		<i>Quercus arizonica</i>	13.82
		<i>Quercus emoryi</i>	13.82
		<i>Dasyllirion wheeleri</i>	13.82
		<i>Nolina microcarpa</i>	13.82
		<i>Pinus cembroides</i>	27.65
		<i>Rhus choriophylla</i>	8.68
		<i>Juniperus deppeana</i>	17.35
		<i>Arbutus arizonica</i>	8.68
		<i>Quercus arizonica</i>	78.09
		<i>Quercus emoryi</i>	60.74
6	Subshrub	<i>Juniperus deppeana</i>	2.09
		<i>Arbutus arizonica</i>	2.09
		<i>Quercus arizonica</i>	18.83
		<i>Quercus emoryi</i>	14.64
		<i>Platanus wrightii</i>	2.09
	Cavity	<i>Yucca schottii</i>	46.29
		<i>Rhus choriophylla</i>	30.86
		<i>Rhus trilobata</i>	46.29
		<i>Baccharis thesioides</i>	15.43
		<i>Opuntia spinosior</i>	46.29
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	46.29
		<i>Quercus arizonica</i>	15.43
		<i>Garrya wrightii</i>	15.43
		<i>Dasyllirion wheeleri</i>	30.86
		<i>Pinus cembroides</i>	15.43

Transect			
Station	Category	Species	Mean density
	Shrub	<i>Rhus choriophylla</i>	170.02
		<i>Rhus trilobata</i>	145.73
		<i>Opuntia spinosior</i>	24.29
		<i>Arctostaphylos pungens</i>	24.29
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	24.29
		<i>Quercus arizonica</i>	24.29
		<i>Quercus emoryi</i>	24.29
		<i>Nolina microcarpa</i>	24.29
		<i>Pinus cembroides</i>	24.29
	Tree	<i>Juniperus deppeana</i>	23.37
		<i>Quercus arizonica</i>	151.93
		<i>Quercus emoryi</i>	46.75
		<i>Quercus hypoleucoides</i>	11.69
	Cavity	<i>Juniperus deppeana</i>	5.73
		<i>Quercus arizonica</i>	17.19
		<i>Quercus emoryi</i>	11.46
7	Subshrub	<i>Agave palmeri</i>	14.47
		<i>Yucca schottii</i>	43.41
		<i>Rhus choriophylla</i>	14.47
		<i>Baccharis pteronioides</i>	14.47
		<i>Opuntia sp.</i>	14.47
		<i>Opuntia spinosior</i>	14.47
		<i>Arctostaphylos pungens</i>	14.47
		<i>Quercus arizonica</i>	43.41
		<i>Quercus hypoleucoides</i>	57.88
		<i>Dasyllirion wheeleri</i>	14.47
		<i>Pinus cembroides</i>	43.41
	Shrub	<i>Yucca schottii</i>	117.44
		<i>Rhus choriophylla</i>	88.08
		<i>Arctostaphylos pungens</i>	146.79
		<i>Quercus emoryi</i>	29.36
	Tree	<i>Quercus hypoleucoides</i>	58.72
		<i>Pinus cembroides</i>	146.79
		<i>Juniperus deppeana</i>	9.03
		<i>Arbutus arizonica</i>	3.01
		<i>Quercus arizonica</i>	24.07
		<i>Quercus emoryi</i>	15.04
		<i>Quercus hypoleucoides</i>	9.03
	Cavity	<i>Juniperus deppeana</i>	6.68
		<i>Arbutus arizonica</i>	3.34
		<i>Quercus arizonica</i>	16.70
		<i>Quercus emoryi</i>	16.70
		<i>Quercus hypoleucoides</i>	10.02
		<i>Pinus cembroides</i>	3.34
	8 Subshrub	<i>Agave palmeri</i>	11.87
		<i>Toxicodendron radicans</i> ssp. <i>radicans</i>	11.87
		<i>Rhus trilobata</i>	11.87
		<i>Brickellia sp.</i>	59.35
		<i>Arctostaphylos pungens</i>	11.87

Transect				
Station	Category	Species	Mean density	
Riparian				
8	Subshrub	<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	47.48	
		<i>Quercus arizonica</i>	35.61	
		<i>Quercus emoryi</i>	11.87	
		<i>Garrya wrightii</i>	23.74	
		<i>Dasyllirion wheeleri</i>	11.87	
	Shrub	<i>Rhus choriophylla</i>	7.45	
		<i>Toxicodendron radicans</i> ssp. <i>radicans</i>	7.45	
		<i>Rhus trilobata</i>	37.24	
		<i>Brickellia</i> sp.	22.34	
		<i>Juniperus deppeana</i>	7.45	
		<i>Arctostaphylos pungens</i>	22.34	
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	14.90	
		<i>Quercus arizonica</i>	7.45	
		<i>Quercus emoryi</i>	7.45	
		<i>Juglans major</i>	14.90	
	Tree	<i>Juniperus deppeana</i>	16.18	
		<i>Arbutus arizonica</i>	5.39	
		<i>Quercus arizonica</i>	48.53	
		<i>Juglans major</i>	16.18	
		<i>Platanus wrightii</i>	16.18	
	Cavity	<i>Salix gooddingii</i>	5.39	
		<i>Juniperus deppeana</i>	20.03	
		<i>Arbutus arizonica</i>	6.68	
		<i>Quercus arizonica</i>	16.69	
		<i>Quercus</i> sp.	3.34	
	Wash	<i>Juglans major</i>	3.34	
		<i>Platanus wrightii</i>	13.35	
		<i>Salix gooddingii</i>	3.34	
1		Subshrub	<i>Baccharis sarothroides</i>	29.97
			<i>Ericameria nauseosa</i> var. <i>nauseosa</i>	2.73
			<i>Isocoma tenuisecta</i>	5.45
			<i>Opuntia spinosior</i>	2.73
			<i>Calliandra eriophylla</i>	2.73
			<i>Eysenhardtia orthocarpa</i>	2.73
			<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	5.45
	<i>Sapindus saponaria</i>		2.73	
	Shrub	<i>Agave palmeri</i>	10.14	
		<i>Baccharis sarothroides</i>	152.08	
		<i>Ericameria nauseosa</i> var. <i>nauseosa</i>	20.28	
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	10.14	
		<i>Sapindus saponaria</i>	10.14	
	Tree	<i>Chilopsis linearis</i>	8.13	
		<i>Prosopis velutina</i>	10.83	
		<i>Quercus emoryi</i>	2.71	
		<i>Quercus oblongifolia</i>	4.06	
		<i>Sapindus saponaria</i>	1.35	
	Cavity	<i>Quercus emoryi</i>	4.83	

Transect			
Station	Category	Species	Mean density
2	Subshrub	<i>Agave palmeri</i>	3.20
		<i>Baccharis sarothroides</i>	4.27
		<i>Opuntia spinosior</i>	3.20
		<i>Calliandra eriophylla</i>	7.46
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	2.13
	Shrub	<i>Prosopis velutina</i>	1.07
		<i>Agave palmeri</i>	8.43
		<i>Baccharis sarothroides</i>	42.14
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	2.81
		<i>Prosopis velutina</i>	2.81
	Tree	<i>Baccharis sarothroides</i>	0.35
		<i>Chilopsis linearis</i>	0.35
		<i>Prosopis velutina</i>	2.79
	Cavity	<i>Quercus arizonica</i>	0.16
		<i>Quercus emoryi</i>	1.41
3	Subshrub	<i>Agave palmeri</i>	75.13
		<i>Baccharis sarothroides</i>	16.10
		<i>Calliandra eriophylla</i>	10.73
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	5.37
	Shrub	<i>Agave palmeri</i>	2.52
		<i>Baccharis sarothroides</i>	35.24
		<i>Eysenhardtia orthocarpa</i>	10.07
		<i>Prosopis velutina</i>	2.52
		<i>Chilopsis linearis</i>	2.14
	Tree	<i>Prosopis velutina</i>	1.84
		<i>Quercus emoryi</i>	2.14
		<i>Quercus emoryi</i>	2.72
	Cavity	<i>Quercus emoryi</i>	2.72
		<i>Quercus emoryi</i>	2.72
4	Subshrub	<i>Agave palmeri</i>	39.18
		<i>Baccharis pteronioides</i>	3.92
		<i>Baccharis sarothroides</i>	31.35
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	3.92
	Shrub	<i>Agave palmeri</i>	7.13
		<i>Baccharis sarothroides</i>	60.64
		<i>Dasyllirion wheeleri</i>	3.57
		<i>Prosopis velutina</i>	1.99
		<i>Quercus emoryi</i>	1.27
	Tree	<i>Quercus oblongifolia</i>	0.36
		<i>Quercus emoryi</i>	1.92
		<i>Quercus oblongifolia</i>	0.10
	Cavity	<i>Quercus emoryi</i>	1.92
		<i>Quercus oblongifolia</i>	0.10
5	Subshrub	<i>Agave palmeri</i>	122.93
		<i>Baccharis pteronioides</i>	49.17
		<i>Acacia angustissima</i>	12.29
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	36.88
		<i>Quercus emoryi</i>	24.59
	Shrub	<i>Agave palmeri</i>	45.69
		<i>Baccharis sarothroides</i>	57.11
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	11.42

Transect			
Station	Category	Species	Mean density
<b>Wash</b>			
5	Tree	<i>Prosopis velutina</i>	2.05
		<i>Quercus emoryi</i>	5.33
		<i>Quercus oblongifolia</i>	0.82
	Cavity	<i>Quercus emoryi</i>	2.70
		<i>Quercus oblongifolia</i>	0.90
6	Subshrub	<i>Agave palmeri</i>	5.53
		<i>Baccharis pteronioides</i>	13.83
		<i>Baccharis sarothroides</i>	11.06
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	22.12
		<i>Prosopis velutina</i>	2.77
	Shrub	<i>Agave palmeri</i>	2.43
		<i>Baccharis sarothroides</i>	26.74
		<i>Opuntia spinosior</i>	2.43
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	7.29
		<i>Prosopis velutina</i>	7.29
	Tree	<i>Dasyllirion wheeleri</i>	2.43
		<i>Prosopis velutina</i>	2.05
		<i>Quercus emoryi</i>	5.33
		<i>Quercus oblongifolia</i>	0.82
	Cavity	<i>Quercus emoryi</i>	2.29
		<i>Quercus oblongifolia</i>	0.27
	Subshrub	<i>Agave palmeri</i>	2.99
		<i>Baccharis sarothroides</i>	3.99
		<i>Opuntia spinosior</i>	2.99
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	9.97

Transect			
Station	Category	Species	Mean density
	Shrub	<i>Baccharis sarothroides</i>	4.11
		<i>Opuntia spinosior</i>	6.85
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	8.22
		<i>Prosopis velutina</i>	1.37
		<i>Quercus emoryi</i>	6.85
	Tree	<i>Prosopis velutina</i>	1.73
		<i>Quercus emoryi</i>	14.69
		<i>Quercus oblongifolia</i>	0.86
	Cavity	<i>Quercus emoryi</i>	11.28
		<i>Quercus oblongifolia</i>	1.25
8	Subshrub	<i>Agave palmeri</i>	4.00
		<i>Opuntia spinosior</i>	2.00
		<i>Arctostaphylos pungens</i>	8.00
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	16.00
		<i>Prosopis velutina</i>	4.00
	Shrub	<i>Quercus emoryi</i>	6.00
		<i>Arctostaphylos pungens</i>	39.78
		<i>Mimosa aculeaticarpa</i> var. <i>biuncifera</i>	14.46
		<i>Prosopis velutina</i>	7.23
		<i>Quercus emoryi</i>	10.85
	Tree	<i>Rhus choriophylla</i>	2.97
		<i>Prosopis velutina</i>	5.93
		<i>Quercus emoryi</i>	41.53
	Cavity	<i>Quercus oblongifolia</i>	8.90
		<i>Quercus emoryi</i>	12.98
		<i>Quercus oblongifolia</i>	3.25

**Appendix K. Total number of observations and mean relative abundance (RA) from reconnaissance VCP surveys, Coronado NM, 2002–2004.** Observations included all observations at the stations along the transect; while the mean relative abundance estimates exclude flyovers and birds observed >75 m from stations. Sample sizes (*n*) are the number of stations surveyed. See Appendix C for scientific names.

Species	Transect-High elevation area						
	Ridge		Coronado Peak		Crest Trail		
	2004 ( <i>n</i> = 7)		2002 ( <i>n</i> = 3)		2003 ( <i>n</i> = 5)		2004 ( <i>n</i> = 5)
	Total obs.	RA	Total obs.	RA	Total obs.	RA	RA
turkey vulture					5		
American kestrel					1		
white-winged dove			1				
mourning dove	2		1	0.33	3	0.20	
greater roadrunner	1		2		1		
white-throated swift					3		
ladder-backed woodpecker	1	0.14			1	0.20	
western wood-pewee	3	0.29			1		0.20
Say's phoebe	1	0.14			1		0.20
dusky-capped flycatcher			1	0.33			
ash-throated flycatcher	1		1		2		0.20
Cassin's kingbird	2		1	0.33			
Hutton's vireo					1		
curve-billed thrasher	4	0.29			2		
crissal thrasher					1		
western scrub-jay	4	0.57	1		9	1.00	0.80
Mexican jay	4	0.14					
violet-green swallow	15		3		6		
bushtit					5	0.20	0.80
cactus wren	2	0.14	3	1.00	3	0.20	0.20
rock wren	1		1				
canyon wren	2	0.14					
Bewick's wren	3	0.29	1	0.33	6	0.60	
blue-gray gnatcatcher					1		
hepatic tanager	2	0.29	1	0.33			
western tanager					2		0.40
spotted towhee	3	0.43	2		7	0.80	0.20
canyon towhee	7	0.57			2		0.20
rufous-crowned sparrow	11	1.14	1	0.33	4	0.40	0.40
brown-headed cowbird	1	0.14					
Scott's oriole	8	0.57	2		1		
house finch	1	0.14	1		2		

**Appendix K. cont.**

Species	Transect-Grassland area					
	Level		Ranch		Uplands	
	2004 (n = 5)		2004 (n = 5)		2004 (n = 5)	
	Total obs.	RA	Total obs.	RA	Total obs.	RA
turkey vulture	4		8		1	
prairie falcon	1					
white-winged dove			1	0.20	1	
mourning dove			2	0.40	1	
greater roadrunner	1					
white-throated swift	6				3	
black-chinned hummingbird					2	0.40
ladder-backed woodpecker					1	0.20
Say's phoebe			2	0.40		
ash-throated flycatcher	1	0.20	2	0.20	1	0.20
Cassin's kingbird					2	
western kingbird			5	0.80	1	0.20
crissal thrasher	2	0.20				
Mexican jay	8	0.80	10	1.60	4	
common raven	1					
bridled titmouse	5	0.40			3	0.20
verdin	1		3	0.60		
bushtit	39	5.00	7	1.40		
white-breasted nuthatch	1	0.20				
cactus wren			2	0.40		
Bewick's wren	10	1.80	2	0.20	8	1.20
blue-gray gnatcatcher	2	0.40				
northern mockingbird					2	
Lucy's warbler			1	0.20		
canyon towhee			2	0.20	5	0.20
rufous-crowned sparrow			2		5	0.40
lark sparrow	1	0.20			1	
black-throated sparrow	1	0.20			1	0.20
black-headed grosbeak					1	
blue grosbeak			2	0.40		
eastern meadowlark			5	0.60		
brown-headed cowbird	2	0.20	1	0.20		
hooded oriole			1	0.20		
Scott's oriole	4	0.40			4	0.20
house finch	1		9	0.80		

**Appendix L. Total number of observations (sum) and mean relative abundance (RA) of birds observed during the non-breeding season (August–March) by Coronado NM volunteers, 2002–2004.** Relative abundance estimates include flyovers and birds observed >100 m from stations. Sample sizes (*n*) are the number of stations surveyed. See Appendix C for scientific names.

Species	Ranch ( <i>n</i> = 25)		Grassland ( <i>n</i> = 38)	
	Sum	RA	Sum	RA
Montezuma quail			2	0.05
turkey vulture	1	0.04		
northern harrier			3	0.08
red-tailed hawk			4	0.11
American kestrel			1	0.03
prairie falcon			1	0.03
mourning dove	8	0.32	1	0.03
ladder-backed woodpecker			6	0.16
northern flicker	29	1.16	2	0.05
Say's phoebe	1	0.04	1	0.03
western kingbird	4	0.16		
loggerhead shrike	3	0.12	3	0.08
Steller's jay			1	0.03
curve-billed thrasher	1	0.04	1	0.03
Mexican jay	6	0.24	4	0.11
pinyon Jay			1	0.03
common raven	1	0.04	9	0.24
verdin	5	0.20	1	0.03
bushtit			10	0.26
cactus wren	2	0.08	3	0.08
rock wren			2	0.05
canyon wren				
Bewick's wren	4	0.16	2	0.05
ruby-crowned kinglet	1	0.04		
western bluebird	1	0.04		
mountain bluebird			2	0.05
American robin	1	0.04		
phainopepla	3	0.12		
yellow-rumped warbler			1	0.03
green-tailed towhee	1	0.04	1	0.03
canyon towhee	11	0.44	3	0.08
rufous-crowned sparrow	2	0.08	33	0.87
chipping sparrow	9	0.36	8	0.21
Brewer's sparrow	5	0.20		
vesper sparrow	9	0.36	42	1.11
lark sparrow	1	0.04		
black-throated sparrow			5	0.13
white-crowned sparrow	1	0.04		
pyrrhuloxia	4	0.16		
eastern meadowlark			50	1.32
western meadowlark	1	0.04		
house finch	17	0.68	1	0.03



**Appendix M. Total number of observations (sum) and mean relative abundance (RA) of birds observed during the breeding season (April–July) by Coronado NM volunteers, 2002–2004.**

Relative abundance estimates include flyovers and birds observed >75 m from stations. Sample sizes (*n*) are the number of stations surveyed. See Appendix C for scientific names.

Species	Transect					
	Ranch ( <i>n</i> = 50)		Upland ( <i>n</i> = 13)		Grassland ( <i>n</i> = 10)	
	Sum	RA	Sum	RA	Sum	RA
turkey vulture	14	0.28			6	0.60
northern harrier	1	0.02			1	0.10
sharp-shinned hawk	1	0.02				
Cooper's hawk	1	0.02			1	0.10
red-tailed hawk	3	0.06				
prairie falcon					1	0.10
white-winged dove	4	0.08	1	0.08		
mourning dove	43	0.86	5	0.38	11	1.10
greater roadrunner	1	0.02				
common nighthawk	4	0.08				
white-throated swift	5	0.10				
black-chinned hummingbird	1	0.02				
broad-tailed hummingbird					3	0.30
Gila woodpecker			1	0.08		
ladder-backed woodpecker	3	0.06	1	0.08		
Arizona woodpecker	2	0.04				
northern flicker	2	0.04			1	0.10
greater pewee	1	0.02				
western wood-pewee	1	0.02				
Say's phoebe	2	0.04			1	0.10
ash-throated flycatcher	13	0.26	3	0.23	2	0.20
brown-crested flycatcher	1	0.02				
Cassin's kingbird	6	0.12	10	0.77		
loggerhead shrike	5	0.10				
Steller's jay	1	0.02				
curve-billed thrasher	3	0.06				
crissal thrasher			1	0.08		
Mexican jay	22	0.44	21	1.62	3	0.30
common raven	1	0.02			1	0.10
violet-green swallow	5	0.10				
cactus wren	9	0.18	3	0.23	2	0.20
canyon wren	1	0.02	2	0.15		
Bewick's wren	4	0.08	8	0.62	2	0.20
ruby-crowned kinglet	1	0.02			2	0.20
Virginia's warbler	1	0.02				
Wilson's warbler	1	0.02			1	0.10
green-tailed towhee	1	0.02			1	0.10
spotted towhee	3	0.06				
canyon towhee	13	0.26			3	0.30
Cassin's sparrow	1	0.02				

Species	Transect					
	Ranch ( <i>n</i> = 50)		Upland ( <i>n</i> = 13)		Grassland ( <i>n</i> = 10)	
	Sum	RA	Sum	RA	Sum	RA
rufous-crowned sparrow	9	0.18	5	0.38		
chipping sparrow	6	0.12	44	3.38	6	0.60
vesper sparrow	12	0.24			3	0.30
black-throated sparrow	4	0.08	2	0.15	2	0.20
song sparrow	1	0.02				
dark-eyed junco	1	0.02				
eastern meadowlark	12	0.24	2	0.15	2	0.20
brown-headed cowbird	1	0.02				
Scott's oriole	3	0.06	3	0.23		
house finch	27	0.54			7	0.70
lesser goldfinch	1	0.02				

**Appendix N. Most common species at each transect and season based on data published in Russell and Danforth (1979) and mean relative abundance (RA) data from Tables 5.3 and 5.4.** Relative abundance data from Russell and Danforth was the number of individuals per transect km. We only summarized data for species that had an average of >1 and >0.6 individuals per transect km for spring and winter surveys, respectively. For Russell and Danforth's Riparian transect, we averaged number of individuals per kilometer of transect for "Lower Wash" and "Woodland" because the two transects run parallel to the riparian area that we surveyed. Species in bold are those that are not found on the corresponding list of the most common species for that area. Relative abundance scores cannot be directly compared because of different methods of data collection and analysis.

Transect	Season	Study			
		Russell and Danforth (1977–1978)		UA inventory (2003–2004)	
		Species	RA	Species	RA
Riparian	Breeding	Mexican jay	6.2	Bewick's wren	1.26
		Bewick's wren	4.0	Mexican jay	0.88
		bridled titmouse	3.5	dusky-capped flycatcher	0.45
		black-throated gray warbler	2.6	black-throated gray warbler	0.45
		bushtit	2.6	bridled titmouse	0.42
		<b>chipping sparrow</b>	2.4	<b>spotted towhee</b>	0.38
		rufous-crowned sparrow	2.1	Scott's oriole	0.35
		ruby-crowned kinglet	2.0	rufous-crowned sparrow	0.27
		ash-throated flycatcher	1.5	<b>Hutton's vireo</b>	0.25
		Scott's oriole	1.5	ash-throated flycatcher	0.23
		<b>house finch</b>	1.3	bushtit	0.22
		dusky-capped flycatcher	1.3	brown-headed cowbird	0.18
		brown-headed cowbird	1.2	<b>black-headed grosbeak</b>	0.15
		<b>black-chinned hummingbird</b>	0.6	<b>northern flicker</b>	0.15
		<b>acorn woodpecker</b>	0.6	<b>sulphur-bellied flycatcher</b>	0.13
		<b>Hammond's flycatcher</b>	0.6	ruby-crowned kinglet	0.12
	Non-breeding	chipping sparrow	17.8	Mexican jay	2.33
		Mexican jay	4.5	bushtit	1.33
		Bewick's wren	3.7	ruby-crowned kinglet	0.89
		dark-eyed junco	3.5	spotted towhee	0.78
		bushtit	3.5	chipping sparrow	0.72
		ruby-crowned kinglet	3.4	<b>western bluebird</b>	0.67
		bridled titmouse	2.6	dark-eyed junco	0.67
		<b>eastern bluebird</b>	2.5	Bewick's wren	0.61
		<b>cedar waxwing</b>	0.9	bridled titmouse	0.39
		<b>lesser goldfinch</b>	0.5	<b>northern flicker</b>	0.17
Wash	Breeding	rufous-crowned sparrow	0.5	<b>white-breasted nuthatch</b>	0.17
		hermit thrush	0.4	hermit thrush	0.17
		Hutton's vireo	0.4	Hutton's vireo	0.11
		spotted towhee	0.4	rufous-crowned sparrow	0.11
	Non-breeding	<b>chipping sparrow</b>	5.6	Bewick's wren	0.82
		Mexican jay	5.1	ash-throated flycatcher	0.48
		<b>bushtit</b>	3.6	<b>Botteri's sparrow</b>	0.44
		<b>western tanager</b>	3.3	Mexican jay	0.39
		ash-throated flycatcher	2.1	<b>rufous-crowned sparrow</b>	0.28
		canyon towhee	1.9	<b>verdin</b>	0.26
	Breeding	Bewick's wren	1.8	canyon towhee	0.24
		mourning dove	1.6	mourning dove	0.24
	Non-breeding	<b>chipping sparrow</b>	75.6	bushtit	2.42
		<b>vesper sparrow</b>	5.4	ruby-crowned kinglet	1.17

Transect	Season	Study			
		Russell and Danforth (1977–1978)		UA inventory (2003–2004)	
		Species	RA	Species	RA
		bushtit	5.0	<b>verdin</b>	0.42
		<b>canyon towhee</b>	3.8	Mexican jay	0.33
		<b>Cassin's sparrow</b>	2.8	Bewick's wren	0.25
		<b>Mexican jay</b>	2.0	<b>rufous-crowned sparrow</b>	0.25
		ruby-crowned kinglet	1.4	<b>northern flicker</b>	0.17
		Bewick's wren	1.1	<b>white-breasted nuthatch</b>	0.17
		<b>house finch</b>	1.0	<b>common raven</b>	0.17
		<b>pyrrhuloxia</b>	0.8	<b>Say's phoebe</b>	0.17

**Appendix O. Species of terrestrial mammals (exclusive of nocturnal rodents) confirmed at Coronado NM and number of documented observations, including photographs by infrared-triggered photography, approximate number of observations during road transects and time-area constrained search, and number of observations recorded in CORO sightings database, 1972–1997.**

Species	Number infrared-triggered photographs	Number observations	Number observations, CORO database
desert shrew	0	7 <sup>a</sup>	0
black bear	27	0	11
common raccoon	4	0	2
white-nosed coati	12	2	357
ringtail	93	0	11
American badger	0	0	1
western spotted skunk	12	1	0
striped skunk	36	4	0
hooded skunk	11	0	3
common hog-nosed skunk	15	1	1
feral dog	1	0	1
coyote	2	3	43
common gray fox	50	12	57
feral cat	0	1	0
mountain lion	5	1	15
bobcat	2	1	20
rock squirrel	8	38	17
Arizona gray squirrel	0	0	10
Botta's pocket gopher	0	0	0
southern pocket gopher	0	0	0
black-tailed jackrabbit	0	4	43
eastern cottontail	2	0 <sup>a</sup>	0
desert cottontail	1	81 <sup>b</sup>	25
collared peccary	33	2	84
mule deer	0	0	5
white-tailed deer	0	157	1187

<sup>a</sup> Includes sightings by staff.

<sup>b</sup> Some sightings listed as desert cottontail may be eastern cottontail.

**Appendix P. Total number of animals captured during trapping on the Joe's Canyon grid at Coronado National Memorial, November 1997–2003.** Species richness is the number of species trapped in a year; accumulated species is the total number of species trapped since 1997.

Species	1997	1998	1999	2000	2001	2002	2003
desert shrew			1				
spotted ground squirrel							
silky pocket mouse							
desert pocket mouse							
rock pocket mouse	3	4	3		8	5	8
western harvest mouse	1						
fulvous harvest mouse	8		4	10	13	2	11
deer mouse	1			2			
white-footed mouse	9	4	25	5	3	2	3
unknown white-footed mouse	1		2	2			3
brush mouse		6	2	14	23	4	25
pygmy mouse	4	2	3	5	11	1	1
white-throated woodrat	8	9	13	10	12	8	20
yellow-nosed cotton rat	17		3	2	6	8	14
Arizona cotton rat		11	2	3	11		
unknown cotton rat							1
<b>Total</b>	<b>52</b>	<b>36</b>	<b>59</b>	<b>53</b>	<b>88</b>	<b>30</b>	<b>87</b>
<b>Species Richness</b>	<b>8</b>	<b>6</b>	<b>9</b>	<b>8</b>	<b>9</b>	<b>7</b>	<b>7</b>
<b>Accumulated Species</b>	<b>8</b>	<b>10</b>	<b>11</b>	<b>11</b>	<b>12</b>	<b>12</b>	<b>12</b>

**Appendix Q. Total number of animals captured during trapping at the Grassland grid, Coronado National Memorial, November 1997–2003.** Species richness is the number of species trapped in a year; accumulated species is the total number of species trapped since 1997.

Species	1997	1998	1999	2000	2001	2002	2003
spotted ground squirrel				1			
silky pocket mouse				1	2	1	
desert pocket mouse						4	1
hispid pocket mouse	1		4	1	4	2	1
Ord's kangaroo rat							9
western harvest mouse	6	7	1	4	10	8	11
fulvous harvest mouse	6	3	8	3	2	8	3
unknown harvest mouse		2		2			
deer mouse				1	3	3	13
unknown deer mouse							1
pygmy mouse	3	3	11	21	19	17	22
southern grasshopper mouse	4	10	10	22	14	6	9
Arizona cotton rat	4	2	8	38	58	10	8
tawny-bellied cotton rat				1	5	2	1
yellow-nosed cotton rat	3	4		3		7	
unknown cotton rat		1	3	2	1		
house mouse					1		
<b>Total</b>	<b>27</b>	<b>32</b>	<b>45</b>	<b>100</b>	<b>119</b>	<b>68</b>	<b>79</b>
<b>Species Richness</b>	<b>7</b>	<b>6</b>	<b>6</b>	<b>11</b>	<b>10</b>	<b>11</b>	<b>10</b>
<b>Accumulated Species</b>	<b>7</b>	<b>7</b>	<b>7</b>	<b>11</b>	<b>12</b>	<b>13</b>	<b>14</b>

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